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General fungi ITS2 (ITS4ngsUni - fITS7) for Illumina amplicon sequencing

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SOWA

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Protocol status: Working

We use this protocol and it's working

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Keywords: fungal communities by illumina amplicon, fungal community, illumina amplicon, its2 region primer pair, specific primer, rna, ribosomal rna gene, 2nd internal transcribed spacer region, pcr amplicons for further analysis, fits7 primer, primer, universal primer, pcr amplicon, amplicons in the range, amplicon, eukaryote

Abstract

A general assay for the preparation of PCR amplicons for further analysis of fungal communities by Illumina amplicon sequencing.

The primers target the 2nd internal transcribed spacer region (ITS2) of the ribosomal RNA gene and were specifically designed for Illumina amplicon sequencing. The ITS4ngsUni primer was designed by Tedersoo et al. (2016) as a universal primer for nearly all eukaryotes, and the fITS7 primer was designed by Ihrmark et al. (2012) as a fungal-specific primer. The ITS2 region primer pair should capture a higher diversity compared to primers targeting the ITS1 region.

For barcoding, we use the [Fludigm Access Array](#), and therefore, the primers are synthesized with the CS1 and CS2 regions.

When working with soil samples, we usually obtain amplicons in the range of 300-500bp. Double bands are often observed.

Protocol materials

✕ Agarose Merck MilliporeSigma (Sigma-Aldrich) Catalog #A9539

✕ GeneRuler DNA Ladder Mix Thermo Fisher Scientific Catalog #SM0331

✕ DreamTaq DNA Polymerase (5 U/μL) Thermo Fisher Catalog #EP0701

✕ 0.2 mM dNTPs Thermo Fisher Scientific Catalog #AM8200

✕ Bovine Serum Albumin (BSA) Merck MilliporeSigma (Sigma-Aldrich) Catalog #A7906

Primers

1

A	B	C	D
Name	Sequence	Ref.	Target region 1
ITS4ngsUni_CS1	ACA CTG ACG ACA TGG TTC TAC ACG CCT SCS CTT ANT DAT ATG C	Tedersoo et al. (2016)	5.8S
fITS7_CS2	TAC GGT AGC AGA GAC TTG GTC TGG GTG ART CAT CGA ATC TTT G	Ihrmark et al. (2012)	ITS-flanking site in LSU

¹ within the ribosomal RNA gene

PCR reaction

2 Prepare the following master mixture  On ice .

Do not forget to prepare some additional mixture for the negative (NTC = no template) and positive controls, and to account for pipetting errors. Work in a clean PCR box.

A	B	C	D
Reagent	Final. conc.	1 tube(25µl) µl	96 tubes (25µl x100)
PCR H ₂ O		16,775	1677,5
10X DreamTaq Green Buffer	1X	2,5 ul	250
dNTP (2 mM each)	0.2 mM	2,5	250
BSA (20 µg µl ⁻¹)	80 ng µl ⁻¹	0,1	10
ITS4ngsUni-CS1 linker	0.5 µM	1	100
fITS7-CS2 linker	0.5 µM	1	100
DreamTaq Green DNA Polymerase	0.625 U	0,125	12,5
Final volume		25	2500



DreamTaq DNA Polymerase (5 U/μL) **Thermo Fisher Catalog #EP0701**

0.2 mM dNTPs **Thermo Fisher Scientific Catalog #AM8200**

Bovine Serum Albumin (BSA) **Merck MilliporeSigma (Sigma-Aldrich) Catalog #A7906**

3 Vortex and spin down 00:02:00 .

2m

4 Distribute 24 μL of the mixture to each tube or well of 96-well plate and add 1 μL of template DNA or cDNA.

PCR program

5 Run the following PCR program:

2h

1. 95 °C 00:10:00

2. x 30{

a. 95 °C 00:00:45

b. 52 °C 00:00:45

c. 72 °C 00:00:45

}

4. 72 °C 00:10:00

5. 10 °C 00:00:00 hold

Evaluate PCR products on an agarose gel

1h 10m

6 Prepare a 1.5% agarose gel by mixing:

1h

100 mL TAE

1.5 g agarose

Heat in the microwave until dissolved and pour into a gel frame.

Place solid gel into an electrophoresis bath filled with TAE buffer.

Agarose **Merck MilliporeSigma (Sigma-Aldrich) Catalog #A9539**

GeneRuler DNA Ladder Mix **Thermo Fisher Scientific Catalog #SM0331**



GeneRuler DNA Ladder Mix Thermo Fisher Scientific Catalog #SM0331

- 7 Load  5 μL of the sample into a well. 10m
In addition load  5 μL of DNA ladder mix (80–10,000 bp) into an empty well, as a marker.
 - 8 Run the gel at 110 V, 265 mA for approx.  00:40:00 40m
 - 9 Stain gel for at least  00:30:00 in an Ethidium bromide TAE bath (or any other DNA stain). 30m
- #### Safety information
- Ethidium bromide is not regulated as hazardous waste at low concentrations, but is treated as hazardous waste by many organizations. Material should be handled according to the manufacturer's **Safety Data Sheet** (SDS).
- Most use of ethidium bromide in the laboratory (0.25–1 $\mu\text{g}/\text{ml}$) is below the LD50 dosage, making acute toxicity unlikely. Testing in humans and longer studies in a mammalian system would be required to fully understand the long-term risk ethidium bromide poses to lab workers, but it is clear that ethidium bromide can cause mutations in mammalian and bacterial cells.
- [Wikipedia](#)
- 10 Visualize the gel using a gel documentation system.

Protocol references

Tedersoo, L. *et al.* Tree diversity and species identity effects on soil fungi, protists and animals are context dependent. *ISME J.* **10**, 346–362 (2016).

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