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Gel Electrophoresis

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Protocol status: Working

We use this protocol and it's working

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Abstract

How to prepare and run gel electrophoresis using agarose gel for the verification of nucleic acid amplification products.

Guidelines

Gloves should be worn at all times.

Materials

- 50x TAE Buffer
- Agarose
- DNA ladder
- Loading dye (optional)
- Nucleic acid amplification solution (e.g. PCR or LAMP)
- Ethidium bromide
- Deionized water



Troubleshooting

Safety warnings


- ❗ Gel bottle will be hot from microwave, use a hot mitt and be cautious to not get hot gel on your skin. Ethidium bromide may be a mutagen; skin contact should be avoided.



Make TAE buffer

- 1 In a large glass bottle, add  750 mL DI water.
- 2 Add  15 mL 50x TAE buffer to the deionized water.
- 3 Cap the bottle and mix by swirling



Make gel

- 4 Pour  300 mL TAE buffer into microwavable glass bottle.
- 5 Add the following amount of agarose powder to TAE buffer for different gel concentrations:

	Concentration (w/v)	Agarose (g)
	2.0%	6
	2.5%	7.5
	3%	9

Note



Swirl solution lightly to mix agarose into buffer. Swirling too vigorously will introduce bubbles that will make casting difficult.

- 6 Microwave in  00:00:40 intervals until completely dissolved, swirling bottle between heating intervals.
- 7 Pour into gel mold with well insert. Add to black line (~  100 mL)






- 8 Allow gel to set for approximately  00:20:00 .

Load and run gel


- 9 Transfer gel boat with gel into electrophoresis chamber.
- 10 Add buffer to line marked on chamber, completely covering the gel.
- 11 Carefully load the wells with ladder (green) and amplification product solutions at volumes between  1 μL and  5 μL .

Note

Optionally, you can mix  1 μL loading dye with  4 μL amplification product solution in a separate microcentrifuge tube to make the loading process easier to visualize.

- 12 Put the lid on the chamber, turn on the power supply and press 'RUN'. Run for  00:50:00 .

Note

The power supply does not turn off automatically, so set a timer and shut off after  00:50:00 . This time may change based on factors such as gel concentration and amplicon length.

Stain the gel

- 13 Transfer the gel into the dying tray.





- 14 Check buffer level and add more if necessary.
- 15 Add  8 μL ethidium bromide to dying tray.
- 16 Turn on stage, and allow to stain for  00:20:00 .

Image gel

- 17 Place gel in imaging machine.
- 18 Turn on the machine by the switch on left side of machine. Press 'Trans UV'.
- 19 Take a picture with the camera.