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Version 2

Fluorescence assay for Enterovirus coxsackievirus A16 2A protease activity measurement V.2

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Protocol status: Working

We use this protocol and it's working

Created: February 13, 2025

Last Modified: May 21, 2025

Protocol Integer ID: 120175

Keywords: FRET assay, enzymatic assay, Enterovirus coxsackievirus A16, Picornaviridae , IC50 measurement, activity of viral protease, enterovirus coxsackievirus a16, viral protease, 2a protease activity measurement, sequence specific to the tested protease, cleavage of the peptide, protease activity measurement, detection of the fluorescence, tested protease, protein production protocol, fluorescence emission, fluorescence, protein production, protease, peptide, labelled peptide, assay, proximity of dabcyI

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Disclaimer

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Abstract

This protocol details the fluorescence assay for Enterovirus coxsackievirus A16 (CVA16) 2A protease activity measurement. This method is intended to measure the activity of viral proteases by using a specific labelled-peptide that allows the detection of the cleaved product. The substrate contains the cleavage-sequence specific to the tested protease and is labeled in C-terminal by the fluorophore Edans (ex 336 nm; em: 455 nm) and in N-terminal by the quencher DabcyI (ex 472 nm). In the case of a non-cleaved substrate, the proximity of DabcyI to Edans prevents the emission and the detection of the fluorescence at 455 nm. The cleavage of the peptide by the protease allows Edans' fluorescence emission and detection. This version we added the protein production protocol and the addgene ID.

Attachments



[nyiwb5id7.docx](#)

203KB



Materials

Reagents

- **Assay buffer:** 25 millimolar (mM) HEPES pH 7.5, 150 millimolar (mM) NaCl, 10 millimolar (mM) glycerol, 7 millimolar (mM) DTT, 5 millimolar (mM) EDTA, 1 millimolar (mM) TCEP (optional) and 0.05 % volume Tween-20 .
- **Incubation:** 01:00:00 at Room temperature .
- **EV-A16 2A:** Protein stocks were stored at -80 °C and used as 2x solution (5 micromolar (μM) , 2.5 micromolar (μM) final assay concentration) in assay buffer.
- **Substrate:** Dabcyl-KEALFQGPPQFE-Edans (LifeTein, USA) prepared as a stock solution at 10 millimolar (mM) in DMSO and used at 2x solution (20 micromolar (μM) , 10 micromolar (μM) final concentration assay concentration) in assay buffer.
- **Positive control:** Telaprevir (Pubchem CID 3010818), 25 micromolar (μM) top final assay concentration.
- **Plates:** Greiner, 384-well plate, black, small volume, non-binding #7840900
- **Liquid handler:** Echo® acoustic liquid handler (Beckman Coulter, USA).
- **Plate reader:** Pherastar FS, BMG Labtech (Germany), 350-460 FI optic module, the plate is read every 30 s for 2 hours and shaken during 5 s before the first reading.

Protocol materials

 Enterovirus Coxsackievirus A16 2A protease **addgene Catalog #228632**



Protein expression and purification

- 1 The protein used in this assay was expressed and purified using the protocol bellow with the addgene.

 Enterovirus Cocksackievirus A16 2A protease **addgene** Catalog #228632

Protocol

NAME

Enterovirus coxsackievirus A16 2A protease small scale expression and purification protocol

CREATED BY

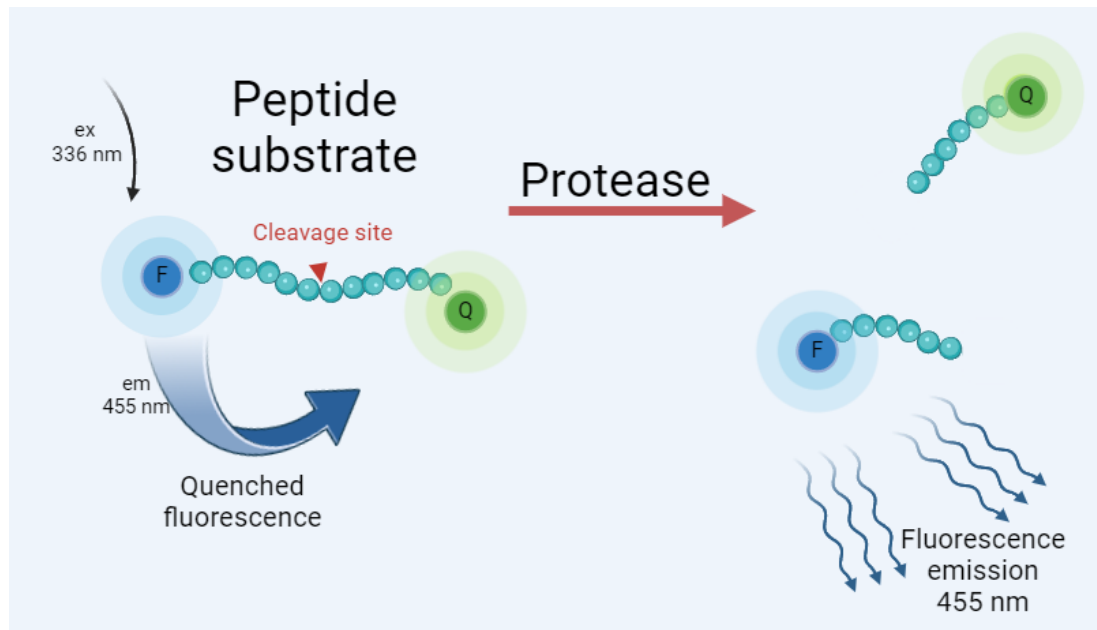
korvus.wang

Preview

CVA16 2A protease Assay

3h

- 2 This method is intended to measure the activity of viral proteases by using a specific labelled-peptide that allows the detection of the cleaved product. The substrate contains the cleavage-sequence specific to the tested protease and is labeled in C-terminal by the fluorophore Edans (ex 336 nm; em: 455 nm) and in N-terminal by the quencher Dabcyl (ex 472 nm). In the case of a non-cleaved substrate, the proximity of Dabcyl to Edans prevents the emission and the detection of the fluorescence at 455 nm. The cleavage of the peptide by the protease allows Edans' fluorescence emission and detection.



Asset URL:

Reagents and equipment

3 **Assay buffer:** [M] 25 millimolar (mM) HEPES pH 7.5, [M] 150 millimolar (mM) NaCl, [M] 10 millimolar (mM) glycerol, [M] 7 millimolar (mM) DTT, [M] 5 millimolar (mM) EDTA, [M] 1 millimolar (mM) TCEP (optional) and [M] 0.05 % volume Tween-20 .

3h 0m 35s

Incubation: ⌚ 01:00:00 at room temperature.

CVA16 2A protease: protein stocks were stored at -80C and used as [M] 2 x solution ([M] 10 micromolar (μM) , [M] 5 micromolar (μM) final assay concentration) in assay buffer.

Positive control: Telaprevir (Pubchem CID 3010818), [M] 25 micromolar (μM) top final assay concentration.

Substrate: Dabcyl-RDKITTLGKFGQDE-Edans (LifeTein, USA) prepared as a stock solution at 10 mM in DMSO and used at 2x solution [M] 20 micromolar (μM) ([M] 10 micromolar (μM) final concentration assay concentration) in assay buffer.

Plates: Greiner, 384-well plate, black, small volume, non-binding #7840900

Liquid handler: Echo® acoustic liquid handler (Beckman Coulter, USA).

Plate reader: Pherastar FS, BMG Labtech (Germany), 350-460 FI optic module, the plate is read every ⌚ 00:00:30 for ⌚ 02:00:00 and shaken ⌚ 00:00:05 before each reading.



CVA16 2A protease IC50 Measurement

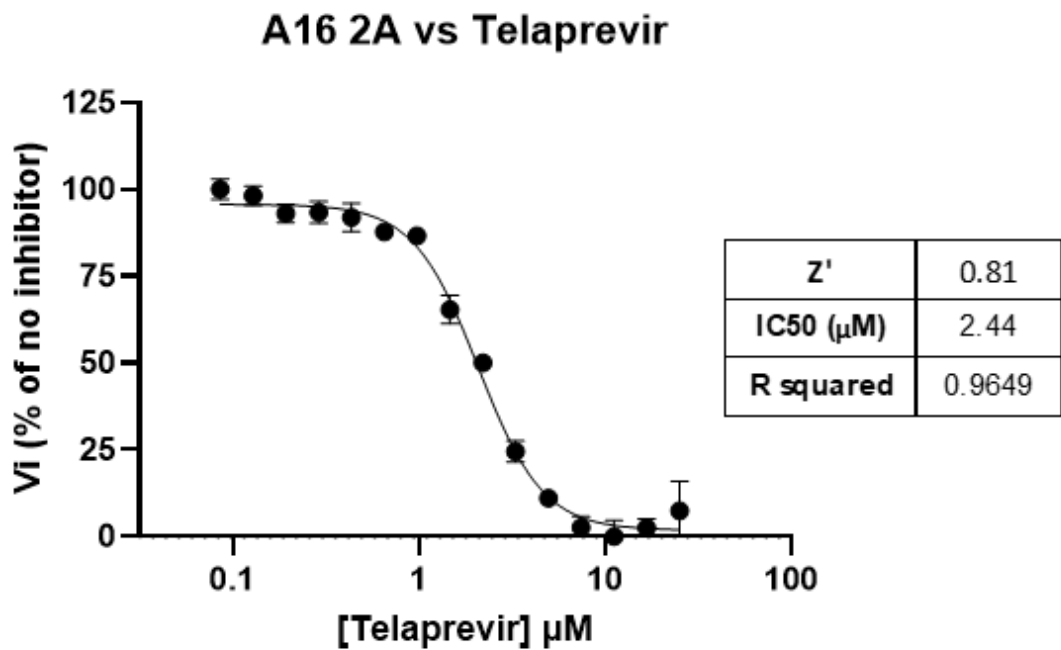
3h

- 4 Add  10 μL of 2x protein [M] 10 micromolar (μM) solution to each well containing the compounds to be tested previously dispensed onto the plate. 
- 5 Incubate the mix for  01:00:00 at  Room temperature and initiate the enzymatic reaction by the addition of  10 μL of 2x ([M] 20 micromolar (μM)) substrate solution using the plate reader injector.  1h
- 6 Read the fluorescence intensity at 350/460 nm every  00:00:30 for  02:00:00 2h 0m 30s
in kinetic mode, which includes a shaking step of the plate before each measurement.
- 7 Calculate the IC50 by plotting the initial velocity against various concentrations of tested inhibitor by using a four-parameter dose-response curve in Prism (v8.0) software.
- 8 Calculate the mean (μ) and the standard deviation (σ) of fluorescence intensity and then calculate the signal-to-background ratio and the Z' or Z-factor. (s: signal; c: control).

$$\text{Estimated Z-factor} = 1 - \frac{3(\hat{\sigma}_s + \hat{\sigma}_c)}{|\hat{\mu}_s - \hat{\mu}_c|}$$

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9



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