

Jul 14, 2019

Flow Cytometry-Based Gamma-H2AX Assay for Radiation Biodosimetry

DOI

dx.doi.org/10.17504/protocols.io.5gzg3x6

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Protocol Citation: Ghazi Alsbeih, Krishna Mishra, Maha Alrashd, Subramanian M. Pulicat, Najla Al-Harbi, Sara Bin Judia, Belal Moftah 2019. Flow Cytometry-Based Gamma-H2AX Assay for Radiation Biodosimetry. **protocols.io**
<https://dx.doi.org/10.17504/protocols.io.5gzg3x6>

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Protocol status: Working

We use this protocol and it's working

Created: July 14, 2019

Last Modified: July 14, 2019

Protocol Integer ID: 25849

A. Lymphocytes separation and fixation:

- 1 Collect blood samples in heparinized Vacutainers (BD, USA).
- 2 Irradiate 2 ml aliquot of whole blood with the required radiation dose.
- 3 Incubate for 30 minutes (or other time) at 37°C.
- 4 Take the 2 ml whole blood and dilute (1:1) with 2ml PBS in a 15 ml tube.
- 5 Carefully layer the mix on 2 ml of Histopaque (Ficoll).
- 6 Centrifuge at 1,200 g (Eppendorf 5810 R) for 15 min at RT.
- 7 Isolate middle cloudy lymphocytes layer and wash 2-3 times with cold PBS*.
- 8 Fix lymphocytes($1-5 \times 10^6$) with 2% paraformaldehyde (PFA) for 10-20 minutes on ice.
At this stage, fixed lymphocytes can either be processed further for immunostaining or stored overnight at +4°C.

B. γ H2AX immunostaining and flow cytometry measurement

- 9 Wash fixed lymphocytes twice with PBS*.
- 10 Incubate lymphocytes for 1 hour at 37°C with primary antibodies (anti-phospho-histone H2A.X, Ser139, clone JBW301; Millipore, CA, USA), in a 1:200 dilution in a buffer containing 1% bovine serum albumin (BSA) and 0.12 % TX-100.
- 11 Wash twice with PBS*.
- 12 Incubate with secondary antibodies (FITC-goat antimouse Ig BD Pharmingen, San Jose, CA, USA), in a 1:200 dilution in a buffer containing 1% bovine serum albumin (BSA) and 12



% TX-100, for 1 hour at 37°C.

13 Wash twice with PBS*.

14 Suspend lymphocytes in fresh 400-500 µl of PBS according to cells concentration.

15 Measure gamma-H2AX fluorescence in lymphocytes using flow cytometer (LSR II, Becton Dickinson, USA) and analyze the data using PC-based BD FACSDiva software. The following settings are used:

15.1 FITC laser excitation of 488 nm and emission of 533 nm.

15.2 A single population of lymphocytes is gated and used for mean fluorescence intensity (MFI) calculations.

15.3 Yields of γH2AX are calculated as the ratio of mean fluorescence intensity of irradiated versus control samples.

***Note: All PBS washing step include suspending cells in 500 µl PBS and centrifuging (Eppendorf 5810 R) at 100rpm for 10 minutes.**

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