Sep 09, 2020

FloodLAMP Inactivation Protocol v3.1

DOI

dx.doi.org/10.17504/protocols.io.bk5nky5e

Randy True¹

¹FloodLAMP Biotechnologies PBC

FloodLAMP XPRIZE Rapid Covid Tes...





DOI: dx.doi.org/10.17504/protocols.io.bk5nky5e

Protocol Citation: Randy True 2020. FloodLAMP Inactivation Protocol v3.1. protocols.io <u>https://dx.doi.org/10.17504/protocols.io.bk5nky5e</u>

Manuscript citation:

[1] Rabe B, Cepko C. SARS-CoV-2 Detection Using an Isothermal Amplification Reaction and a Rapid, Inexpensive Protocol for Sample Inactivation and Purification. medXriv preprint 4-28-20 https://www.medrxiv.org/content/10.1101/2020.04.23.20076877v1

License: This is an open access protocol distributed under the terms of the **<u>Creative Commons Attribution License</u>**, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

Protocol status: Working We use this protocol and it's working

Created: September 08, 2020

Last Modified: September 09, 2020

Protocol Integer ID: 41870

Abstract

The inactivation protocol uses a chemical plus heat to break open cells (including virus, if present) and preserve the RNA. The inactivation can be done outside a lab setting, at the point of sample collection or drop-off. This is our preferred mode, and we refer to these as "inactivation stations". Prior to inactivation, the samples may contain live, contagious virus, so it is crucial that personnel use proper PPE and safe handling procedures. Many current community screening efforts are performing non-lab based sample processing (for example, <u>Dave O'Connor</u> and <u>Chris Mason</u>), and we have followed their examples with the use of splash guards in addition to PPE.

After inactivation, samples should be refrigerated or stored on ice before same-day processing through the assay.

On our **website** are our protocols in worksheet form as we use in the lab. This and more information will be live soon.

Guidelines

Individuals are responsible for the chemical and bio safety training to safely complete this protocol. Be careful of aerating samples pre-inactivation. The inactivation solution should also be handled cautiously.

Materials

MATERIALS

Sodium hydroxide Sigma – Aldrich Catalog #S8045

🔀 UltraPure 0.5M EDTA, pH 8.0 Thermo Fisher Scientific Catalog #15575-038

X UltraPure Distilled Water Thermo Fisher Scientific Catalog #10977015

X Tris(2-carboxyethyl)phosphine hydrochloride (TCEP) Sigma Aldrich Catalog #C4706

Sigma 100% Ethanol Catalog #E7203

X Phosphate Buffered Saline **Thermofisher Catalog #**10010023

Tubes:

- 1.5mL Eppendorf DNA LoBind Tubes (\$39 for 250)
- 5mL Eppendorf LoBind with screw cap (\$79 for 200)
- 5mL colorless Eppendorf DNA tube with screw cap (\$65 for 200)
- 5mL amber Eppendorf DNA tube with screw cap (\$85 for 200)
- 5mL Eppendorf DNA tube with snap cap (\$58 for 200)
- 30mL Self Stading Tubes Chubs by Stellar Scientific (\$99 for 500)

Reagents:

- Zeptometrix

Safety warnings

• Both TCEP and EDTA should be handled cautiously as they can cause severe eye damage and are toxic if inhaled. See SDS for TCEP, EDTA, NaOH, HCI and NaI for more safety information.

Before start

- Set Up: turn on dry heater, get out fozen lab armor, turn on centrifuge, make temp QC tube for each size and volume of tube going through inactivation process

- Safety Procedures: always cover heater with plastic cover, wear appropriate PPE

Sample Preparation 2m		
1 For dry pasal swape add 2 Emt. 1xDPS, eask swape for 2 minutes, remove and dispard		
	swabs properly	2m
2	For positive controls, spike selected samples with inactivated virions	
Inactivation 20m		
3	Spray all closed tubes with 70% ethanol	
4	Vortex the 100x Inactivation Solution within five minutes of use	
5	Add 100x Inactivation Solution to collection tube, add 50uL per 5mL collected sample	2m
6	For each sample, vortex for 5 seconds, shake and then vortex for another 5 seconds	
7	Heat the samples at 95C for 8 minutes, be sure to add the QC tube(s) for in situ temperature check	8m
8	Chill samples using the frozen lab armor for 4 minutes, alteratively on ice	4
-		4m
9	Check caps are tight, then Centrifuge samples at 5K rpm for 4 minutes	4m
10	Aspirate the clarified samples into new tubes, be sure not to disturb any solids at the bottom of the original tube	2m