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Flame Photometry Protocol

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Mariam Awlia¹ ¹University of Cape Town

Salt Lab KAUST



Mariam Awlia University of Cape Town





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Protocol status: Working We use this protocol and it's working

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Flame Photometry

Attachments



Safety warnings

- Work in the fume hood.
 - Wear lab coat, eye protection, gloves and lower screen in front of face.
 - Always add <u>acid to water</u> slowly, never water to acid.
 - Please see SDS (Safety Data Sheet) for hazards and safety warnings.

Preparation of 1% nitric acid	
1	
	Safety information
	 Work in the fume hood. Wear lab coat, eye protection, gloves and lower screen in front of face. Always add <u>acid to water</u> slowly, never water to acid.
	Note
	For 250 samples (250*5=1.25L), 1.5L of 1% nitric acid should be sufficient.
	If stock is 70% nitric acid, pour 469 mL of MilliQ-water for every 41 mL of nitric acid. (x 21.4 for 1.5L).
	If stock is 69% nitric acid, pour 468 mL of MilliQ-water for every 41 mL of nitric acid. (x 21.8 for 1.5L)
	Pour 21.8 mL nitric acid to 1483.4 mL MilliQ-water.

Preparation of samples

- 2 Put all samples in <u>I 10 mL</u> falcon tubes and determine fresh/dry weight of plant material.
- 3 Add \underline{A} 3 mL of 1% nitric acid to \underline{A} 10 mL tubes of samples and close lids tightly.
- 4 Put in oven at **#** 70 °C overnight.
- 5 After 3:00:00, shake gently without breaking the rosette to cover all in nitric acid.
- 6 Remove samples and let them cool down on the bench before diluting.

Preparation of dilutions

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Note

- Prepare dilutions in fresh MilliQ-water and make sure to be extremely precise in all measurements.
- Prepare replicates for each sample, just in case.

Use 2 salt and 2 control Eppendorfs to test the dilutions and standards before doing the rest of the samples.

Flip the falcon tube to remove condensation, before pipetting.

Note

Suggested dilutions: Salt 1:20 (1.90ml $H_2O + 100\mu L$ sample)

Control 1:5 (1.6ml $H_2O + 400\mu L$ sample)

Suggested standards: Salt 500µM NaCl, 100µM KCl

Control 10-100µM NaCl, 1000µM KCl

Note

Make sure to avoid carrying over any plant-fibre to the dilutions.

Operation of flame photometer

- 8 Fill two beakers with fresh MilliQ-water; <u>the running solution</u> and <u>blank solution</u>.
- 9 Put the running solution under the U-tube and add water with droplet to the connecting tube.
- 10 Check that the U-tube doesn't have any air bubbles; wiggle it around if necessary.

- 11 Check waste container and dispose of solution down the sink.
- 12 Open "air" on the main valve and check that liquid is being drawn into the machine and that waste liquid is flowing out into waste bucket.
- 13 Check at the back of instrument that pressure is approx. 11 psi.
- 14 Open gas bottle by turning the valve all the way, then a half-turn back.
- 15 Switch flame photometer and check that the flame has been lit, and then close the hatch.
- 16 Turn computer on to record the measurements manually. Leave the running solution under the U-tube and let it run for 😢 00:30:00 to 🕑 01:00:00 before use.

Calibration

- 17 Shake the standard solution bottle before use, then add around $\boxed{10 \text{ mL}}$ to a beaker.
- 17.1 Set system to dual and peak only. Put in blank solution, press "Blank", wait till "100" is flashing, then remove blank solution.
- 17.2 Put in selected standard solution and press "Cal" for Na and K and when "100" flashes, press "Cal" again.
- 17.3 Press "Measure" and wait till it blinks to measure **Blank** (should be 0±2) and **Standard** (should be 100±2).
- 17.4 Start measuring Eppendorf samples and wipe the tube with tissue when changing solutions.

Note

Correct measurements for Salt (20 - 120) and Control (10 - 20)

17.5 Run standard as test every 12-24 Samples

Note

When switching between standard solutions, press "Blank" for 3 seconds to restart calibration.

Shut down and clean-up

- 18 Make sure the running solution is still under the U-tube, so the flame photometer doesn't dry out.
- 18.1 Turn gas valve off and wait until the error E61 is displayed and flame is out, then switch off and close air.Dispose of solution and plant material down sink with excess water.