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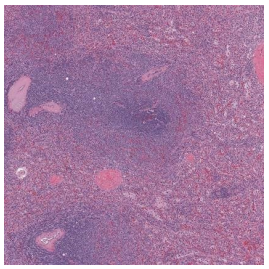
FFPE Tissue Sectioning for Staining V.1



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Protocol status: Working
We use this protocol and it's working

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Abstract

Method for sectioning paraffin blocks for staining

Guidelines

FFPE tissues sectioned onto poly-lysine-coated coverslips can be stored at 4° C for up to six months prior to staining. It is critical not to exceed a thickness of 10 µm because it can disrupt the autofocusing capabilities of the microscope. For best results, the tissue should be devoid of folds and tears. To ensure the integrity of the tissue slices, it is critical that they are not stacked on top of one another after they have been placed on the coverslips.

1 Prepare a water bath at 42°C and place it next to the Microtome.

2 Label positive charged microscopes slides.

3 Insert a new blade for sectioning each block

4 Section the tissue between 4-5 µm thick.

5 Place the sectioned tissue in the water bath for a few seconds and observe it expanding.

6 When the observed expansion is enough to remove folds and wrinkles from the tissue, quickly place positive charged microscope slide in the water bath and gently move it upward towards the tissue.

7 Doing so, the tissue will lay on the slide as this is moved out from the water bath.

8 Put the slides upright in a rack to let air dry overnight or dry in 50°C oven for 1 hour.