

Apr 25, 2024

Fecal Carmine Red Protocol

DOI

dx.doi.org/10.17504/protocols.io.eq2lywpwwvx9/v1

Adam Hamilton¹, Ian N Krout¹, Tim Sampson¹

¹Emory University



Ian N Krout

Emory University

Create & collaborate more with a free account

Edit and publish protocols, collaborate in communities, share insights through comments, and track progress with run records.

Create free account





DOI: https://dx.doi.org/10.17504/protocols.io.eq2lywpwwvx9/v1

Protocol Citation: Adam Hamilton, Ian N Krout, Tim Sampson 2024. Fecal Carmine Red Protocol. protocols.io https://dx.doi.org/10.17504/protocols.io.eq2lywpwwvx9/v1

License: This is an open access protocol distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

Protocol status: Working

We use this protocol and it's working

Created: April 24, 2024



Last Modified: May 31, 2024

Protocol Integer ID: 98671

Keywords: ASAPCRN, GI Transit, Gut, Motility, fecal carmine red protocol, bright red fecal pellet, large intestinal transit, whole gut transit time, bright red carmine dye, assay, volume of bright red carmine dye, pellet expulsion, time the dye, dye, gastric emptying, time between gavage, mice, gi tract, length of the gi tract

Funders Acknowledgements:

Aligning Science Across Parkinson's

Grant ID: ASAP020527

Abstract

This assay is used to determine whole gut transit time. Mice are given an oral gavage containing a known volume of bright red carmine dye. Mice are placed into empty cages and are then observed in 15-minute intervals until they produce a bright red fecal pellet. The time between gavage and pellet expulsion is the time the dye takes to travel down the length of the GI tract, which is influenced by gastric emptying, and small and large intestinal transit.

Materials

Carmine Red Solution

6% w/v carmine red (Sigma cat #C1022), 0.5% w/v methylcellulose (Sigma cat #M7027) in water.

Sterile Cages

Standard Mouse Chow (whatever the mice currently are eating)

Gavage needles

Troubleshooting



Pre-protocol

- 1 Prepare Carmine Red Soln. (see materials)
- 2 Autoclave Carmine Red Soln. if sterility is required. (i.e. microbiome analysis)
- 3 Shake well or stir on low heat to homogenize
- 4 Once resuspended, immediately aliquot into 4 1.5 mL tubes and store in fridge to prevent spoilage.
- 5 Allow aliquots to come to room temperature prior to gavage.
- 6 Shake/vortex each right before gavage.

Day of Set-up

1h



Day of Assay



Orally gavage mice with $\[\] \] 100 \ \mu L$ carmine red solution. Generally, soft-tipped, disposable (Instec FTP-20-30, or similar) feeding needles, attached to a $\[\] \] 1 \ m L$ sliptip syringe.



Equipment	
Polypropylene Feeding Tubes for Rodents	NAME
Gavage Needle	TYPE
Instech Labs	BRAND
FTP-20-30	SKU
https://www.instechlabs.com/products/feeding-tubes/polypropylene ^{LINK}	

- 9 Record time of gavage for each mouse.
- Following gavage, place animals back into home cage with cage mates, food, and water.
- Allow animals to rest with food, water, in home cage for 01:00:00 (wildtype SPF mice will start producing red pellets 02:00:00 after gavage, while germ-free mice often take longer than 00:00:00).
- •Split individual mice into single-housed, clean (or sterile, if needed) cages, with <u>no</u> <u>bedding</u>, which will interfere with the observation of a red fecal pellet. Cover with cage top.
- Assay can be set up with or without food as long as all comparable runs are done the same way. If adding food, place 1-2 food pellets into a portion cup with water to create moistened chow (sterile if needed). Moistened chow reduces the risk of mice spilling water in the cage. Glass dishes can be used as cups to prevent flipping of the cups.
- Optional: record number of pellets produced, cumulatively in 5-minute bins, for the first30minutes of separation (this can be informative of fecal output).

7h

15 Every (5) 00:15:00 , check cages for a bright red pellet. Can observe through sides (with pen light if needed) or by opening cage top. Placing the cages on a white sheet of paper, rather than a black benchtop, will also make visualization easier. If food was provided, check in the

15m

- 16 Pellets can be collected at each cage check, for water content or molecular assays, as long as
 - all cages are similarly disrupted during collection. Removing fecal pellets will also aid in determination of the first red fecal pellet produced.

portion cups for red pellets as well. Ensure that whichever method of observation is used, all animals are disrupted similarly (ie. all animals have their cages opened).

- 17 Record time at which the first red pellet is observed for each mouse. For a normal, healthy adult mouse, this will be ~3-4hrs following gavage, but can range from 2-8+ hrs.
- 18 Once a red pellet is observed, return that mouse to its home cage.
- 19 To prevent stress associated with single-housing of mice, limit the single-housed portion of the assay to 6-8hrs, putting the maximum recordable transit time at 8-10hrs postgavage.
- 20 If the maximum allotted time is reached without production of a red pellet, return the mouse to their home cage, and record their transit time as the maximum.

Analysis

21 Compare the "time to red pellet" for each genotype or treatment group. This can be done pre- and post- exposures for toxicity assessments.