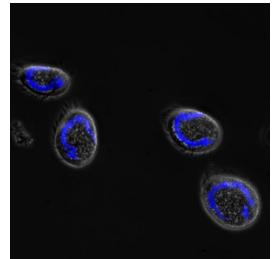


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Euplotes crassus transfection using Lipofectamine 2000 as vehicle (provisional)

Forked from [Euplotes crassus transfection using Lipofectamine 2000 as vehicle \(provisional\)](#)

In 1 collection



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Protist Research to Opti...



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Protocol status: In development

We are still developing and optimizing this protocol

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- 1 Collect 4×10^4 well-fed Euplotes crassus cells (we used E.coli as the only food source) by centrifugation at 400 rcf for 3 minutes.
- 2 Wash the cells twice with artificial sea water (see attachment for the recipe) and once with 500 mM sorbitol, 0.5 mM Tris-HCl, pH 7.0 (400 rcf for 3 minutes each time). Then resuspend Euplotes crassus cells in 50 μ l of the medium (500 mM sorbitol, 0.5 mM Tris-HCl, pH 7.0). To get this small volume you may require an additional minute of centrifugation.

Note

Recipe for complete sea water (1 L):

36 g Reef Crystals
1 ml Walne's solution
1 ml of 10 μ g/ml FeSO₄
0.2 ml of 2 mg/ml thiamine (light sensitive; store at 4 °C)
Add distilled water up to 1 L

- 3 Transfer the cells into wells within a 96-well plate for transfection.
- 4 Dilute 2.5 μ l of Lipofectamine 2000 Reagent in 25 μ l of the same medium of the cells (500 mM sorbitol, 0.5 mM Tris-HCl, pH 7.0).
- 5 Dilute 5 μ g of DNA dissolved in MilliQ H₂O (0.5-5 μ g/ μ l) in 125 μ l of the same medium of the cells (500 mM sorbitol, 0.5 mM Tris-HCl, pH 7.0).
- 6 Add 25 μ l of the diluted DNA to 25 μ l of the diluted Lipofectamine 2000 Reagent (1:1 ratio), and incubate them for 10 minutes at room temperature (before incubation mix them by pipetting up and down 5 times or vortex them for 10 seconds).
- 7 Add 10 μ l of the transfection complexes drop-wise to the 50 μ l of Euplotes crassus cells in medium (gently swirl the dish to ensure uniform distribution of the transfection complexes).
- 8 One hour after addition of Lipofectamine 2000 complexes, add 50 μ l of artificial sea water to the cells.
- 9 After another hour, add other 50 μ l of artificial sea water to the cells.

- 10 An hour later harvest the cells (400 rcf for 3 minutes), and wash them twice with artificial sea water (400 rcf for 3 min each time). Then resuspend them in 400 µl of the artificial sea water.
- 11 Transfer the cells into depression wells.
- 12 Incubate the cells at 24°C, then analyze them by fluorescence microscopy to determine gene expression.