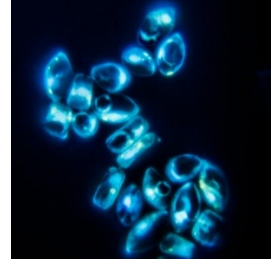


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## Establishment of axenic sea-ice diatom cultures, modified from Jaeckisch et al. (2011)

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**Protocol status:** Working

This protocol has been successful for the three tested species, *Nitzschia lecontei*, *Navicula perminuta*, *Fragilariopsis cylindrus* CCMP1102. Cultures have been stable and uncontaminated for > 18 months (DAPI screening and no growth on BD Difco 2216 agar plates).

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## Abstract

Table 1. Antibiotic concentrations used for establishing axenic sea-ice diatom cultures.

Antibiotic	Final concentration (µg/ml)
Ampicillin	50
Gentamycin	15
Streptomycin	125
Chloramphenicol	10
Ciprofloxacin	10
Penicillin	100

## References

**Guillard RRL. 1975.** Culture of phytoplankton for feeding marine invertebrates. In: Smith WL, Chanley MH eds. *Culture of marine invertebrate animals*. New York: Plenum, 29-60.

**Jaeckisch N, Yang I, Wohlrab S, Glöckner G, Kroymann J, Vogel H, Cembella A, John U. 2011.** Comparative Genomic and Transcriptomic Characterization of the Toxigenic Marine Dinoflagellate *Alexandrium ostenfeldii*. *PLoS ONE* **6**(12): e28012.

## Attachments



Axenic diatom protoc...

105KB

- 1 Pre-treat an optically dense diatom culture with 3 min of vortex-mixing ( $< 8^{\circ}\text{C}$ ) and 30 sec of ultrasonication (80% in 5-s pulses) on ice to remove attached bacteria from diatoms and EPS aggregates.
- 2 Centrifuge cells at  $1000 \times g$  for 10 min ( $< 8^{\circ}\text{C}$ ) and wash the pellet twice with sterile F/2 medium (Guillard, 1975) to remove the majority of loose bacterial cells.
- 3 Add six antibiotics to the medium according to Table 1 (see abstract), and incubate for 5 days at  $-1^{\circ}\text{C}$  and under light ( $20\text{--}45 \mu\text{mol photons m}^{-2} \text{s}^{-1}$ ).
- 4 Centrifuge cells at  $1000 \times g$  for 10 min ( $< 8^{\circ}\text{C}$ ) wash pellet in fresh F/2 medium.
- 5 Pick  $< 10$  cells to start the axenic cultures (pipetting or plate on F/2 agar to pick single colonies).
- 6 After recovery and growth ( $\sim 2$  months), check for bacterial cells using DAPI staining and after agar plating on half-strength BD Difco™ Marine Agar 2216 ( $< 8^{\circ}\text{C}$ ), kept at seawater salinity using artificial seawater (411 mM NaCl, 9.39 mM KCl, 26.1 mM  $\text{MgCl}_2 \cdot 6\text{H}_2\text{O}$ , 28.4 mM  $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$ , 5.01 mM TAPSO buffer).
- 7 If bacteria are still present, repeat protocol. Axenic cultures are usually established within the first or second attempt.