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Enzolution™ DDX1 ATPase Assay System Technical Manual

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Abstract

The Enzolution™ DDX1 Assay System is intended for use with the Transcreener® ADP^2 Assay Kits to measure the ATP-dependent RNA helicase activity of DDX1. DDX1, a DEAD Box helicase protein characterized by the conserved motif Asp-Glu-Ala-Asp (DEAD), is an ATP-dependent RNA helicase that unwinds RNA-RNA and RNA-DNA duplexes. ADP formation by the RNA helicase activity of DDX1 can be easily measured by the Transcreener ADP^2 Assay, a far-red, competitive fluorescence assay that enables single addition, mix-and-read detection in a continuous or endpoint format. The assay has been optimized and extensively validated for high throughput screening (HTS) and inhibitor dose response measurements using most multimode plate readers.

The Enzolution™ DDX1 Assay System provides the reagents needed for screening and profiling DDX1 inhibitors, including purified, full length human DDX1 (aa 1-740) and yeast RNA, when used in combination with Transcreener ADP^2 Assay Kits with FP, FI and TR-FRET readouts. The protocol is configured for 384-well plates; use of different multi-well plate formats will require adjustment of reagents concentrations utilized in the assay.

Image Attribution

Figure 1. Schematic Overview of the Enzolution™ DDX1 Assay System with the Transcreener ADP2 Assays.

Safety warnings

Note: Antibody (TR-FRET) and Tracer (FP and FI) concentrations remain constant in the 20 µL Complete Assay regardless of changes to other reaction conditions.

Before start

1. Read the entire protocol and note any reagents or equipment needed (see Section 2.2).
2. Check the plate reader and verify that it is compatible with the Transcreener® ADP2 Assays. [Full list of compatible plate readers and settings.](https://www.bellbrooklabs.com)
3. Please read and understand the Transcreener ADP2 Assay Technical Manuals prior to use with this kit.

Introduction

1 The Enzolution™ DDX1 Assay System is intended for use with the Transcreener® ADP^2 Assay Kits to measure the ATP-dependent RNA helicase activity of DDX1. DDX1, a DEAD Box helicase protein characterized by the conserved motif Asp-Glu-Ala-Asp (DEAD), is an ATP-dependent RNA helicase that unwinds RNA-RNA and RNA-DNA duplexes. ADP formation by the RNA helicase activity of DDX1 can be easily measured by the Transcreener ADP^2 Assay, a far-red, competitive fluorescence assay that enables single addition, mix-and-read detection in a continuous or endpoint format. The assay has been optimized and extensively validated for high throughput screening (HTS) and inhibitor dose response measurements using most multimode plate readers.

The Enzolution™ DDX1 Assay System provides the reagents needed for screening and profiling DDX1 inhibitors, including purified, full length human DDX1 (aa 1-740) and yeast RNA, when used in combination with Transcreener ADP^2 Assay Kits with FP, FI and TR-FRET readouts. The protocol is configured for 384-well plates; use of different multi-well plate formats will require adjustment of reagents concentrations utilized in the assay.

Key Applications:

- Screening for DDX1 inhibitors (or activators)
- Generating dose response curves and IC_{50} values for DDX1 inhibitors
- Kinetic and mechanistic analyses

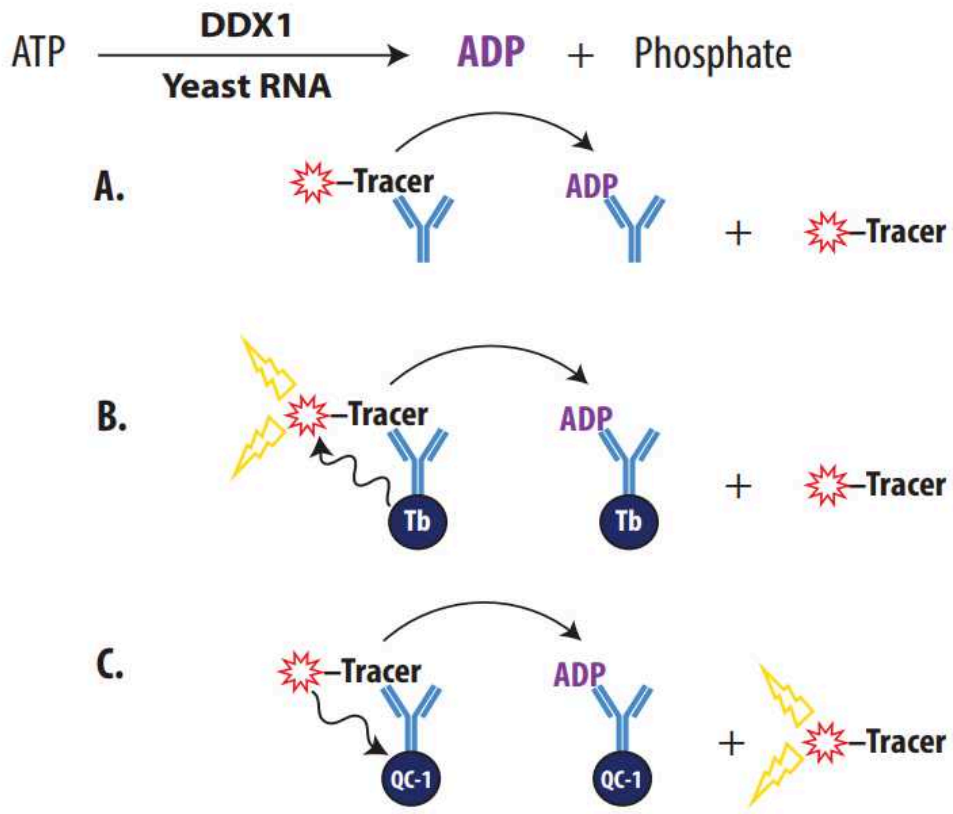


Figure 1. Schematic Overview of the Enzolution™ DDX1 Assay System with the Transcreener ADP^2 Assays.

For FP readout (A): ADP produced by the RNA helicase activity of DDX1 displaces an Alexa Fluor® 633 Tracer from the ADP^2 antibody, resulting in decreased fluorescence polarization.

For TR-FRET readout (B): ADP produced by the RNA helicase activity of DDX1 displaces a HiLyte647 Tracer from the ADP^2 Antibody conjugated to terbium (Tb), resulting in a decrease in TR-FRET.

For FI readout (C): ADP produced by the RNA helicase activity of DDX1 displaces an Alexa Fluor® 594 Tracer from the ADP^2 Antibody conjugated to an IRDye® QC-1 quencher, resulting in an increase in fluorescence intensity.

Product Specifications

2

	Product	Quantity	Part #	Part #
	Enzolution™ DDX1 ATPase Assay System	1,000 assays*	FP	3037-1K-FP

	Product	Quantity	Part #	Part #
			FI	3037-1K-FI
			TR-FRET	3037-1K-TR
		10,000 assays*	FP	3037-10K-FP
			FI	3037-10K-FP
			TR-FRET	3037-10K-TR

*The exact number of assays depends on the enzyme reaction conditions. The kits are designed for use with 384-well plates, using a 10 µL Enzyme Reaction and a 20 µL Complete Assay volume

Storage

DDX1 Enzyme and yeast RNA should be stored at -80°C; other reagents can be stored at –20°C. Though we have confirmed that the DDX1 Enzyme and the yeast RNA are stable and maintain greater than 80% activity up to 5 freeze-thaw cycles, we recommend aliquoting the reagents and snap-freezing for multiple uses to minimize loss of activity.

Use the reagents provided in this kit within 6 months from date of receipt.

Materials Provided

2.1

	Component	Composition	Notes
	DDX1 Enzyme	0.1 mg/mL (1.18 µM)* in 50 mM Tris-HCl, 100 mM NaCl, 1 mM TCEP, 1 mM EDTA, 10% Glycerol, pH 7.5	Full length (amino acids 1-740), N-Terminal 6xHis, 84.8 kDa. Sufficient enzyme is included in the kit to complete at least 1,000 assays (Part # 3037-1K) or 10,000 assays (Part # 3037-10K).
	Yeast RNA	10 mg/mL in H_2O	Yeast RNA purified from *Torulla*. Storage solution is nuclease-free water.
	Enzyme Assay Buffer D, 10X	500 mM TRIS (pH 7.5), 20 mM $MgCl_2$, 0.1% Triton X-100	Use Enzyme Assay Buffer D in the Enzyme Reaction and for preincubation with inhibitors. Changes to the assay buffer could affect enzyme activity and/or detection of ADP.
	384-Well Low	Corning #4514 -FP and FI Only	Polystyrene non-binding surface assay plates in either a 3-pack

	Component	Composition	Notes
	Volume Assay Plates	Corning #4513 – TR-FRET Only	(1,000+ Assays) or a 30-pack (10,000+ Assays). We strongly recommend the use of these plates as inconsistent results have been observed with other plates.

*The exact concentration may vary from batch to batch. Please refer to the Certificate of Analysis for an accurate concentration.

Materials Required But Not Provided

2.2

	Component	Notes
	Ultrapure Nuclease Free Water	Some deionized water systems are contaminated with nucleases that can degrade both nucleotide substrates and products, reducing assay performance. Use nuclease free water such as: Invitrogen Part # AM9930
	Plate Reader	A multimode microplate reader configured to measure FP, FI, or TR-FRET is required. Transcreener Assays have been validated on the following instruments: BioTek Synergy™2 and Synergy™4; BMG Labtech PHERAstar® Plus and CLARIOstar® Plus; Molecular Devices SpectraMax™ Paradigm; Perkin Elmer EnVision® and ViewLux; and Tecan Infinite® F500, Safire2™, and M1000. Full list of compatible plate readers and settings.
	Liquid Handling Devices	Use liquid handling devices that can accurately dispense sub-microliter volumes into 384-well plates.
	Laboratory Incubator	An incubator model that is capable of maintaining temperature stability at 30°C is required.

Transcreener® ADP^2 FP Assay - SOLD SEPERATELY

	Component	Composition	Notes
	ADP^2 Antibody	4 mg/mL solution in PBS with 10% glycerol*	1,000 assays (Part # 3010-1K) or 10,000 assays (Part # 3010-10K).
	ADP^2 Alexa Fluor® 633 Tracer	400 nM solution in 2 mM HEPES (pH 7.5) containing 0.01% Brij-35	1,000 assays (Part # 3010-1K) or 10,000 assays (Part # 3010-10K).

	Component	Composition	Notes
	Stop & Detect Buffer B, 10X	200 mM HEPES (pH 7.5), 400 mM EDTA, and 0.2% Brij-35	The EDTA in the Stop & Detect Buffer B quenches the DDX1 Enzyme Reaction by chelating Mg^{2+}. The final concentrations of Mg^{2+} and EDTA in the Complete Assay are 2 mM and 20 mM, respectively.
	ATP	5 mM	The ATP supplied in this kit can be used for the DDX1 Enzyme Reaction and the ATP/ADP standard curve.
	ADP	5 mM	The ADP is used for the ATP/ADP standard curve.

Transcreener® ADP^2 FI Assay - SOLD SEPERATELY

	Component	Composition	Notes
	ADP^2 Antibody-IRDye® QC-1	1.4 mg/mL solution in 100 mM KH_2PO_4 (pH 8.5)*	1,000 assays (Part # 3013-1K) or 10,000 assays (Part # 3013-10K).
	ADP^2 Alexa Fluor® 594 Tracer	800 nM solution in 2 mM HEPES (pH 7.5) containing 0.01% Brij-35	1,000 assays (Part # 3013-1K) or 10,000 assays (Part # 3013-10K).
	Stop & Detect Buffer B, 10X	200 mM HEPES (pH 7.5), 400 mM EDTA, and 0.2% Brij-35	The EDTA in the Stop & Detect Buffer B quenches the DDX1 Enzyme Reaction by chelating Mg^{2+}. The final concentrations of Mg^{2+} and EDTA in the Complete Assay are 2 mM and 20 mM, respectively.
	ATP	5 mM	The ATP supplied in this kit can be used for the DDX1 Enzyme Reaction and the ATP/ADP standard curve.
	ADP	5 mM	The ADP is used for the ATP/ADP standard curve.

Transcreener® ADP^2 TR-FRET Assay - SOLD SEPERATELY

	Component	Composition	Notes
	ADP^2 Antibody-Terbium Conjugate	800 nM solution in HEPES-buffered saline	1,000 assays (Part # 3011-1K) or 10,000 assays (Part # 3011-10K).
	ADP HiLyte647 Tracer	10 µM solution in 2 mM HEPES (pH 7.5) containing 0.01% Brij-35	1,000 assays (Part #3011-1K) or 10,000 assays (Part #3011-10K).

	Component	Composition	Notes
	Stop & Detect Buffer C, 10X	500 mM HEPES (pH 7.5), 200 mM EDTA, and 0.2% Brij-35	The EDTA in the Stop & Detect Buffer C quenches the DDX1 Enzyme Reaction by chelating Mg^{2+}. The final concentrations of Mg^{2+} and EDTA in the Complete Assay are 2 mM and 10 mM, respectively.
	ATP	5 mM	The ATP supplied in this kit can be used for the DDX1 Enzyme Reaction and the ATP/ADP standard curve.
	ADP	5 mM	The ADP is used for the ATP/ADP standard curve.

*The exact concentration may vary from batch to batch. Please refer to the Certificate of Analysis for an accurate concentration.

Before You Begin

3 1. Read the entire protocol and note any reagents or equipment needed (see **Section 2.2**).
2. Check the plate reader and verify that it is compatible with the Transcreener® ADP^2 Assays.
Full list of compatible plate readers and settings.
3. Please read and understand the Transcreener ADP^2 Assay Technical Manuals prior to use with this kit.

Protocol

4 The methods described below are for single-addition, endpoint detection: the DDX1 Enzyme Reaction is quenched by the addition of EDTA along with the detection reagents (see **Figure 2**). The methods were designed for 384-well plates using a 10 µL DDX1 Enzyme Reaction and 10 µL of detection/quench reagents (final volume 20 µL when the plates are read). The use of different plate densities or reaction volumes will require changes in reagent quantities (see **Section 5.1** for example reaction volumes).

The methods were optimized for initial velocity detection of ADP formation (≤ 20% conversion of ATP to ADP) by DDX1 with a sub-Km concentration of ATP (100 µM) and a saturating concentration of yeast RNA (1 mg/mL). These conditions will ensure sensitive detection of inhibitors that compete with ATP while minimizing the effects of compounds that inhibit DDX1 via non-specific interactions with RNA. Significant changes in the ATP concentration may require optimization of the ADP^2 Antibody concentration (in the case of the FP and FI readout modes) or the ADP Tracer (in the case of the TR-FRET readout mode) to adjust the dynamic range as described in the Transcreener ADP^2 Assay

Manual.

Note: Antibody (TR-FRET) and Tracer (FP and FI) concentrations remain constant in the 20 µL Complete Assay regardless of changes to other reaction conditions.

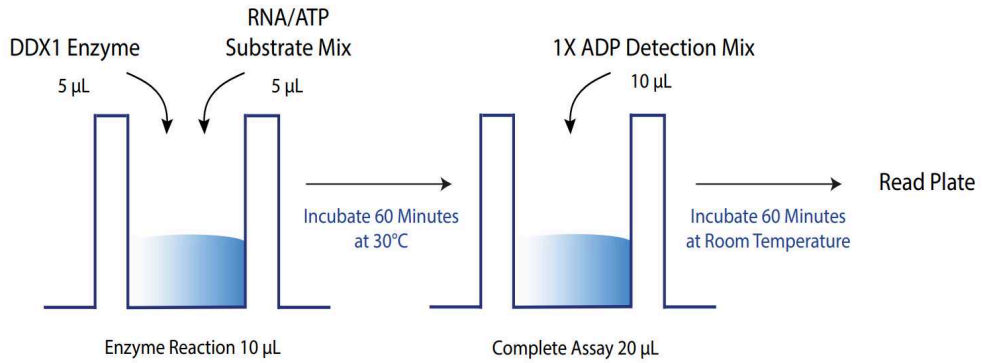


Figure 2. An Outline of the Procedure. The DDX1 Enzyme Reaction is initiated by the addition
of yeast RNA and ATP. After the Enzyme Reaction incubation is completed, ADP detection reagents are added (Transcreener ADP^2 Antibody and Tracer) along with EDTA to quench the DDX1 reaction.

10 µL Enzyme Reaction Components

	Component	Working Stock	Final Concentration in 10 µL
	Enzyme Assay Buffer D, 10X	1X in Nuclease Free Water	1X (50 mM Tris-HCl (pH 7.5), 2 mM $MgCl_2$, 0.01% Triton X-100)
	DDX1 Enzyme, 0.1 mg/mL (1.18 µM)	2X in 1X Enzyme Assay Buffer D	10 nM – 15 nM*
	ATP, 5 mM	200 µM in 1X Enzyme Assay Buffer D (with 2 mg/mL Yeast RNA)	100 µM
	Yeast RNA, 10 mg/mL	2 mg/mL in 1X Enzyme Assay Buffer D (with 200 µM ATP)	1 mg/mL

*See Section 4.1 for Determining the Optimal Enzyme Concentration.

Table 1. DDX1 Enzyme Reaction Components. Concentrations are provided for the standard protocol using 5 µL of DDX1 Enzyme Mix and 5 µL of RNA/ATP Substrate Mix for the Enzyme Reaction.

Component	1X ADP Detection Mix - Add 10 µL Per Well					
	FP		FI		TR-FRET	
	Detection Mix Concentration	Complete Assay	Detection Mix Concentration	Complete Assay	Detection Mix Concentration	Complete Assay
Stop & Detect Buffer	1X	0.5X	1X	0.5X	1X	0.5X
ADP Tracer	4 nM	2 nM	8 nM	4 nM	500 nM	250 nM
ADP Antibody	109 µg/mL	54.5 µg/mL	101 µg/mL	50.5 µg/mL	8 nM	4 nM

Table 2. 1X ADP Detection Mix Components. The optimal concentrations for each of the detection reagents based on the preferred readout mode are shown. Changes to the concentrations may require re-optimization of the assay.

Determining the Optimal Enzyme Concentration

4.1 Using the enzyme concentration suggested in the DDX1 Enzyme Certificate of Analysis should provide a robust signal that is within the linear range for ADP formation. However, for best results, we suggest performing an enzyme titration to identify the optimal enzyme concentration (EC_{50} to EC_{80}), especially when running the assay in a different buffer system or with a different ATP or Yeast RNA concentration. This example uses a 2X serial dilution; it should be performed at least in duplicate. If a compound screen is planned, you should include the solvent (e.g., DMSO) at its final assay concentration.

4.1.1 Enzyme Titration Steps

1. Prepare 1000 µL 1X Enzyme Assay Buffer D: Dilute 100 µL of 10X Enzyme Assay Buffer D in 900 µL Ultrapure Nuclease Free Water.

2. Prepare 30 µL of 500 nM DDX1 Enzyme: dilute 12.71 µL of 1.18 µM DDX1 Enzyme in 17.29 µL 1X Enzyme Assay Buffer D.

3. Add 10 µL of the DDX1 Enzyme to well 1 (including replicates).

4. Add 5 µL of 1X Enzyme Assay Buffer D to wells 2-11, DO NOT add the Assay Buffer to well 1.

5. Transfer 5 µL from well 1 to well 2 and mix by pipetting, then transfer 5 µL from well 2 to well 3 and mix by pipetting; repeat this serial dilution process until well 11 has received DDX1 Enzyme. Well 12 is to be used as a blank and should not include enzyme. **IMPORTANT: After mixing the last well (11) in the dilution series, remove 5 µL from that well only and discard, so that all the wells contain 5 µL final volume.**

6. Prepare 300 µL of RNA/ATP Substrate Mix: mix 60 µL of 10 mg/mL Yeast RNA and 12 µL of 5 mM ATP in 228 µL 1X Enzyme Assay Buffer D.

7. Start the Enzyme Reaction by adding 5 µL of the RNA/ATP Substrate Mix to every well (1-12). Gently mix for 40 to 60 seconds on a plate shaker. Incubate at 30°C for 60 minutes.

8. Prepare 600 µL 1X ADP Detection Mix based on the concentrations provided in

Table 2: 60 µL 10X Stop & Detect Buffer B, 6 µL ADP[2] Alexa Fluor 633 Tracer and 109 µg/mL ADP[2] Antibody in Ultrapure Nuclease Free Water.

Note: The 1X ADP Detection Mix varies for the readout mode used. Table 2 lists the 1X ADP Detection Mix for each readout.

9. Add 10 µL of 1X ADP Detection mix to every well (1-12), in replicate.
10. Gently mix on a plate shaker for 40 to 60 seconds and then allow it to incubate at room temperature for 60 minutes before measuring signals.

Note: The reagent volumes indicated above are sufficient for running the enzyme titration in duplicate plus excess for pipetting dead volume. Scaling of volumes can be performed if necessary.

For detection of inhibitors at single concentration or in dose response mode, we recommend selecting an enzyme concentration that produces a 50–80% change in signal (EC_{50} to EC_{80}) (see **Figure 3**). This will result in initial velocity conditions, which correspond to the linear phase of the reaction after conversion of values to ADP formation (see **Figure 6**). The EC_{50} is provided by common graphing programs; the EC_{80} enzyme concentration can be calculated from the EC_{50}, as follows:

$$EC_X = (X \div (100 - X))^{(1 \div |hillslope|)} \times EC_{50}$$

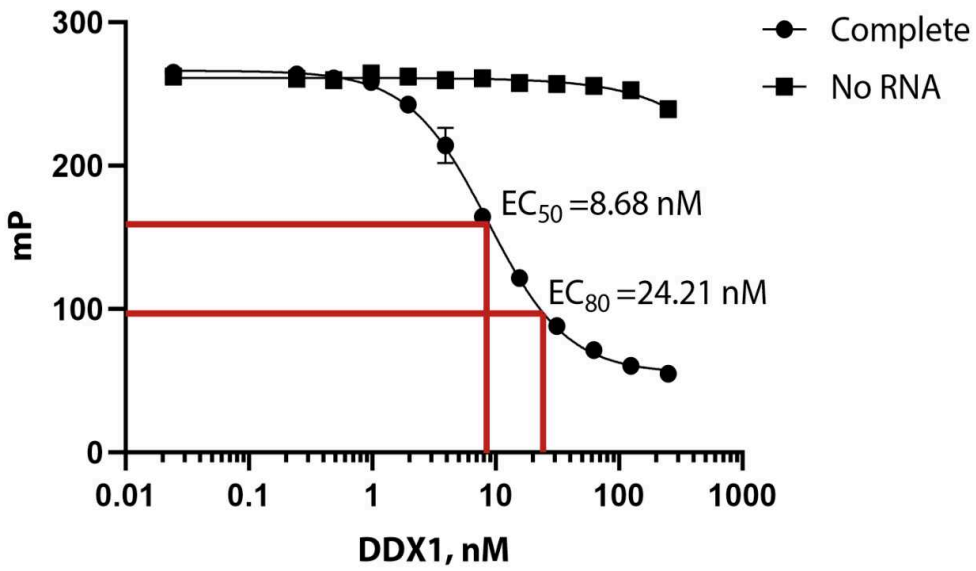


Figure 3. Enzyme Titration Curve. Example DDX1 Enzyme titration. The ideal range of enzyme concentrations is between EC_{50} and EC_{80}; the specific concentration may vary depending on the enzyme lot.

Performing Single Compound Screening and Dose-Response Assays

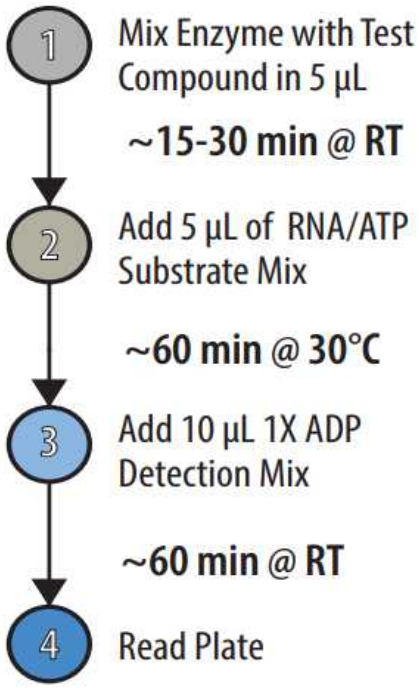
4.2 **4.2.1 Experimental Samples**

1. Perform a serial dilution of test compounds with your method of choice. Add the enzyme to the test compounds at the desired concentration so that the total volume of this mixture is 5 µL. Mix gently on a plate shaker for 40 to 60 seconds. Preincubate the Enzyme Inhibitor Mix for the desired time (typically at least 15 minutes) at room temperature to allow equilibration of the E-I complex.
Note: Final concentration of test compounds should be based on the volume of the Enzyme Reaction.

2. Start the Enzyme Reaction by adding 5 µL of the RNA/ATP Substrate Mix in each well. It is recommended to incubate the Enzyme Reaction at 30°C for 60 minutes.
Note: The final volume of the Enzyme Reaction mixture should be 10 µL for 384 well plates. See Section 5.1 for a list of other plate formats.

3. After the incubation, add 10 µL of 1X ADP Detection Mix to the 10 µL Enzyme Reaction and mix the 20 µL Complete Assay using a plate shaker.
Note: The 1X ADP Detection Mix varies for the readout mode used. Table 2 lists the 1X ADP Detection Mix for each readout.

4. Incubate at room temperature for 60 minutes before measuring signals.



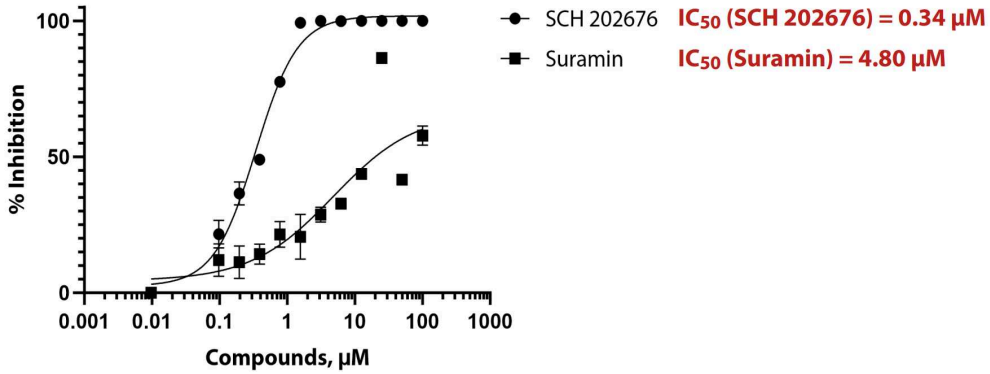


Figure 4. Dose-Response Curve. Example titration of probe inhibitors SCH 202676 and Suramin in the presence of DDX1 Enzyme.

Setting Up a Standard Curve

4.3 Use of a standard curve for conversion of values to amount of ADP formed allows quantitative measurement of the enzyme activity and accurate IC_{50} determinations; it is not typically done for screening at single concentrations. The standard curve mimics the Enzyme Reaction (as ATP concentration decreases, ADP concentration increases); the adenine nucleotide concentration remains constant. The ADP/ATP standard curve allows calculation of the concentration of ADP produced in the Enzyme Reaction and, therefore, the percent ADP conversion.

In this example, a 12-point standard curve was prepared using the concentrations of ADP and ATP shown in the following Table. Commonly, 8- to 12-point standard curves are used. Here we describe preparation of a standard curve from 100 µM to 0.5 µM ADP, which encompasses the appropriate range for this assay.

	% Conversion	ATP (µM)	ADP (µM)
	100	0	100
	50	50	50
	25	75	25
	15	85	15
	10	90	10
	8	92	8
	4	96	4

	% Conversion	ATP (µM)	ADP (µM)
	2	98	2
	1	99	1
	0.5	99.5	0.5
	0.2	99.8	0.2
	0	100	0

Table 3. Concentration of ATP and ADP to prepare a 12-point standard curve.

4.3.1 Preparing the Standard Curve

1. Prepare 1X Enzyme Assay Buffer D.

2. Dilute 5 mM ATP and ADP to 100 µM in 1X Enzyme Assay Buffer D. The volume for both dilutions should be based on the total amount of volume to be utilized during preparation of the ATP/ADP percent conversion solutions.

3. Starting from 100 µM, add proportional values of ATP and ADP with the method of choice (i.e., Automatic Dispenser). For manual preparations, add reagents in separate microcentrifuge tubes to generate the desired percentage conversions utilized in the standard curve. An example of the dilution scheme can be found in **Table 3**. Once all dilutions are completed, add 10 µL of each solution to the respective wells.

4. Afterwards, add 10 µL of 1X ADP Detection Mix. The 1X ADP Detection Mix varies for the readout mode used. **Table 2** lists the 1X ADP Detection Mix for each readout.

5. Finally, mix for 40 to 60 seconds. Incubate at room temperature for 60 minutes before measuring signals.

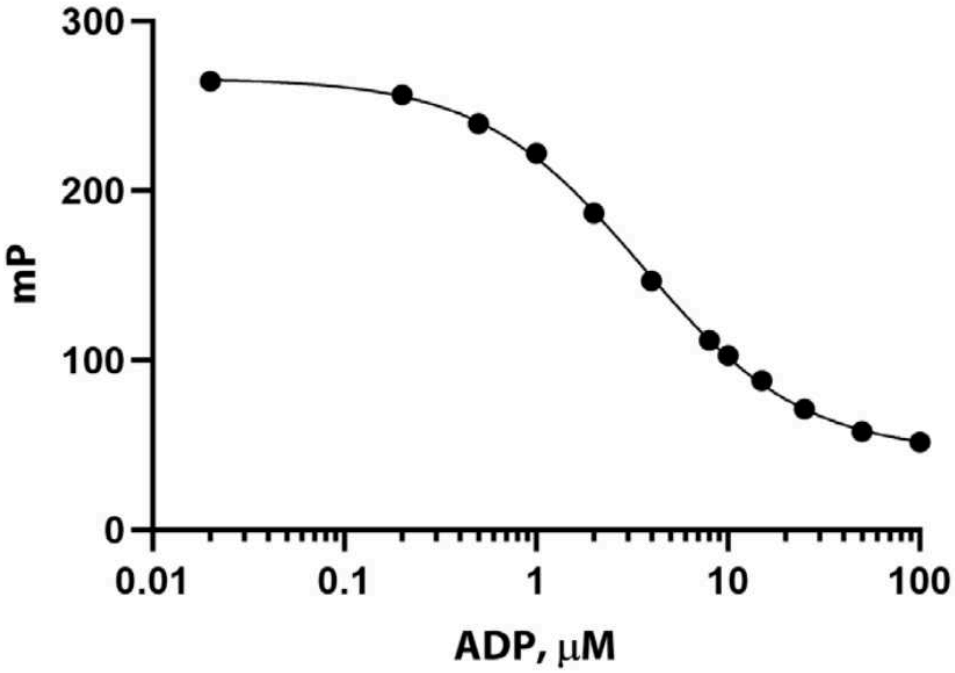


Figure 5. ADP Standard Curve. Standard curve using 1X ADP Detection Mix for FP readout as shown in **Table 2** (109 µg/mL ADP^2 antibody, 4 nM ADP^2 Tracer).

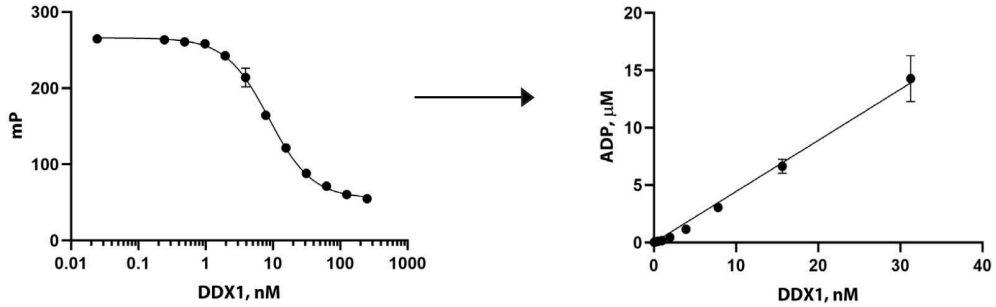


Figure 6. Enzyme Titration Curve Converted to ADP Formed. Raw polarization signal (mP) is converted to ADP formed using a standard curve as described in **Section 4.3**. Only the linear portion of the graph is shown; interpolation was performed using GraphPad Prism.

Measuring Assay Robustness with Z'

4.4 By taking into account both dynamic range and data variability at the high and low ranges of the assay, the Z′ statistic provides a measure of what is of most interest when considering the suitability of an assay for HTS: the usable screening or “assay window.” It is a dimensionless coefficient for the quality of the screening window that is relevant

for any assay, regardless of detection method or readout, without the intervention of test compounds. As a guideline, a Z′ value of 0.5 or greater is generally considered to be indicative of a very good screening window for a biochemical assay, thus the assay is an excellent assay. When running the DDX1 Assay, run the controls with and without enzyme (no test compound) to achieve final results. Use the following formula to determine Z′.

$$Z' = 1 - \frac{[(3 \times SD_{\text{No Enzyme}}) + (3 \times SD_{\text{Complete Reaction}})]}{|(\text{Mean}_{\text{No Enzyme}}) - (\text{Mean}_{\text{Complete Reaction}})|}$$

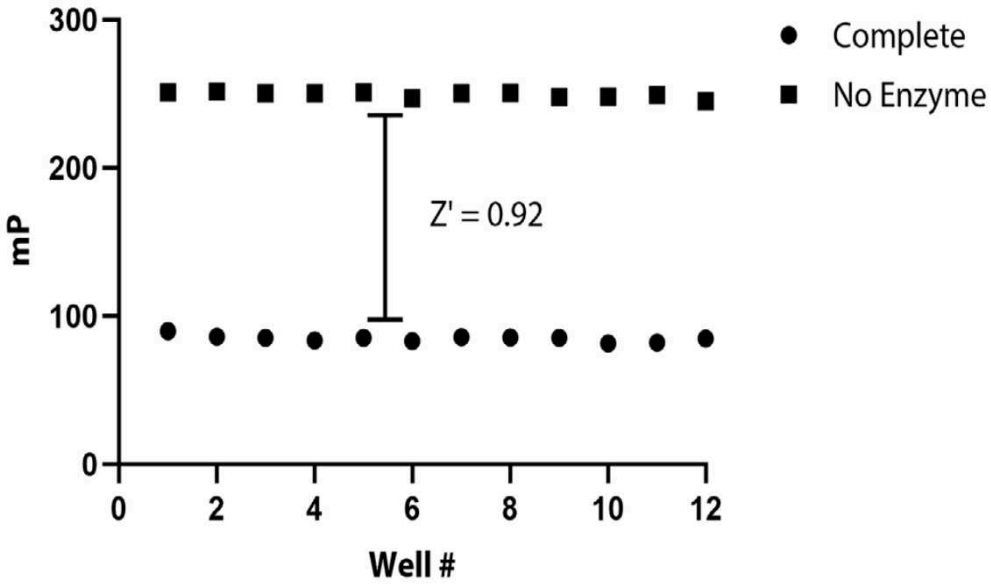


Figure 7. Z′ Measurement. Complete Assay is performed with and without DDX1 Enzyme (n=12). Z′ is then calculated based on the formula shown in **Section 4.4**.

Appendix

5

Using the Assay with Different Volumes and Plate Formats

5.1

	Component	Total Volume	Enzyme Reaction Volume	1X ADP Detection Mixture Volume
	96 Well Low Volume Plate	50 µL	25 µL	25 µL
	384 Well Low Volume Plate	20 µL	10 µL	10 µL
	1536 Well Low Volume Plate	8 µL	4 µL	4 µL

Please check the working plate volumes from the manufacturer to ensure they are within the suggest volumes ranges of your plate.

Links to Applicable Application Notes

5.2

- A Guide to Navigating Hit Prioritization After Screening Using Biochemical Assays
- A Guide to Measuring Drug-Target Residence Times with Biochemical Assays
- Using GraphPad Prism to Interpolate Data from a Standard Curve to Generate a Dose-Response
- List of Commonly Used Plate Readers and Settings

Contact Information

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