ELISA for quantification of macrophage-colony stimulating factor (M-CSF) in human serum or plasma.

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1. An anti-human macrophage-colony stimulating factor (M-CSF) coating antibody is adsorbed onto the microwells by incubation overnight at 4°C with carbonate-bicarbonate buffer.

2. Add 50 µl of human serum or plasma. M-CSF present in the serum sample binds to antibodies adsorbed into the microwells.

3. The microplate is blocked with 3% non-fat milk-PBS buffer and later wash to remove unbound proteins.

4. Fifty (50) µl of biotin-conjugated anti-M-CSF antibody is added. The optimal dilution must be investigated.

5. The microplate is rewashed with PBS-Tween 20 buffer, pH 7.4.

6. One hundred µl of streptavidin-HRP conjugate is added and it binds to the biotin-conjugated anti-M-CSF antibody.

7. The plate is washed following incubation to remove the unbound Streptavidin-HRP conjugate.
Add 100 µl of 3,3',5,5'-tetramethylbenzidine (TMB; Sigma-Aldrich) into each well.

Incubate the microwells in the dark for 15 min.

A colored product is formed in proportion to the quantity of M-CSF present in the sample or standard.

The reaction is terminated by addition of 100 µl 3M H2SO4 and the absorbance is measured at 450 nm.

A standard curve is made from 7 human M-CSF standard dilutions and the human M-CSF sample concentration is determined.

For better results place the microplate on a microplate shaker in every incubation.