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Electroporation Protocol



Forked from [Electroporation Protocol](#)

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External link: <https://www.neb.com/protocols/2012/06/21/making-your-own-electrocompetent-cells>

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Protocol status: In development

We are still developing and optimizing this protocol

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Abstract

This protocol may be used with electrocompetent cells prepared by you according to [this protocol](#).

Guidelines

Appropriate Antibiotics for Your Application

Antibiotics for Plasmid selection

Antibiotic	Working Concentration
Ampicillin	100 µg/ml
Carbenicillin	100 µg/ml
Chloramphenicol	33 µg/ml
Kanamycin	30 µg/ml
Streptomycin	25 µg/ml
Tetracycline	15 µg/ml

Electroporation Protocol


The electroporation protocol will vary depending on the strain so this protocol may need to be optimized. For control electroporation dilute pUC19 to 10 pg/µl with Milli-Q water.

Calculation:

If the culture was diluted 1000-fold when plated, the total cfu per ml is 1000 times the number of colonies counted. The cfu is divided by the amount of pUC19 (10 pg per ml)

$$\text{cfu/} \mu\text{g} = (\text{colonies counted} \times 1000) / (0.00001 \mu\text{g pUC19})$$






Safety warnings

 The electroporation protocol will vary depending on the strain so this protocol may need to be optimized.

Before start

For control electroporation dilute pUC19 to 10 pg/µl with Milli-Q water.




- 1 Turn on electroporator and set to 1.7-2.5 kv (optimize for strain), 200 ohms and 25 μ F
- 2 Place recovery SOC in 37°C water bath
- 3 Pre-warm LB-antibiotic plates at 37°C
- 4 Thaw cells on ice for 10 min or use freshly made cells
 00:10:00
- 5 Place appropriate number of microcentrifuge tubes and 1 mm-electroporation cuvettes on ice
- 6 Flick the tube containing cells a few times to mix and add **25 μ l** of competent cells to the microcentrifuge tube
 25 μ L
- 7 Add **1 μ l** of a 100 pg/ μ l to 1 ng/ μ l DNA solution (in DI water) to the cells in the microcentrifuge tube
 1 μ L
- 8 Transfer the DNA-cell mixture to the cold cuvette, tap on countertop 2X, wipe water from exterior of cuvette and place in the electroporation module and press pulse (**you don't hold the button down**)
- 9 Immediately add **975 μ l** of 37°C SOC, mix by pipetting up and down once and transfer to a microcentrifuge tube, 5 ml culture tube, or 15 ml centrifuge tube.
 975 μ L
- 10 Place in the shaker/incubator at 37°C incubator for 1 h
 01:00:00
- 11 Make appropriate dilutions

Note

When using 100 pg - 1 ng of DNA, make three dilutions:
Dilute 1 μ l of cells into 990 μ l SOC and plate 100 μ l. (10000-fold dilution)
Dilute 10 μ l of cells into 990 μ l SOC and plate 100 μ l. (1000-fold dilution)
Dilute 100 μ l of cells into 900 μ l SOC and plate 100 μ l. (100-fold dilution)



12 Incubate overnight at 37°C

 16:00:00