

Nov 14, 2018

Electroporation of *Oxyrrhis marina*

 [Nature Methods](#)

DOI

<https://dx.doi.org/10.17504/protocols.io.vcne2ve>

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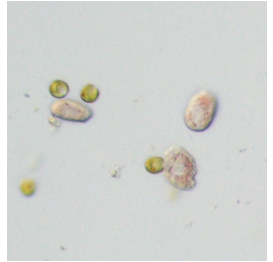
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External link: <https://doi.org/10.1038/s41592-020-0796-x>

Protocol Citation: Yutaka Hanawa, Yoshihisa Hirakawa, Patrick Keeling 2018. Electroporation of *Oxyrrhis marina*. **protocols.io**
<https://dx.doi.org/10.17504/protocols.io.vcne2ve>

**Manuscript citation:**

Faktorová D, Nisbet RER, Robledo JAF, Casacuberta E, Sudek L, Allen AE, Ares M, Aresté C, Balestreri C, Barbrook AC, Beardslee P, Bender S, Booth DS, Bouget F, Bowler C, Breglia SA, Brownlee C, Burger G, Cerutti H, Cesaroni R, Chiurillo MA, Clemente T, Coles DB, Collier JL, Cooney EC, Coyne K, Docampo R, Dupont CL, Edgcomb V, Einarsson E, Elustondo PA, Federici F, Freire-Beneitez V, Freyria NJ, Fukuda K, García PA, Girguis PR, Gomaa F, Gornik SG, Guo J, Hampl V, Hanawa Y, Haro-Contreras ER, Hehenberger E, Highfield A, Hirakawa Y, Hopes A, Howe CJ, Hu I, Ibañez J, Irwin NAT, Ishii Y, Janowicz NE, Jones AC, Kachale A, Fujimura-Kamada K, Kaur B, Kaye JZ, Kazana E, Keeling PJ, King N, Klobutcher LA, Lander N, Lassadi I, Li Z, Lin S, Lozano J, Luan F, Maruyama S, Matute T, Miceli C, Minagawa J, Moosburner M, Najle SR, Nanjappa D, Nimmo IC, Noble L, Vanclová AMGN, Nowacki M, Nuñez I, Pain A, Piersanti A, Pucciarelli S, Pyrih J, Rest JS, Rius M, Robertson D, Ruaud A, Ruiz-Trillo I, Sigg MA, Silver PA, Slamovits CH, Smith GJ, Sprecher BN, Stern R, Swart EC, Tsaousis AD, Tsypin L, Turkewitz A, Turnšek J, Valach M, Vergé V, Dassow Pv, Haar Tvd, Waller RF, Wang L, Wen X, Wheeler G, Woods A, Zhang H, Mock T, Worden AZ, Lukeš J, Genetic tool development in marine protists: emerging model organisms for experimental cell biology. *Nature Methods* 17(5). doi: [10.1038/s41592-020-0796-x](https://doi.org/10.1038/s41592-020-0796-x)

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Protocol status: In development

We are still developing and optimizing this protocol

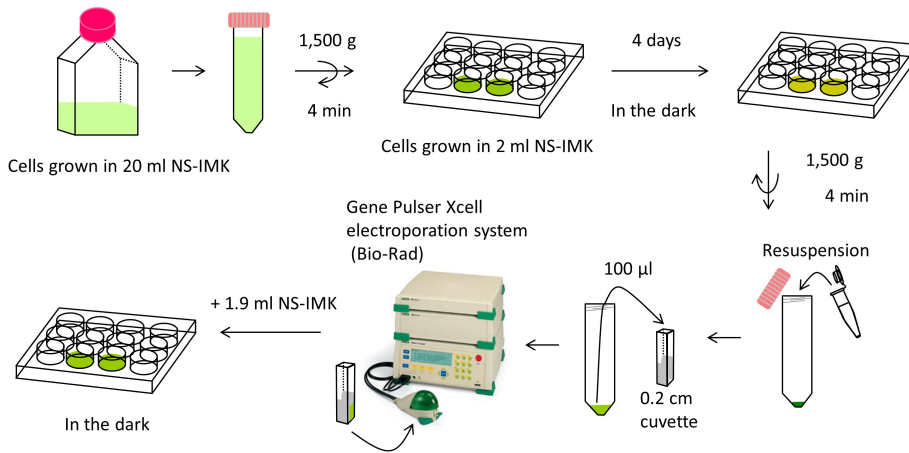
Created: November 06, 2018

Last Modified: November 14, 2018

Protocol Integer ID: 17518

Keywords: electroporation of oxyrrhis marina, electroporation, marina

Abstract



Culture condition of *O. marina*

- 1 Transfer *Oxyrrhis marina* (NIES-494) cells to 20 mL of fresh IMK medium (Nihon Pharmaceutical Co., Ltd.) in a plastic flask (IWAKI 75 cm²) at the concentration of 200 cells/mL, and add *Pyramimonas parkeae* (NIES-254) as feed at the concentration of 1×10⁴ cells/mL.

Grow cells at 22°C with a light/dark cycle of 14h/8h for 14 days. After two weeks, the cell density of *O. marina* will reach approximately 1×10⁴ cells/mL.

Concentration of *O. marina* cells

- 2 Collect *O. marina* and *P. parkeae* cells from 20 mL culture by centrifugation at 1,500 g for 4 min with a swing rotor.

Resuspend cells with 2 mL fresh IMK medium, and then transfer to a 12-well plastic plate. Incubate the plate in a dark condition at 22°C.

After 4 days, the cell density of *O. marina* will be increased from 1×10⁵ cells/mL (0 day) to 5×10⁵ cells/mL (4 days), in contrast with the drastic decrease of *P. parkeae* cells.

Electroporation with Bio-Rad Gene Pulser Xcell

- 3 Harvest *O. marina* cells at 1,500 g for 4 min with a swing rotor, and then resuspend the cell pellet by 100 µL Gene Pulser electroporation buffer (Bio-Rad #1652676) at the final concentration of 1×10⁶ to 5×10⁶ cells/mL.

Add DNA (5–25 µg) or RNA (5 µg) to the cell solution, and transfer it to a 0.2 cm cuvette (Bio-Rad). Immediately after electrophoresis, add 1.9 mL fresh IMK medium into the cuvette, and transfer the cells to a 12-well plastic plate.