



Mar 18, 2023

Economic and easy bacterial and yeast colony PCR with 2x premix Gotaq-Green (Promega) or DreamTaq-Green (Thermo Fisher)

DOI

<https://dx.doi.org/10.17504/protocols.io.eq2ly74jqlx9/v1>

Dariusz Abramczyk¹

¹University of Edinburgh



Dariusz Abramczyk

University of Edinburgh

Create & collaborate more with a free account

Edit and publish protocols, collaborate in communities, share insights through comments, and track progress with run records.

Create free account

OPEN  ACCESS



DOI: <https://dx.doi.org/10.17504/protocols.io.eq2ly74jqlx9/v1>

Protocol Citation: Dariusz Abramczyk 2023. Economic and easy bacterial and yeast colony PCR with 2x premix Gotaq-Green (Promega) or DreamTaq-Green (Thermo Fisher) . **protocols.io** <https://dx.doi.org/10.17504/protocols.io.eq2ly74jqlx9/v1>

License: This is an open access protocol distributed under the terms of the **Creative Commons Attribution License**, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

Protocol status: Working

We use this protocol and it's working



Created: March 10, 2023

Last Modified: March 18, 2023

Protocol Integer ID: 78495

Keywords: yeast colony pcr, colony pcr, yeast, pcr, method gotaq promega, modified method gotaq promega, thermo fisher, promega, thermo

Abstract

Slightly modified method Gotaq **Promega** or DreamTaq (**Thermo**), economical, quick and ready-to-load. Perfect for yeast genotyping and screening

Protocol materials



GoTaQ Green Master Mix **Promega Catalog #M7122**

1 Bacterial colony PCR

 GoTaq Green Master Mix **Promega Catalog #M7122**

PCR reactions using ProFlex PCR system, Applied Biosystems. **Thermal Cycler**

1.1

	A	B	C
	Reagent Name	Initial Conc.	Volume (μL)
	gotaq green 2x	2X	5
	oligo 1	10uM	0.6
	oligo 2	10 uM	0.6
	cell suspension in water	cells suspension	0.8
	ddH2O	N/A	3
	TOTAL Volume:	10 uL	

	A	B	C	D
	Number of Cycles	Step Name	Temperature	Duration
	1	Initial Denaturation	98oC	10min
	30	Denaturation	98oC	10 sec
		Annealing	50-55oC	30 sec
		1min (depends on the length)	72oC	1min per 1kb
	1	Hold	4-10oC	hold

Note

Maximum (good quality) PCR product up to ~3kb. The best yield is 0.2kb-2kb

Note

pick a colony from agar plate and swirl in sterile water  50-100 µL . From liquid culture, dilute saturated culture  5-10 µL in sterile  50-100 µL water. This is a source for reaction (taking  0.8 µL).

Keep the rest in ice, for re-culture of positively verified clones.

- 1.2 Load on 1-1.2% agarose gel  8-10 µL and visualise under GelDoc or another UV source.

2 Yeast colony PCR (S.cerevisiae, Pichia, Candida)

- 2.1 Alkaline yeast cells lysis treatment. Pick a yeast colony from agar plate and swirl in sterile 20mM NaOH  50-100 µL .
 Optionally, a liquid culture or a agar-plate pick colony resuspend in the sterile water or YPD, take small volume (~20uL) and mix with the same volume of 40mM NaOH.
 20mM NaOH-cells resuspension incubate  95 °C  00:10:00 and immediately transfer to ice/  4 °C for at least  00:05:00

15m

2.2

	A	B	C
	Reagent Name	Initial Conc.	Volume (µL)



	A	B	C
	gotaq green 2x	2X	5
	oligo 1	10uM	0.6
	oligo 2	10 uM	0.6
	cell suspension in 20mM	yeast suspension	0.8
	ddH2O	N/A	3
	TOTAL Volume:	10 uL	

	A	B	C	D
	A	B	C	D
	Number of Cycles	Step Name	Temperatur e	Duration
	1	Initial Denaturatio n	98oC	2min
	30-33	Denaturatio n	98oC	10 sec
		Annealing	52-55oC	1min 30sec
		1min (depends on the length)	72oC	1min per 1kb
	1	Hold	4-10oC	hold

Note

Maximum PCR product up to ~3kb. The best yield is 0.2kb-2kb



Note

Important - annealing 1min30sec

- 2.3 Load on 1-1.2% agarose gel  8-10 μL and visualise under GelDoc or another UV source.