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Dye-terminator DNA sequencing V.3

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We use this protocol and it's working

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Abstract

This protocol (based on the BigDye® Terminator v3.1 Cycle Sequencing Kit) is for performing terminator cycling sequencing reactions for Sanger sequencing of amplified PCR products or plasmid DNA on the 3130X genetic analyser (Applied Biosystems).

Attachments

PDF

BigDye Terminator v3...

350KB

Materials

MATERIALS

- Antarctic Phosphatase - 1,000 units **New England Biolabs Catalog #**M0289S
- 96 well PCR Plate Non-skirted **Phenix Research Catalog #**MPS-499
- Nuclease-free water (e.g. MilliQ or HPLC grade water)
- primers
- EDTA
- 10 mM dNTPs **Life Technologies Catalog #**10297-018
- Ethanol **Merck Millipore (EMD Millipore) Catalog #**100983
- BigDye™ Terminator v3.1 Cycle Sequencing Kit **Thermo Fisher Catalog #**4337454
- Exonuclease I (E. coli) **NEB Catalog #**M0293S
- Hi-Di™ Formamide **Thermo Fisher Scientific Catalog #**4311320

Troubleshooting

Before start

Optimize PCR cycling (if sequencing amplified PCR products) to ensure your reaction produces a single product. Perform gel excision or PCR clean-up with the potential inclusion of incubating with Antarctic phosphatase and Exonuclease 1 to dephosporylate and degrade unincorporated dNTPs in PCR reactions to prepare templates for DNA sequencing.

Terminator cycling reaction

1 Perform sequencing reaction in PCR tubes (or 96-well plate) with BigDye Terminator cycling kit and forward or reverse primers. 30m

	Component	Volume (µl)
	v3.1 Ready reaction mix	1
	5X Sequencing buffer	1.5
	20 µM F/R Primer	0.5
	Template (plasmid or cleaned PCR product)	50-150 ng DNA (plasmid or PCR product)
	Nuclease-free water	to 10 µl

BigDye Terminator Cycling reaction

5x reaction buffer: 400 mM TRIS, 10 mM $MgCl_2$

2 Run the following thermal cycling protocol: 4h

1. 1 min at 96 °C
2. 30-40 cycles: 96 °C for 10 seconds, 50 °C for 5 seconds, and 60 °C for 4 min.
3. Hold at 4-12 °C.

Purification

1h 30m

3 Transfer PCR reaction to eppendorf tube. To the reaction, add 2.5 µL of 125 mM EDTA (make sure it touches bottom of tube).

4 Add 30 µl of 100% ethanol, mix well (inversion).

5 Incubate at room temperature for 15 minutes.

6 Centrifuge at 4 °C at max speed for 30 minutes.

7 Discard supernatant and add 50 µl of ice-cold 70% ethanol.

8 Centrifuge at 4 °C at max speed for 5 minutes.

9 Discard supernatant and allow to air-dry in the dark for >15 minutes.

Prepare for sequencing

10 Resuspend the pellet (likely transparent) in 7.5 µL HiDi Formamide (add to any empty wells). Incubate at RT for 5 minutes then transfer to plate. Spin down briefly.

11 Incubate plate at 95 °C for 3 minutes (denature) then place immediately on ice. Spin down briefly.

12 Submit for sequencing on 3130X genetic analyser (Applied Biosystems). Keep samples on ice.