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# Dry-Sponge DNA extraction

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**Protocol status:** In development
**We are still developing and optimizing this protocol**

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## Abstract

Sample preparation optimization by sodium acetate precipitation for DNA extraction from Dry-Sponge samples.

## Sample Preparation

15h 55m

1 Extract the sponge liquid by squizing the sponge. 5m

2 Move (pour) the liquid content extracted from the sponge into 50 mL tubes (expect 15 mL eg. Longmire's buffer) 5m

3 Add 33 mL of pure ethanol and 1.m mL of Sodium Acetate 3M to the sample tube (achieving approx. 50 mL of liquid voulme). 5m

4 incubate the samples overnight at -20 °C 12h

5 Cool down the centrifuge to 4-8 °C 15m

6 Centrifuge at 4000g for 80 min at 6°C. 1h 20m

7 Discard the supernatant and take up to 250 mg of the sediment into PowerBead tubes (Qiagen, Hilden, Germany) 5m

8 Follow the DNeasy PowerSoil kit instruction for the DNA extraction. 2h