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Dry-Sponge DNA extraction

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Amir reza Varzandi¹

¹University of Turin



Amir reza Varzandi

University of Turin

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Protocol status: In development

We are still developing and optimizing this protocol

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Abstract

Sample preparation optimization by sodium acetate precipitation for DNA extraction from Dry-Sponge samples.



Sample Preparation

15h 55m

- 1 Extract the sponge liquid by squeezing the sponge. 5m
- 2 Move (pour) the liquid content extracted from the sponge into 50 mL tubes (expect 15 mL eg. Longmire's buffer) 5m
- 3 Add 33 mL of pure ethanol and 1.m mL of Sodium Acetate 3M to the sample tube (achieving approx. 50 mL of liquid voulme). 5m
- 4 incubate the samples overnight at -20 °C 12h
- 5 Cool down the centrifuge to 4-8 °C 15m
- 6 Centrifuge at 4000g for 80 min at 6°C. 1h 20m
- 7 Discard the supernatant and take up to 250 mg of the sediment into PowerBead tubes (Qiagen, Hilden, Germany) 5m
- 8 Follow the DNeasy PowerSoil kit instruction for the DNA extraction. 2h