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# DNA/RNA Radiolabeling Protocol

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Liz O'Brien<sup>1</sup>, Connor Tsuchida<sup>1</sup>

<sup>1</sup>University of California, Berkeley

The Center for Genome ...



Meredith Triplet

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**Protocol status:** Working

**We use this protocol and it's working**

**Created:** October 18, 2019

**Last Modified:** November 22, 2019

**Protocol Integer ID:** 28818

**Keywords:** Radiolabeling, DNA, RNA, CasX, TS, NTS

## Attachments



Radiolabeling\_CasX\_D...

141KB

## Guidelines

### CasX TS/NTS with non-hydrolysable spacers:

#### TS:

5'-CGCTAGCTACGT**T\*T\*G\*A\*T\*T\*T\*C\*T\*G\*C\*T\*G\*C\*A\*G\*G\*A\****TGAAATCCCGTAATCGCGC*-3'

MW: 15664.2 g/mol

Concentration: X  $\mu$ M

\* = phosphothioate, **bold letters** = PAM, *italic letters* = spacer

For 10 pmol of TS: X  $\mu$ l of stock

#### NTS:

5'-GCGCGATTACGGGAT *TT*CAT**C\*C\*T\*G\*C\*A\*G\*C\*A\*G\*A\*A\*A\*A\*T\*C\*A\*A\*A**\*CGTAGCTAGCG-3'

MW: 15749.3 g/mol

Concentration: X  $\mu$ M

\* = phosphothioate, **bold letters** = PAM, *italic letters* = spacer

For 10 pmol of NTS: X  $\mu$ l of substrate

### Labelling reaction setup:

#### \*TS:

XX  $\mu$ l DNA or RNA (10 pmoles)

2.5  $\mu$ l 10x PNK buffer

0.5  $\mu$ l PNK enzyme

1.5  $\mu$ l P32-gamma-ATP

XX mL dH<sub>2</sub>O (DEPC for labeling RNA) to 25  $\mu$ l

#### \*NTS:



XX  $\mu$ l DNA or RNA (10 pmoles)  
2.5  $\mu$ l 10x PNK buffer  
0.5  $\mu$ l PNK enzyme  
1.5  $\mu$ l P32-gamma-ATP  
XX mL dH<sub>2</sub>O (DEPC for labeling RNA) to 25  $\mu$ l

## Materials

### MATERIALS

⊗ T4 Polynucleotide Kinase (3' phosphatase minus) - 200 units **New England Biolabs Catalog #M0236S**

⊗ 10X T4 PNK Reaction Buffer **New England Biolabs**

⊗ ATP [ $\gamma$ -<sup>32</sup>P]- 3000Ci/mmol 10mCi/ml Lead 100  $\mu$ Ci (P32-gamma-ATP) **Perkin Elmer Catalog #NEG002A100UC**

⊗ HiTrap Desalting columns with Sephadex G-25 resin **GE Healthcare Catalog #29048684**

## Safety warnings

⚠ Please see SDS (Safety Data Sheet) for hazards and safety warnings.



## 1 Set up labeling reaction:

X $\mu$ l	DNA or RNA (10 pmol es)
2.5 $\mu$ l	10x PNK buffer
0.5 $\mu$ l	PNK enzyme
1.5 $\mu$ l	P32-gamma-ATP
	dH <sub>2</sub> O (DEPC for labeling RNA) to 25 $\mu$ l

### Note

Mix the DNA, buffer, enzyme, and H<sub>2</sub>O at the bench, and then add the DNA/enzyme mixture to ATP-filled tubes in a radioactive use area.

2 Incubate at 37 °C for 00:30:00 .



3 Heat inactivate the PNK at 65 °C for 00:20:00 .








4 Prepare G25 columns (from GE, green box): vortex thoroughly, twist cap ¼ turn, snap off bottom, spin for 00:01:00 at 3000 rpm to get rid of liquid.



5 Add 50  $\mu$ L H<sub>2</sub>O to a labeled eppendorf tube, place G25 column in it.





- 6 Add  25  $\mu\text{L}$   $\text{H}_2\text{O}$  to each labeling reaction after heat inactivation is done. 
- 7 Apply entire reaction (now 50  $\mu\text{l}$  total) to G25 column resin.
- 8 Spin for  00:02:00 at  3000 rpm . 
- 9 Since 50  $\mu\text{l}$   $\text{H}_2\text{O}$  were in bottom of tube and you add your 50  $\mu\text{l}$  reaction, you should end with up to 100  $\mu\text{l}$  of  100 nanomolar (nM) labeled DNA/RNA.
- 10 Measure  1  $\mu\text{L}$  of each reaction with the black rad counter on shelf to get cpm readings. 