

Jan 01, 2020

DNA Quantification

 In 2 collections

DOI

dx.doi.org/10.17504/protocols.io.4g8gtzw

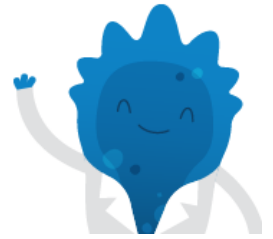
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DOI: dx.doi.org/10.17504/protocols.io.4g8gtzw

External link: <https://www.addgene.org/protocols/dna-quantification/>

Protocol Citation: Addgene The Nonprofit Plasmid Repository 2020. DNA Quantification. **protocols.io**
<https://dx.doi.org/10.17504/protocols.io.4g8gtzw>

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Protocol status: Working

We use this protocol and it's working

Created: June 21, 2019



Last Modified: August 18, 2020

Protocol Integer ID: 24832

Abstract

This protocol is for DNA quantification. To see the full abstract and additional resources, please visit <https://www.addgene.org/protocols/dna-quantification/>.



- 1 Before measuring any samples, be sure to 'blank' the spectrophotometer using the solution the DNA is resuspended in, but with no DNA added. 'Blanking' measures the background inherent to the machine and your solvent.
- 2 If using a NanoDrop to measure your samples, place  1 μL -  2 μL of mini-prepped DNA onto the pedestal.
- 3 Close the lid and click measure, be sure to record the concentration and purity.

Note

Note, purity is measured under the 260/280 column (A good purity ranges from 1.80-2.00).

- 4 Repeat for each sample.

Note

Notes:

- Keep in mind that despite the accuracy of the NanoDrop, if two consecutive samples have significantly different concentrations, it is possible that the difference between them has affected the accuracy of the NanoDrop. It is a good idea to re-zero any spectrophotometer between samples if they are expected to vary significantly in concentration.
- DNA dissolved in water is going to have a greater variance in concentration readings than a DNA sample dissolved in buffer (such as TE). You will get much more accurate and consistent readings from DNA in a buffered solution.