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# DNA extraction from fish tissue using the Wizard® SV 96 Genomic DNA Purification System (Promega)

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**We use this protocol and it's working**

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## Abstract

This protocol describes a method for extracting genomic DNA from fish tissue (e.g., fin tissue) using the Wizard Genomic DNA Purification kit (Promega), adapted from the manufacturers' recommendations. A summary sheet of the DNA extraction procedure is included to support its practical implementation in the laboratory.

## Attachments



Rogissart-et-al-DNA-...

204KB

## Image Attribution

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## Materials

### ■ Samples

- Fish tissue preserved in ethanol (final concentration > 90 % of ethanol) or at -20 °C
- Weight required = 10-15 mg

### ■ Reagents

To clean the workspace

- DNA/RNA-ExitusPlus
- Ethanol (70 %)

For prepare sample:

- Ethanol 96 - 100 %
- Water

For DNA extraction

- DNA extraction kit: **Wizard Genomic DNA Purification kit**
- Ethanol 96 - 100 %, molecular grade to prepare wash buffer
- Nuclease-Free Water

### ■ Materials (excluding solutions preparation)


- Vac-Man® 96 Vacuum Manifold
- Vacuum trap for waste collection
- Vacuum pump with tubing
- 96-well deep well plate
- Adhesive plate sealers
- Bunsen burner
- Chisel and lab pincer
- Balance
- Specific DNA-work station (sterile area equipped with air filtration)
- Horizontal vortexer with microtube holder
- Vortexer
- Stove (to have 55 °C)
- Pipettes: 100-1000 µL; 10-100 µL or Multichannel pipettors: 10-1,000 µL
- 2 trash cans: 1 for liquid and 1 for solid

### ■ Consumables

- Tips with filter:
  - > 1000 µL
  - > 100 µL
- Gloves

## Protocol materials


 Qubit™ dsDNA BR Assay Kit **Thermo Fisher Scientific Catalog #Q32853**

 Wizard® SV 96 Genomic DNA Purification System **Promega Catalog #A2370**

 Proteinase K 20 mg/ml SOLUTION (STABILISEE) **eurobio Catalog #GEXPRK01-I5**




## Troubleshooting

## Safety warnings



 Safety informations of all buffers are available at: [Safety Data Sheets](#)

## Before start

Wear gloves throughout the extraction process.

1. Preheat dry bath or stove to  55 °C
2. Clean the workspace with DNA-ExitusPlus followed by  70 % volume ethanol
3. Unpack  Wizard® SV 96 Genomic DNA Purification System **Promega Catalog #A2370**
4. Check/Prepare solutions

### → Proteinase K Solution

- Ready-to-use solution:  Proteinase K 20 mg/ml SOLUTION (STABILISEE) **eurobio Catalog #GEXPRK01-I5**
- Store at  4 °C





### → Column Wash Solution (CWA):

- Add  95 % volume ethanol to the **CWA** bottle as directed on the bottle label
- Store at  Room temperature

## Prepare sample:

- 1
  - Before start
    - Organize the samples based on the plate map
    - Decontaminate cutting tools: clean with DNA ExitusPlus, rinse with water and cover with ethanol
    - Sterilize cutting tools using bunsen burner
  - Cut approximatively **10–15 mg of tissue**
  - Place tissue in dedicated well of 96-Well Deep Well Plate

### Note

- Decontaminate or replace cutting tools for each sample to prevent cross-contamination
- Cutting the tissue into small pieces may enable more efficient lysis
- Store at  +4 °C for quick use (<  08:00:00 ) or at  -20 °C for further use (>  08:00:00 )

## Lyse sample:


- 2
  - Prepare the lysis mixture containing the following per sample

	Digestion Solution Master Mix
	200 µL Nuclei Lysis Solution
	50 µL 0.5M EDTA (pH 8.0)
	20 µL proteinase K, 20 mg/ml
	5 µL RNase A Solution, 4 mg/ml

Lysis mixture composition

### Note

- Homogenize Digestion Solution Master Mix
- When preparing the mix for multiple samples, include an extra volume equivalent to 1 to 1.5 additional samples per every 8 samples

- Add  275 mL lysis mixture to each sample in the 96-well deep well plate



- Incubate at 55 °C for at least 18:00:00 in a stove

#### Note

- *Make sure the incubator is preheated to 55°C otherwise the lysis time must be increased*
- *Cover the plate with an adhesive plate sealer*
- *To prevent condensation or liquid accumulation on the sealer, place a sheet of aluminum foil on top*
- Overnight incubation is applied to ensure that digestion will be complete and/or to facilitate the organization of extraction.

## Purification of Genomic DNA:

- 3
  - Following incubation, dispense 250 µL of the **Wizard SV Lysis Buffer** into each well of the deep-well plate containing lysate

#### Note

- *Lysate must be warm during processing*
- *Mix each well's contents by pipetting up and down three times to ensure homogeneity*



- Prepare the Vacuum Manifold (see figure on summary sheet)
- Place the Binding Plate in the Vacuum Manifold Base
- Orient the Binding Plate in the Vacuum Manifold with the numerical column headers toward the vacuum port
- Attach the vacuum line to the vacuum port on the Manifold Base
- Transfer the tissue lysates to the wells of the Binding Plate
- Apply vacuum until all of the lysate has passed through the Binding Plate

#### Note

- **Caution:** *Always place a liquid trap (e.g., vacuum flask) between the manifold and the vacuum pump to avoid damaging the pump with aspirated liquid*
- *Vacuum pressure should be approximately 508 mBar (or 50.8 kPa) for efficient processing*
- *To maintain strong vacuum across the wells, cover the plate with adhesive plate sealer*







## Wash & Dry:

- 4
  - Add  900  $\mu\text{L}$  of **Column Wash Solution (CWA)** to each well of the Binding Plate
  - Apply vacuum until the **CWA** passes through the Binding Plate
  - Repeat two more times for a total of **3 washes** with the **CWA**
  - After the wells have emptied, continue to apply vacuum for an additional  00:06:00 to allow the binding matrix to dry
  - Turn off the vacuum
  - Release the vacuum line from the Manifold Base, and snap it into the vacuum port in the Vacuum Manifold Collar
  - Remove the Binding Plate from the Manifold Base
  - Blot by gently tapping onto a clean paper towel to remove residual ethanol and repeat if necessary to remove all residual ethanol

### Note

- A fourth wash can be performed if necessary

## Elute:

- 5
  - Place the **96-Well Deep Well Plate** in the Manifold Bed and position the Vacuum Manifold Collar on top
  - Orient the plate with the numerical column headers toward the vacuum port
  - Position the Binding Plate on top of the Manifold Collar
  - Place the Manifold Collar containing the Binding Plate on top of the 96-Well Deep Well Plate sitting on the manifold bed
  - The Binding Plate tips must be centered on the Deep Well Plate wells, and both plates must be in the same orientation
  - Add  250  $\mu\text{L}$  of  Room temperature **Nuclease-Free Water** to each well of the Binding Plate and incubate for  00:02:00 at  Room temperature
  - Apply vacuum until the **Nuclease-Free Water** passes through the Binding Plate
  - Release the vacuum and remove the Wizard SV 96 Binding Plate
  - Optional: transfer the samples into 1.5 mL tubes






### Safety information

- ***Gently detach the Manifold Collar ensuring the Deep Well Plate stays properly aligned in the Manifold Bed***
- ***In case droplets are visible on the upper edge of the wells, gently tap the plate on the bench to allow them to settle at the bottom***

## DNA storage

6 **DNA is ready** to use immediately or store at:



-  4 °C for use a few days
-  -20 °C for use within a few weeks
-  -70 °C for long-term storage

### Note

*To store the plate, seal it tightly using a plate sealer*

## DNA quantification and DNA quality control:

7 ***NanoDrop measurement:***

- Use  2 µL of **elution buffer** (without DNA) as a blank to calibrate the NanoDrop and correct for background absorbance
- To validate the blank, add  2 µL of **elution buffer** onto the NanoDrop pedestal and measure



## Equipment

NanoDrop™ One UV-Vis Spectrophotometer

NAME

spectrophotometer

TYPE

Thermo Scientific

BRAND

ND-ONE-W


SKU

<https://www.thermofisher.com/order/catalog/product/ND-ONE-W>

LINK

Sample Volume (Metric): Minimum 1µL; Spectral Bandwidth: ≤1.8 nm (FWHM at Hg 254 nm); System Requirements: Windows™ 8.1 and 10, 64 bit; Voltage: 12 V (DC); Wavelength Range: 190–850 nm

SPECIFICATIONS

- To measure DNA concentration of sample, add  2 µL of sample DNA onto the NanoDrop pedestal and measure
  - Note down the A260/280, A260/230 and concentration (ng/µl)
  - or export the data to a USB storage device

## Note

- *Clean the surface between each read*

## 8 Qubit Measurement:

- Ensure the Qubit is calibrated before starting

## Equipment

Invitrogen™ Qubit™ 3 Fluorometer

NAME

Accurately measures DNA, RNA, and protein using the highly sensitive fluorescence-based Qubit quantitation assays

TYPE

Invitrogen™ Q33216

BRAND

Q33216


SKU

<https://www.fishersci.co.uk/shop/products/qubit-3-0-fluorometer/15387293>

LINK

### ■ Use the

☒ Qubit™ dsDNA BR Assay Kit Thermo Fisher Scientific Catalog #Q32853

- Analyze  1 µL of **extracted DNA**
- Note the concentration (ng/µl)