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DNA extraction from dermatophytes using the Qiagen DNEasy™ UltraClean Microbial kit (REF: 12224-50) V.1

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We are still developing and optimizing this protocol

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Disclaimer

Abstract

This protocol describes the steps necessary to extract and purify genomic DNA from dermatophytes (and more specifically from dermatophytes of the genus Trichophyton).

Guidelines

Perform the DNA dosage directly after extraction and not after a freeze/thaw cycle.
Be careful when preparing the medium : work under sterile conditions as much as possible to avoid contaminating the liquid medium.

Materials

Qiagen DNEasy UltraClean Microbial kit (REF: 12224-50)
Sabouraud Dextrose Broth (REF : Merck S3306)

Protocol materials

Sabouraud dextrose broth **Merck MilliporeSigma (Sigma-Aldrich) Catalog #**S3306

MilliQ water

Troubleshooting

Medium preparation

35m

1 Dissolve 30 g of

Sabouraud dextrose broth **Merck MilliporeSigma (Sigma-Aldrich) Catalog #**S3306

in 1 L

of MilliQ water and let mix on the heated magnetic stirrer for 00:05:00

(temperature and mixing speed knob at mid-step).

5m

2 Cover the flask with glass wool and aluminium foil. Autoclave it at 121 °C 121 °C for 00:30:00 .

30m

Cultivation of the strains

3 After allowing to cool, transfer 25 mL of this medium into a tube. Label the tube with the strain number.

4 Using a sterile swab (or a sterile inoculation loop), gently collect the primary culture and dip the swab (or the sterile inoculation loop) into the tube containing the culture medium (prepared in the previous step). Close the tube halfway to allow gas exchange.

5 Allow to grow in the incubator at 30 °C until a sufficient flocculate is formed (requires at least 96 hours). Incubation time varies from strain to strain but flocculate should be visible after 5 days. If this is not the case, repeat the cultivation step.

Preliminary steps

3m

6 Using a Pasteur pipette, carefully remove the flocculate from the tube containing the previously cultured dermatophyte strain. Transfer this flocculate to a PowerBead tube containing glass beads, let's call it primary tube. Add 300 µL of PowerBead solution and 50 µL of SL solution to this tube.

7 Cool this tube to -196 °C in liquid nitrogen for 00:01:00. Then, heat this tube in a water bath at 56 °C for 00:01:00. Finally, run this tube through the cell disruptor at maximum speed for 00:01:00. This constitutes 1 cycle of 3 steps. You must repeat this cycle 3 times. The recovered mixture is referred to as primary lysate in the following steps. 3m

DNA extraction 6m 50s

8 Centrifuge the tube at 10000 x g for 00:00:30 at room temperature. Gently transfer the supernatant to a clean collection tube (provided in the kit) and discard the PowerBead tube. 30s

9 Add 100 µL of IRS solution to the supernatant and vortex for 00:00:10. Incubate at 4 °C for 00:05:00. 5m 10s

10 After that, centrifuge the tube at 10000 x g for 00:01:00 at room temperature. Gently transfer the supernatant to a new collection tube and discard the tube containing the pellet. 1m

11 Add 900 µL of SB solution to the tube containing the supernatant from the previous step and vortex for 00:00:10. Load approximately 700 µL of this suspension into a silica membrane chromatography column (provided in the kit). 10s

DNA purification 4m

12 Centrifuge the column at 10000 x g for 00:00:30 at room temperature. Keep the column and discard the flows-through. Repeat until the entire volume from step 11 is loaded into the column. 30s

13 Add 300 µL of CB solution into the column and centrifuge it at 10000 x g for 00:00:30 at room temperature. Keep the column and discard the flows-through. 30s

14 Centrifuge the column alone (empty) to remove the last residues of CB solution. The conditions are identical to the previous step : 10000 x g for 00:00:30 at room temperature. 30s

15 Place the column in a new collection tube. Add 50 µL of EB solution to the center of the silica membrane. Let stand for 00:01:00 at room temperature. Let stand for 00:01:00 at room temperature and then centrifuge the column at 10000 x g for 00:00:30 to elute the DNA. 2m 30s

16 Discard the column and keep the flows-though which is the purified DNA. Store DNA at -80 °C to ensure stability.

Spectrophotometric dosage

17 To determine the purity and concentration of the DNA, a NanoDrop dosage was performed. For this purpose, a negative control was prepared beforehand. This control will have undergone all the extraction steps but will not contain any material from dermatophytes.

18 Launch the computer program and select the "nucleic acid" mode. Make sure the sample deposit spot is clean and dry. If necessary, clean it with the wipes provided for this purpose. Then drop 2 µL of the negative control and click on the "blank" box.

19 Proceed in the same way to measure the sample containing the DNA, but click on "measure" instead of "blank". There is no need to redo a blank between measurements.