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🌐 DNA Bisulfite conversion, using Zymo EZ-96 DNA Methylation-Lightning MagPrep Kit and Integra VIAFLO 96, for processing on Illumina methylation arrays

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We use this protocol and it's working

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Abstract

The Genetics Core of the Edinburgh Clinical Research Facility provides DNA methylation analysis services using optimized automated protocols. This protocol details a high-throughput DNA bisulfite conversion method that uses Zymo EZ-96 DNA Methylation-Lightning MagPrep Kits and automated using the Integra VIAFLO 96 platform. The method is used as the first step in analysing DNA samples on Illumina Infinium Methylation Screening Arrays-48. This protocol allows 192 samples to be processed together.

Guidelines

This protocol details a high-throughput DNA bisulfite conversion method that uses Zymo EZ-96 DNA Methylation-Lightning MagPrep Kits and automated using the Integra VIAFLO 96 platform. The method is used as the first step in analysing DNA samples on Illumina Methylation Screening Arrays.

This protocol allows 2× 96-well plates (192 samples) to be processed together.

Materials

Zymo Research EZ-96 DNA Methylation-Lightning MagPrep Kit (D5046)

Integra VIAFLO 96 electronic pipette

Integra VIAFLO 96 Racks Sterile 300 µl (6434)

Integra VIAFLO 96 Racks Sterile Filter 300 µl (6435)

ThermoMixer e.g. Eppendorf ThermoMixer C (2231001127)

PCR Thermocycler e.g. Eppendorf Mastercycler X50 (2231001152)

Zymo Research ZR-96 Magnetic Stand (P1005)

Troubleshooting

Safety warnings

- ⚠ Step 19 is crucial. Samples need to be incubated for at least 15 mins and no longer than 25 mins. This limits the throughput of plates per operator.



Buffer Preparation

- 1 Add 288 mL of 95–100% ethanol to the 72 mL M-Wash Buffer concentrate. Mix well.
- 2 Label the buffer with preparation date and initials.

Sample Conversion

- 3 Transfer 20 μ L of 50ng/ μ L (500ng) DNA into each well of the Conversion Plate.
- 4 Add 130 μ L of Conversion Reagent (pre-mixed and light-sensitive). Mix thoroughly.
- 5 Seal with foil seal, centrifuge at 280 g for 30 seconds.
- 6 Place plate into the Eppendorf Mastercycler with settings:
98°C for 8 minutes
54°C for 60 minutes
Hold at 4°C (up to 20 hours)

Preparation of Collection Plate During Incubation

- 7 Add 600 μ L of M-Binding Buffer and 10 μ L of EZ-Methylation MagPrep Beads to each well of a fresh Collection Plate.
- 8 Keep beads suspended by gently vortexing or pipetting as needed.

Transfer to Collection Plate

- 9 Spin conversion plate at 280g for 30 seconds
- 10 Using VIAFLO 96 mix programme (aspirate 170 μ L at Asp speed = 1 and Mix speed = 8) and 300 μ L **sterile filter** tips, transfer entire sample volume from Conversion Plate to



corresponding wells of Collection Plate.

- 11 In collection plate, use VIAFLO 96's mix function: mix 300 μ L $\times$ 6 cycles to ensure bead binding. Dispose of tips.
- 12 Incubate at room temperature for 5 minutes.

Bead Separation

- 13 Place Collection Plate onto the ZR-96 Magnetic Stand for 5 minutes to pellet the beads.
- 14 Remove supernatant carefully using VIAFLO 96 at lowest speed setting. (Aspirate 300 μ L at Asp speed = 1 and Disp. speed = 3) using 300 μ L **sterile tips** (these do not need to be filter tips). Dispose tips

Wash Step

- 15 Remove plate from magnetic stand.
- 16 Add 400 μ L of M-Wash Buffer, mix for 30 seconds at 1100 rpm on Eppendorf ThermoMixer C.
- 17 Return plate to magnetic stand for 3 minutes, remove supernatant.

Desulphonation

- 18 Add 200 μ L of L-Desulphonation Buffer, mix on ThermoMixer for 30 seconds at 1100 rpm.
- 19 Incubate at room temperature for 15 minutes.
- 20 Place on magnetic stand for 3 minutes, remove supernatant.



- 21 Note: Take time for handling/re-suspension into account for the total incubation time. Adjust time as necessary to ensure that no sample remains in the L-Desulphonation Buffer for more than 20–25 minutes. This is particularly important when processing two plates together.

Final Washes and Elution

- 22 Add 400 μ L of M-Wash Buffer, mix for 30 seconds at 1100 rpm, return to magnetic stand for 3 minutes, and discard supernatant.
- 23 Repeat the wash step once more (total: 2 washes).
- 24 Note: Remove as much buffer as possible after final wash to aid in the drying of the beads.
- 25 After second wash, dry beads by heating the plate at 55°C for 25 minutes on the ThermoMixer.

DNA Elution

- 26 Add 25 μ L of Elution Buffer. Mix for 30 seconds at 1300 rpm on the ThermoMixer.
- 27 Heat the elution at 55°C for 4 minutes then transfer the plate to the magnetic stand for 1 minute. Remove the supernatant and transfer to a clean Elution Plate using VIAFLO 96 pipette function (Aspirate 30ul at Asp speed = 1 and Disp. speed = 3) and 300 μ L **sterile filter tips**.
- 28 Eluted DNA is now ready for downstream analysis or can be stored at -20°C.



Protocol references

Zymo EZ-96 DNA Methylation-Lightning MagPrep Kit Protocol Ver.2.1.1

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