

Aug 13, 2019

Version 1

# O Detection of Klebsiella pneumoniae and closely related species by real-time PCR with the ZKIR system V.1

dx.doi.org/10.17504/protocols.io.6gvhbw6



Elodie Barbier<sup>1</sup>, Carla Rodrigues<sup>2</sup>, Geraldine Depret<sup>1</sup>, Virginie Passet<sup>2</sup>, Laurent Gal<sup>1</sup>, Pascal Piveteau<sup>1</sup>, Sylvain Brisse<sup>2</sup>

<sup>1</sup>Agroécologie, AgroSup Dijon, CNRS, Univ. Bourgogne, INRA, Univ. Bourgogne Franche-Comté, Dijon, France; <sup>2</sup>Institut Pasteur, Biodiversity and Epidemiology of Bacterial Pathogens, Paris, France

Klebsiella Research and ...



#### **Elodie Barbier**

Université de Bourgogne Franche-Comté - INRAe Dijon - France

### Create & collaborate more with a free account

Edit and publish protocols, collaborate in communities, share insights through comments, and track progress with run records.

Create free account

OPEN ACCESS



DOI: https://dx.doi.org/10.17504/protocols.io.6gvhbw6



**Protocol Citation:** Elodie Barbier, Carla Rodrigues, Geraldine Depret, Virginie Passet, Laurent Gal, Pascal Piveteau, Sylvain Brisse 2019. Detection of Klebsiella pneumoniae and closely related species by real-time PCR with the ZKIR system.

protocols.io https://dx.doi.org/10.17504/protocols.io.6gvhbw6

#### **Manuscript citation:**

Unpublished yet

**License:** This is an open access protocol distributed under the terms of the **Creative Commons Attribution License**, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

Protocol status: Other

Error in the protocol, see version 3

Created: August 13, 2019

Last Modified: August 13, 2019

Protocol Integer ID: 26869

**Keywords:** Klebsiella, phylogroup, ZKIR qPCR, detection, screening, klebsiella pneumoniae, kp ecology, phylogroups kp1, research on kp ecology, phylogroups kp1 to kp7, single bacterium, specific for kp, pneumoniae sensu, time pcr with the zkir system klebsiella, time pcr assay, virulent strain, phylogenetic breadth, human infection, strain, related species, kp1, kp in complex matrice, kp

### Abstract

*Klebsiella pneumoniae* (Kp) is of growing public health concern due to the emergence of multidrug-resistant and virulent strains. Taxonomically, Kp includes seven phylogroups, with Kp1 (*K. pneumoniae sensu stricto*) being medically prominent. Kp can be present in environmental sources such as soils and vegetation, which could act as reservoirs of animal and human infections. However, the current lack of screening methods to detect Kp in complex matrices limits research on Kp ecology.

We designed a novel SYBR green real-time PCR assay, named the ZKIR assay, that targets *Klebsiella pneumoniae* and closely related species (phylogroups Kp1 to Kp7).

Based on 48 Kp representing its phylogenetic breadth, and on 88 non-Kp strains, the ZKIR assay detected all Kp and was totally specific for Kp, as no false positive was found.

We also tested this method on spiked soil microcosms after a 24 h enrichment step (in Lysogeny Broth supplemented with ampicillin 10 mg/l) and a short sample treatment. We showed that this procedure was sensitive enough to detect one single bacterium in 5 g of soil.

#### **Attachments**



Detection of Klebsie...

143KB



### **Materials**

#### **MATERIALS**

X Takyon ROX SYBR MasterMix 2X Blue dTTP

X T4 bacteriophage Gene 32 product MP Biomedicals Catalog #SKU 11TGP32100

#### Primers sequences:

ZKIR F: 5' CTAAAACCGCCATGTCCGAT 3' ZKIR R: 5' TTCCGAAAATGAGACACTTAGA 3'

Primers were synthetized by Eurogentec.

## **Troubleshooting**

#### Before start

Takyon™ qPCR Kits for SYBR® assays containing ROX passive reference is used on the following thermocyclers: ABI Prism® 5700, ABI Prism® 7000, ABI Prism® 7300, ABI Prism® 7700, ABI Prism® 7900 & FAST 7900, ABI Step One & Step One Plus.

For other thermocyclers, see <a href="https://secure.eurogentec.com/egt/files/FileBrowse/Brochures/PCR%20-">https://secure.eurogentec.com/egt/files/FileBrowse/Brochures/PCR%20-</a> %20gPCR/Eurogentec-pcr-gpcr.pdf



### 1 Sample preparation

### 1.1 Sample enrichment step:

Soil samples (10 g) are enriched in 90 ml of LB (Lysogeny Broth: 5 g yeast extract, 5 g sodium chloride, 10 g tryptone for 1 liter) supplemented with ampicillin 10 mg/l for 24 h at 30°C.

Food samples (25g) such as salad and chicken are enriched in 225 ml of BPW (Buffered Peptone Water) for 24 h at 37°C.

### 1.2 Sample treatment step:

 $500 \mu l$  of enrichment is centrifuged 5 min at 5 800 g and washed twice with sterile water before boiling for 10 min. Boiled suspensions are then diluted at 1/10 and 1/100 for qPCR. Crude and diluted DNAs are tested with the ZKIR assay.

2 qPCR mix preparation (final volume 20  $\mu$ l)

Mix reag ents	Volu me per well (μΙ)	Fina I con cent ratio n				
Taky on™ ROX SYB R® Mast erMi x 2X	10					
Forw ard (3 µM)	2	300 nM				
Reve rse (3 µM)	2	300 nM				
T4 gp3 2 (opti onal )	0.5	12.5 μg/ ml				



PCR grad e	QS 17.5 μΙ	
wate r	μ	
DNA tem plate	2.5	

3

### Thermocycler settings

	Temperature (°C)	Time (min.)	
Holding stage	95.0	03:00	Enzyme activation
Cycling stage	95.0	00:10	Denaturation
	60.0	01:00	Annealing (data collection)
Melt Curve stage	95.0	00:15	Dissociation stage Program Step and Hold T° increment +0.3°C
	60.0	01:00	
	95.0	00:15	

Thermocycler settings

#### 4 **Positive and negative controls:**

Negative control: PCR grade water

Positive control: *Kp*1 DNA diluted to 1 ng per μl (Strain ATCC13883T)

5 Melt curve peak established using the ZKIR assay with serial dilutions of K. pneumoniae ATCC13883T. Melting temperature was around 80.2 °C.

