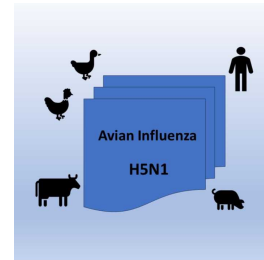


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Detection of avian Influenza Virus H5N1 with a Real time PCR kit

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We use this protocol and it's working

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Disclaimer

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Abstract

Avian Influenza virus H5N1 has caused a number of outbreaks in human beings in recent years. Recently it has caused outbreak in human and cows in USA. Therefore it is essential to detect the presence of this virus as early as possible in order to monitor the presence of this virus, hence we have developed a number of ready to use PCR kits to detect this virus. Highly pathogenic strains are responsible for such outbreak. It spreads also in poultry with very fatal consequences. The kit double- and triple-checks the presence of this virus, therefore it makes possible to detect the H5N1 strains circulating in different countries worldwide, moreover mutated versions are likely to be detected with double and triple check method. The cross reaction panel has done with related and non related pathogens like Influenza virus strains: H3N2, H1N1, H7N3, H1N1 (California strain); RSV, Enterovirus, CMV, Influenza B, Mycoplasma etc.



Materials

FR006 - FluHunter: Avian Influenza Virus H5N1 (Realtime PCR-Kit) CE

-UDI-DI: 04262420430102

Compositions of the Kit:

- Tube A (2 tubes): **2 × 385µl**
- Tube B (2 tubes): **2 × 550µl**
- Tube Y (1 tube): **110µl**
- Positive (+Ve) Control (tube D1) (1 tube): **40µl**
- Negative (-Ve) Control (tube D2) (1 tube): **60µl**

- Realtime PCR thermocycler
- Laboratory centrifuge
- 96 well microplates for PCR or Microtubes (0.2ml)
- Sterile Pipette-tips with filter (1-10µl & 10-100µl)
- Pipettes (quality pipettes)
- Vortexer
- RNA isolation kit: **Genekam Biotechnology AG, Germany or equivalent**

Safety warnings

Precautions:

- It is only for in vitro use.
 - It must be used through trained persons.
 - It should not be used after expiry date.
 - During PCR, one should work very cleanly.
- All works must be done in different parts of laboratory.
- **NEVER STORE THIS KIT WITH FOOD, VACCINE OR BACTERIAL CULTURES.**
 - Decontaminate the instruments regularly (once a week).
 - To dispose of the kit and its contents, add 70% ethanol to the contents of the kit and then dispose of it.



Before start

1. User must read the manual before use.

2. Important Hint:

Real time PCR is based on fluorogenic dyes. **There are 2 probes with 2 different** dyes in this kit hence you have to program them in your machine:

- First the probe, namely Carboxy-fluorescein (reporter) and 6-Carboxy tetramethyl rhodamine (quencher). **The results will be shown as Ct -Values.** Up to 40 Ct should be taken positive. Value between 40-45 Ct should be taken as marginal positive (doubtful).

- Second the probe, namely HEX (reporter, it is available as VIC in some machines) and 6-Carboxy tetramethyl rhodamine (quencher). **The results will be shown as Ct -Values.** Up to 40 Ct should be taken positive. Value between 40-45 Ct should be taken as marginal positive (doubtful).

If any of the both channels or both are positive, it is to be considered positive.

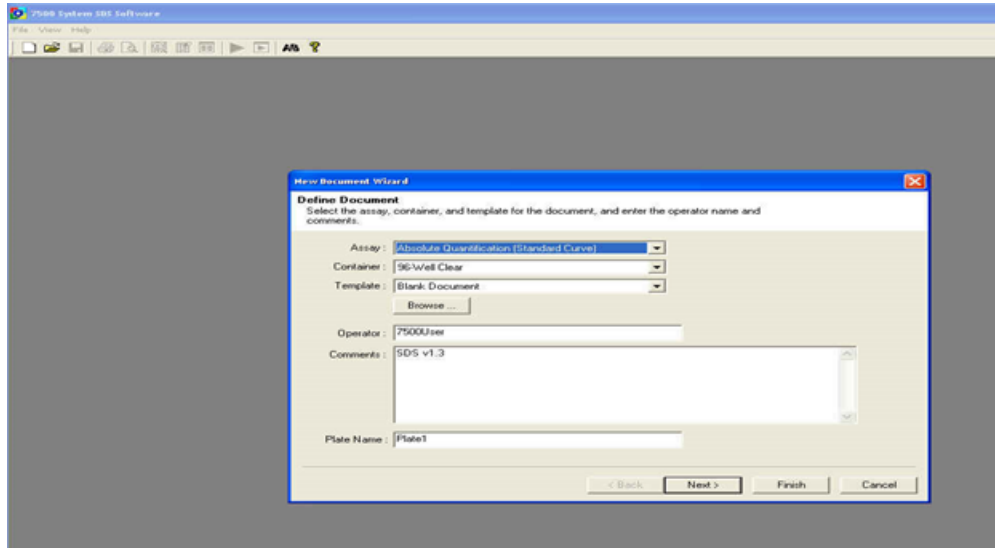


- 1 Thaw **one tube** each: A, B, Y, D1 and D2. After thaw, put the tubes at 4°C (as it is better). If the kit is not in use, store them at -20 ° C.
- 2 Mark your microtubes with a sample number, +ve Control and -ve Control. Otherwise use a 96 well microwell plate instead of tubes.
- 3 Add 7µl of Tube A to each tube.
- 4 Add 10µl of B to each micro tube. Avoid touching the wall of the microtubes.
- 5 Add 1µl of Y to each tube (Avoid touching the wall of the microtubes).
TIP: User can calculate the total requirement of chemicals needed. User can mix 7µl of A + 10µl of B + 1µl Y together in one tube for one reaction, but to have 10 reactions, there will be total volume of 180µl (70µl Tube A, 100µl B and 10µl Y). From this, 18µl can be distributed into each tube. This step saves time and hardware.
- 6 Add 2µl of your RNA with sterile pipette-tip with filter to each micro tube according to your label except +Ve and -Ve (Avoid touching the wall). **Use a new pipette tip for each sample!**
- 7 Use new pipette tip with filter. Add 2µl of Tube D1. This is the positive control supplied with our kit.
- 8 Use a new pipette tip. Add 2µl of -Ve (tube D2) to -Ve Control (Do not touch the wall).
- 9 Centrifuge all tubes at 8000 rpm for 5 sec. (This is not necessary, but it is better).
- 10 Run the program of your thermocycler as follows: Check if everything is added correctly, as the volume of each microtube must be almost the same.
- 11 **Now enter quencher and reporter dye to setup your software (see FAQ) and run the following program (2h30min):**
 1. 60 minutes at 42°C
 - 10 minutes at 70°C

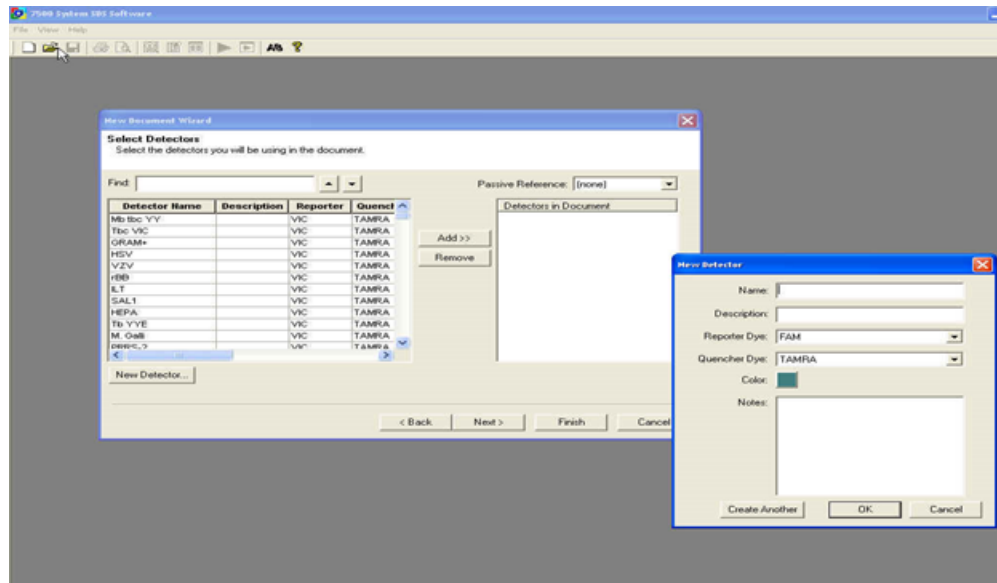
2. 15 seconds at 95°C x 45 cycles
60 seconds at 60°C

12 Example of setting up a real time machine: Instrument Setup Instructions for real time PCR (RT-PCR System: ABI 7500)

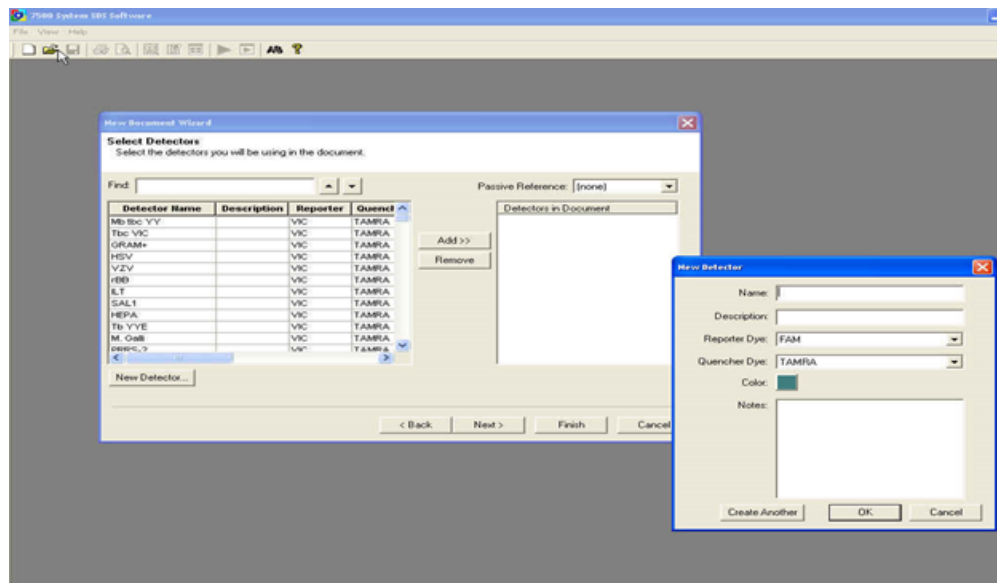
Open SDS Software -**File New** -Select: Absolute Quantification assay -Insert name - Click: **Next**



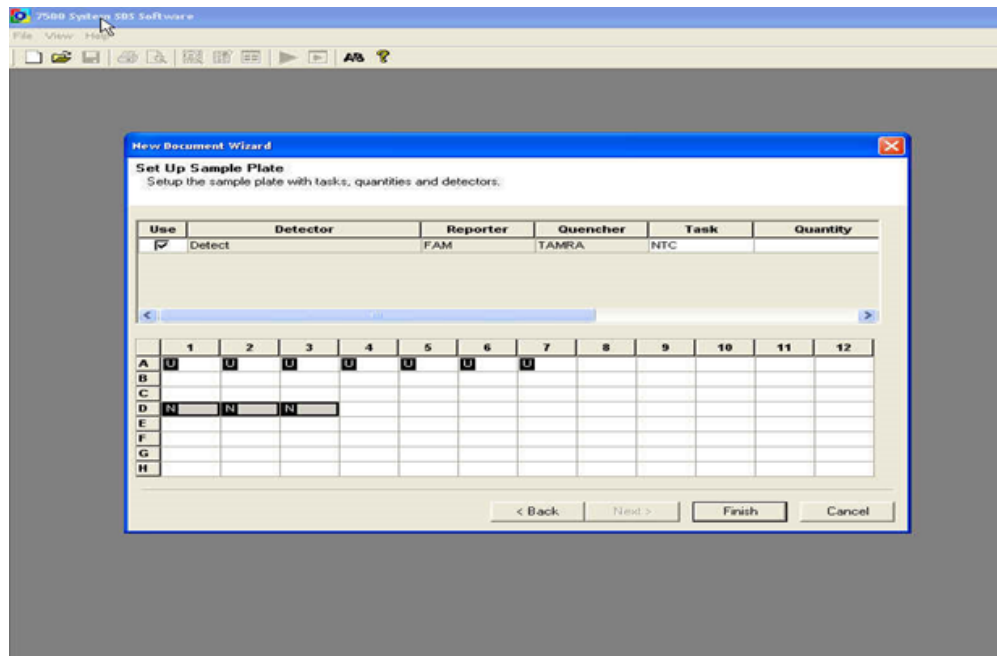
- 13 Create a **New Detector 1**: Select **Reporter** and **Quencher**-Dye: (FAM-TAMRA) -Click: Add
-Select: Passive Reference-none -Click: **Next**



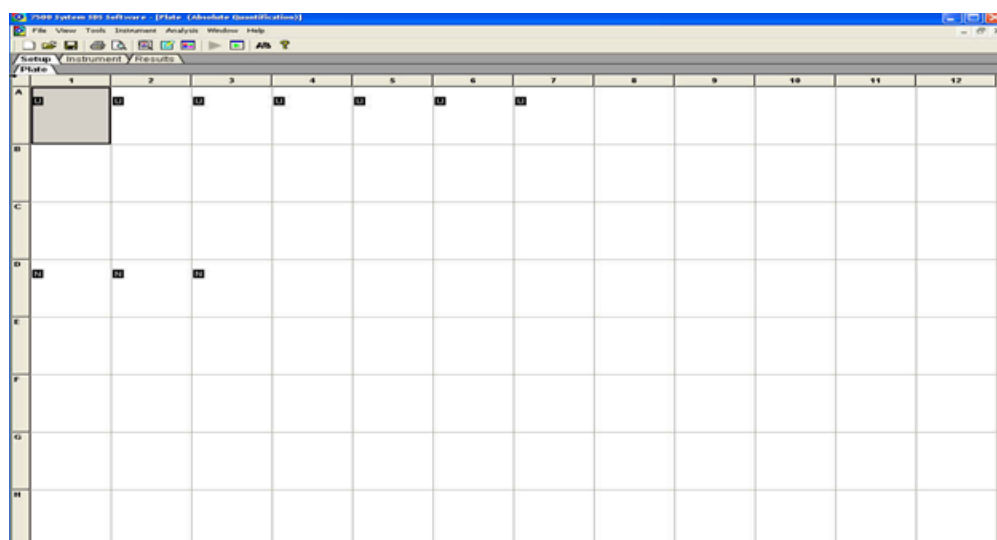
- 14 Create a **New Detector 2** (VIC-TAMRA) -Click: Add -Select: Passive Reference-none - Click: **Next**



- 15 Insert Well positions: **Unknown** – or -**NTC** (No Template Control) - or– **Standard** -Click: Finish: The Plate Document will open.



16 Write your Array: Name of Samples for each well or tube.



17 Select Instrument Tab – Insert the **Program** into the Thermal Profile **as mentioned in point 11** - Change Sample Volume (20µl) -Click: File - **“Save as”** to save the Setup -Click: **“Start”** to begin the PCR run: Shortly the run time will appear on the screen.

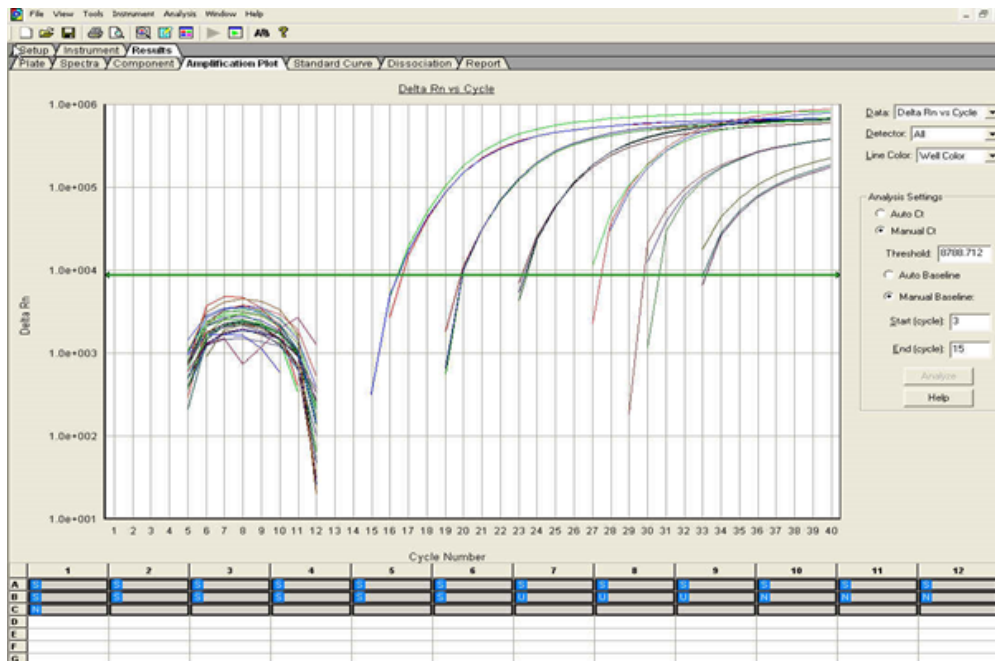
Before starting the PCR program, check that the plate or tubes are properly sealed. **The wells of the well plate or microtubes must be in contact with metal block** (important!). There should be no air or loose contact with metal block of thermocycler. In case of 96-well plate, it should be sealed with adhesive cover. Now run your PCR.

Once the PCR is finished, a small dialog box will appear: "The run has been completed successfully"

Click: OK. Now you can take out the microtubes or well plate.

18 **STEP B**

Click the **Result**-Tab and the **Amplification**-Tab: Place the Threshold line above the background, then select: "Analyse" (see Figure below). Calculate the threshold cycle (Ct) Value for each well. There should be no signal in the negative control. Successful positive controls or positive samples must give a curve in the software graphics.



19 The Report page will give you the Ct-values and Standard deviation for each well.



Well	Sample Name	Detector	Task	Ct	StdDev Ct	Qty	Mean Qty	StdDev Qty	Filtered	Trn
A1	H5N1 real	H5N1 Neu_2	Standard	9.95	2.141	1.00				
A1	H5N1 real	H5N1 Neu	Standard	13.57	0.108	1.00				
A2	H5N1 real	H5N1 Neu_2	Standard	4.93	2.141	1.00				
A2	H5N1 real	H5N1 Neu	Standard	13.43	0.108	1.00				
A3	H5N1 real	H5N1 Neu_2	Standard	8.26	2.141	1.00				
A3	H5N1 real	H5N1 Neu	Standard	13.64	0.108	1.00				
A4	1:10	H5N1 Neu_2	Standard	15.38	5.780	1.00e-001				
A4	1:10	H5N1 Neu	Standard	12.32	1.083	1.00e-001				
A5	1:10	H5N1 Neu_2	Standard	6.37	5.780	1.00e-001				
A5	1:10	H5N1 Neu	Standard	14.23	1.083	1.00e-001				
A6	1:10	H5N1 Neu_2	Standard	4.57	5.780	1.00e-001				
A6	1:10	H5N1 Neu	Standard	12.39	1.083	1.00e-001				
A7	1:100	H5N1 Neu_2	Standard	5.64	1.559	1.00e-002				
A7	1:100	H5N1 Neu	Standard	17.10	1.742	1.00e-002				
A8	1:100	H5N1 Neu_2	Standard	5.76	1.559	1.00e-002				
A8	1:100	H5N1 Neu	Standard	17.24	1.742	1.00e-002				
A9	1:100	H5N1 Neu_2	Standard	9.40	1.559	1.00e-002				
A9	1:100	H5N1 Neu	Standard	20.19	1.742	1.00e-002				
B1	1:1000	H5N1 Neu_2	Standard	10.06	3.134	1.00e-003				
B1	1:1000	H5N1 Neu	Standard	21.84	1.226	1.00e-003				
B2	1:1000	H5N1 Neu_2	Standard	14.95	3.134	1.00e-003				
B2	1:1000	H5N1 Neu	Standard	20.08	1.226	1.00e-003				
B3	1:1000	H5N1 Neu_2	Standard	15.80	3.134	1.00e-003				
B3	1:1000	H5N1 Neu	Standard	19.49	1.226	1.00e-003				
B4	1:10000	H5N1 Neu_2	Standard	13.15	3.183	1.00e-004				
B4	1:10000	H5N1 Neu	Standard	17.77	0.604	1.00e-004				
B5	1:10000	H5N1 Neu_2	Standard	7.86	3.183	1.00e-004				
B5	1:10000	H5N1 Neu	Standard	17.92	0.604	1.00e-004				
B6	1:10000	H5N1 Neu_2	Standard	7.63	3.183	1.00e-004				
B6	1:10000	H5N1 Neu	Standard	19.88	0.604	1.00e-004				
B7	1:100.000	H5N1 Neu_2	Standard	19.25	7.139	1.00e-005				
B7	1:100.000	H5N1 Neu	Standard	21.85	6.203	1.00e-005				
B8	1:100.000	H5N1 Neu_2	Standard	9.76	7.139	1.00e-005				
B8	1:100.000	H5N1 Neu	Standard	17.40	6.203	1.00e-005				
B9	1:100.000	H5N1 Neu_2	Standard	5.26	7.139	1.00e-005				
B9	1:100.000	H5N1 Neu	Standard	23.65	6.203	1.00e-005				

Summary section below the table:

	1	2	3	4	5	6	7	8	9	10	11	12
A	1	1	1	1	1	1	1	1	1	1	1	1
B	1	1	1	1	1	1	1	1	1	1	1	1
C												
D												
E												
F												
G												
H	1	1	1	1	1	1	1	1	1	1	1	1

If either channel is positive, the result should be considered positive regardless of whether the internal control is negative or positive. If the pathogen-specific test and the internal control are negative, the test should be repeated with the same sample or a new isolation. If the internal control is positive and the H5N1 test is negative, the test should be scored negative.