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CycloneSEQ based Pore-C Library Preparation and Sequencing Protocol

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Protocol status: Working

We use this protocol and it's working

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Abstract

This protocol is designed for Pore-C library preparation and sequencing of extracted animal DNA on the CycloneSEQ sequencing platform.

Troubleshooting



Tissue Fixation & Crosslinking

- 1 Homogenization & Washing:
 - Using 100 mg of fresh tissue per sample, homogenize in 2 mL 1x PBS using a sterile pestle.
 - Centrifuge at 2,000xg for 5min at 4°C. Discard supernatant.
- 2 Fixation:
 - Resuspend pellet in 10 mL 1% formaldehyde in 1x PBS
 - Incubate 10 min at RT with gentle rotation.
- 3 Quenching
 - Add 527 μ L 2.5 M glycine.
 - Incubate 5 min at RT, then incubate 10 min on ice.
 - Centrifuge at 2,000xg for 10 min at 4°C. Discard the supernatant.

Nuclei isolation & Digestion

- 4 Lysis:
 - Resuspend pellet in 550 μ L ice-cold lysis buffer
(10 mM Tris-HCl pH 8.0, 10 mM NaCl, 0.2% Igepal-CA630, 50 μ L of protease inhibitor cocktail)
 - Incubate 15 min on ice.
 - Centrifuge at 2,000xg for 10 min at 4°C. Discard supernatant.
- 5 Digestion Prep:
 - Resuspend pellet in 200 μ L ice-cold 1.5 \times digestion buffer.
 - Centrifuge at 2,000 \times g for 10 min at 4°C.
 - Resuspend pellet in 300 μ L 1.5 \times digestion buffer.
 - Add 33.5 μ L 1% SDS.
- 6 Digestion:
 - Incubate in thermomixer: 65°C, 300 rpm, 10 min.
 - Add 37.5 μ L 10% Triton X-100 (to quench SDS).
 - Add 45 μ L NlaIII (10 U/ μ L) + 34 μ L nuclease-free water.
 - Rotate 18 hr at 37°C.
 - Heat-inactivate: 65°C for 20 min.

Proximity Ligation

- 7 Ligation Master Mix:
Prepare 550 μ L mix per sample:
 - 100 μ L 10 \times T4 DNA ligase buffer (with ATP)
 - 10 μ L BSA (10 mg/mL)



- 50 μ L T4 DNA ligase (400 U/ μ L)
- 390 μ L nuclease-free water

8 Ligation:

- Add master mix to sample.
- Rotate 5 hr at RT.

Crosslink Reversal & DNA Purification

9 Protein Digestion:

- Add master mix:
 - (1) 100 μ L 10% SDS
 - (2) 500 μ L 20% Tween-20
 - (3) 100 μ L Proteinase K (20 mg/mL)
 - (4) 300 μ L nuclease-free water
- Incubate 56°C for 18 hr.

10 DNA Extraction:

- Add equal volume phenol:chloroform:isoamyl alcohol (25:24:1).
- Centrifuge 12,000 \times g for 15 min at 4°C.
- Transfer aqueous phase to new tube.

11 Precipitation:

- Add 0.7 \times volume isopropanol.
- Incubate -20°C for 1 hr.
- Centrifuge 12,000 \times g for 30 min at 4°C.
- Wash pellet with 70% ethanol. Air-dry.

Library Construction

12 Size Selection & Bead Cleanup:

- Resuspend DNA in 50 μ L nuclease-free water.
- Perform size selection >1.5 kb using DNA Clean Beads per manufacturer's instructions.

13 CycloneSEQ Library Prep:

- Construct library using CycloneSEQ Long-Read WGS Kit (refer to separate protocol).
- Quantify with Qubit fluorometer.

Sequencing

14 Run Sequencing:

- Load library onto CycloneSEQ WuTong02 platform.
- Follow manufacturer's run parameters.