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## Culture and transfection of HEK293T cells

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**We use this protocol and it's working**

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## Abstract

This protocol describes a standard procedure culturing and transfecting HEK293T cells

### Protocol overview:

A. Culturing HEK293T cells

B. Transfection of HEK293T cells with Lipofectamine 2000



## Materials

|  | A                                    | B             | C         |
|--|--------------------------------------|---------------|-----------|
|  | Item                                 | Vendor        | Catalog # |
|  | DMEM, high glucose                   | Thermo Fisher | 11965118  |
|  | DPBS w/o Calcium and magnesium       | Corning       | MT21031CV |
|  | Fetal Bovine Serum (FBS)             | Corning       | 35-011-CV |
|  | L-Glutamine                          | Sigma         | G8540     |
|  | MEM Non-Essential Amino Acids (100X) | Thermo Fisher | 11140050  |
|  | 0.25% Trypsin with EDTA              | Thermo Fisher | 25200114  |
|  | Opti-MEM                             | Thermo Fisher | 31985062  |
|  | Sodium Pyruvate 100 mM               | Thermo Fisher | 11360070  |
|  | Lipofectamine 2000                   | Thermo Fisher | 11668019  |



## A. Culturing HEK293T cells

1m

- 1 HEK293T cells are cultured in HEK293T medium in 10 cm dishes.

### 1.1 HEK293T Medium

| A                                    | B      |
|--------------------------------------|--------|
| DMEM, high glucose                   | 385 ml |
| Fetal Bovine Serum (FBS)             | 50 ml  |
| L-Glutamine (100X)                   | 5 ml   |
| MEM Non-Essential Amino Acids (100X) | 5 ml   |
| Sodium Pyruvate 100 mM               | 5 ml   |

Final volume: 500 mL

- 2 Passage cells when the culture reaches 80% confluency.
- 3 Remove medium
- 4 Wash once with 5 ml DPBS
- 5 Add 1 ml 0.25% Trypsin with EDTA, tilt and shake the dish so that the Trypsin covers the entire dish.
- 6 Incubate at 37 °C for 00:01:00
- 7 Add 5 ml fresh HEK293T medium to inactivate Trypsin
- 8 Pipet 10 times to dissociate the cells and mix

1m



- 9 Transfer 1 ml of cell suspension to a new dish pre-added with 9 ml HEK293T medium. Shake to mix well. This is a 1:6 splitting.
- 10 HEK293T cells usually need to be passaged every 2 days.

## B. Transfection of HEK293T cells with lipofectamine 2000

45m

- 11 One day before transfection, dissociate HEK293T cells with Trypsin as described above
- 12 Seed 250,000 cells/1 well of 12-well plate
- 13 Change to fresh medium 1 h before transfection
- 14 Label two micro centrifuge tubes as I and II
- 15 In micro centrifuge tube I, add 125 µl Opti-MEM and 6 µl Lipofectamine 2000. Mix by gently pipetting 3 times. Incubate at Room temperature for 00:05:00 . 5m
- 16 While waiting, in micro centrifuge tube II, add 125 µl Opti-MEM and 250 ng total plasmid. Mix by pipetting.
- 17 Mix tube I and II by gently pipetting 3 times. Incubate at Room temperature for 00:20:00 . 20m
- 18 Mix one time by gently pipetting. Transfer all transfection reagents into one well of a 12-well plate, drop-wise.
- 19 After adding the reagent to all wells, shake the plate to mix.
- 20 Culture in 37 °C incubator Overnight . 20m



- 21 Change to fresh medium and culture for another 2 days. Collect samples or passage once if longer culturing is needed.