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CTAB DNA Extraction for genotyping

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External link: [http://mimubase.org/FTP/Protocols/DNA_extraction/CTAB%20DNA%20Extraction%20\(Regular\).pdf](http://mimubase.org/FTP/Protocols/DNA_extraction/CTAB%20DNA%20Extraction%20(Regular).pdf)

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Protocol status: Working

We use this protocol and it's working

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Attachments



CTAB DNA Extraction ...

26KB

Materials

MATERIALS

 Liquid Nitrogen

 70% Ethanol

 CTAB DNA Extraction buffer

 Chloroform: IsoAmyl Alcohol (24:1)

 7.5M Ammonium acetate

 100% Ethanol

 dH₂O

CTAB DNA Extraction Buffer (Recipe to make 100 mL)

10 mL 1 M Tris Buffer

8.3 g NaCl (1.4 M)

0.744 g EDTA

2 g CTAB

2 g PVP

0.088 g Asorbic acid

Safety warnings

 For Safety Warnings and Hazard Information please refer to the SDS (Satety Data Sheet).



- 1 Grind fresh plant tissue with liquid nitrogen or silica-gel dried tissue in a 1.5 ml Eppie tube.

Note

A little silica gel grains in the tube actually helps the grinding.

- 2 Add  750 µL CTAB DNA Extraction buffer .
- 3 Incubate the CTAB/plant extract mixture for  00:15:00 at  55 °C in the heat block and invert to mix throughout the 15 minutes.
- 4 Add  500 µL Chloroform : IsoAmyl Alcohol (24:1) in the hood and mix the solution by inverting the tubes (**do not vortex**).
- 5 Centrifuge at  13000 rpm for  00:10:00 .
- 6 Transfer the upper aqueous phase only to a new eppie tube (~  500 µL).
- 7 Add  50 µL 7.5M Ammonium acetate followed by  500 µL ice cold 100% ethanol and invert to mix.
- 8 Put tubes in  -20 °C freezer for  00:30:00 (or longer) to precipitate the DNA.
- 9 Centrifuge at  13000 rpm for  00:15:00 .

Note

You should see a pellet at the bottom (align the tube so that you know where the pellet is in case you can't see it very well).



10 Remove the supernatant and wash the DNA pellet as follows. (1/2)

10.1 Add  500 μ L ice cold 70% ethanol . (1/2)

10.2 Centrifuge at  130000 rpm for  00:05:00 . (1/2)

11 Remove the supernatant and wash the DNA pellet as follows. (2/2)

11.1 Add  500 μ L ice cold 70% ethanol . (2/2)

11.2 Centrifuge at  130000 rpm for  00:05:00 . (2/2)

12 Remove all the supernatant and allow the DNA pellet to dry in the hood (approx.  00:20:00).

Note

Do not over dry the pellet since it will be hard to re-dissolve.

13 Resuspend the DNA in  100 μ L dH₂O .

14 Run the DNA on a 1.0% agarose gel to check the quality of the DNA.