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# Congo Red Immunostaining

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Protocol status: Working

We use this protocol and it's working



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#### **Abstract**

Staining for amyloid structures in protein aggregates

## **Troubleshooting**



- 1 Wash freshly sectioned tissues 2-3 times with 1X PBS to remove OCT.
- Mount all sections onto microscope slides and dry at Room temperature 00:00:00.
- Transfer slides into a 1% CongoRed solution for 00:30:00.

30m

Place slides into alkaline bath (1% Sodium hydroxide in 50% ethanol) for 00:05:00.

5m

- 5 Transfer slides into 3 sequential baths of water to remove excess staining.
- Transfer slides into 0.4% Toluidine blue solution for 00:10:00.

10m

- Wash slides in 3 sequential bath of water to remove excess staining.
- 8 Finally, dehydrate slides using sequential baths of:

13m

- 1. distilled water (~ (5) 00:02:00 )
  - 2. 70% ethanol (2 times ~ 👏 00:02:00 )
  - 3. 95% ethanol (2 times ~ 👏 00:02:00 )
  - 4. 100% ethanol (2 times ~ 6 00:02:00 )
  - 5. 10% xylene (2 times ~ 👏 00:05:00 )
- 9 Dry sections at room temperature (~ 👏 00:05:00 ).

5m

10 Cover with DPX mounting medium.