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CODA (part 1): setting up environment and preparing sample dataset | HuBMAP | JHU-TMC V.2

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We use this protocol and it's working

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Abstract

CODA workflow part 1. setting up environment and preparing dataset

Software requirements

- 1 MATLAB [MATLAB \(mathworks.com\)](https://www.mathworks.com/).
Image processing toolbox [Image Processing Toolbox - MATLAB \(mathworks.com\)](https://www.mathworks.com/help/images/).
Deep learning toolbox [Deep Learning Toolbox - MATLAB \(mathworks.com\)](https://www.mathworks.com/help/deeplearning/).
MATLAB - Resnet50 model [MATLAB resnet50 \(mathworks.com\)](https://www.mathworks.com/help/deeplearning/ug/resnet50.html).
- 2 Aperio ImageScope [Aperio ImageScope \(leicabiosystems.com\)](https://leica-biosystems.com/aperio-imagescope/).
- 3 FIJI ImageJ [Fiji Downloads \(imagej.net\)](https://imagej.net/).

Download Source code

- 4 Codes are available at the following GitHub: **CODA Github Repository**.

Download Sample dataset

- 5 Here, we discuss application to a sample dataset “lungs”, containing 150 serial histological images. Download the sample dataset (serial images and sample annotations) here: [**Lung Sample Dataset on Google Drive**](#)
- 6 Images are .ndpi format and were scanned at 20x magnification (approximately 0.5 micron / pixel resolution), spaced 10 micron apart. Save the images in a local drive folder (e.g. **\\Users\\Ashley\\Documents\\lungs**).
- 7 Filenames for each image should be created such that tissue sections are read consecutively by Matlab. Therefore, include zero-padding in numerical indices.

CORRECT FILENAMES: lungs_001.ndpi, lungs_002.ndpi, ..., lungs_011.ndpi

INCORRECT FILENAMES (no zero_padding): **lungs_1.ndpi, lungs_2.ndpi, ..., lungs_11.ndpi**

Create downsampled copies of high-resolution images

- 8 The function **create_downsampled_tif_images** will create downsampled copies of the .ndpi files by directly loading each high-resolution images in tiles and down sampling it to the desired pixel resolutions.

- 9 First, decide the resolution of the images you want to create. Here, we create images of 1 micron / pixel, 2 microns / pixel, and 10 micron / pixel resolution:

```
ds=[1 2 10];
```

- 10 Next, decide on the name of the output folders for each of the downsampled images you create. Here, we will save the images downsampled to 1 micron / pixel in a folder named "10x", the images downsampled to 2 micron / pixel in a folder named "5x," and the images downsampled to 10 micron / pixel in a folder named "1x."

```
subfolders=["10x" "5x" "1x"];
```

- 11 Finally, call the function: **create_downsampled_tif_images(pth,ds,subfolders);**

- 12 Using this function, you will make two subfolders within the original folder containing the .ndpi images. One subfolder named "10x" containing the 20x images downsampled by a factor of 2. The other subfolder named "1x" containing the 20x images downsampled by a factor of 20. Most calculations will be performed on these tif images. Note: here we use 10x and 1x for example, but other resolutions could be created as desired.

```
pth10x=[pth,'10x'];
```

```
pth1x=[pth,'1x'];
```

****Note: If this code fails due to memory constraints on your computer, try python Openslide.**