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Co-immunoprecipitation V5-NIX

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We use this protocol and it's working

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Abstract

This protocol details co-immunoprecipitation.



Materials

Materials and Reagents

Lysis Buffer:

	A	B
	KCl	150 mM
	MgCl ₂	2.5 mM
	Tris-HCl pH 7.4	20 mM
	NP-40	0.50%

Washing Buffer:

	A	B
	KCl	150 mM
	MgCl ₂	2.5 mM
	Tris-HCl pH 7.4	20 mM

- V5-NIX (RRID:Addgene_223731)
-  Anti-V5 agarose beads **Merck MilliporeSigma (Sigma-Aldrich) Catalog #A7345**
-  Pierce™ Detergent Compatible Bradford Assay Kit **Thermo Fisher Catalog #23246**
-  NuPAGE™ 4-12% Bis-Tris Protein Gels, 1.0 mm, 12-well **Thermo Fisher Catalog #NP0322BOX**



Co-immunoprecipitation

1h

- 1 Stably transduce BNIP3/NIX double knockout HeLa cells with lentivirus to express V5-NIX (RRID:Addgene_223731).
- 2 After seeding, treat the cells with DFP to induce mitophagy, where indicated.
- 3 After the treatment, collect the cells by trypsinization and wash the cell pellet with PBS once before cells were lysed in lysis buffer.



Lysis buffer:

	A	B
	KCl	150 mM
	MgCl ₂	2.5 mM
	Tris-HCl pH 7.4	20 mM
	NP-40	0.50%

- 4 Lyse samples for 00:20:00 On ice . Then, centrifuge the cell lysates at 20000 x g, 4°C, 00:10:00 .
- 5 Then, determine the protein concentrations of the cleared protein lysates with the Pierce Detergent Compatible Bradford Assay Kit (23246, Thermo Fisher).
- 6 Incubate 7 mg of cell lysate Overnight with 20 µL of anti-V5 agarose beads (A7345, Sigma-Aldrich).
- 7 In the morning, wash the samples three times in washing buffer and resuspend in protein loading dye, supplement it with 100 millimolar (mM) DTT, and boil it for 00:05:00 at 95 °C .

30m



20m



5m



**Washing buffer:**

	A	B
	KCl	150 mM
	MgCl ₂	2.5 mM
	Tris-HCl pH 7.4	20 mM

- 8 Load the samples on 4-12% SDS-PAGE gels (NP0322BOX, Thermo Fisher) and analyze them by western blotting as described above.

