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# Clusterin cellular uptake assay

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We use this protocol and it's working

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### **Abstract**

This protocol details how to efficiently monitor Clusterin and Clusterin/Substrate uptake in different cell types, like HEK293T, iNeurons and iMicroglia.

### **Attachments**



# Guidelines

- To study Clusterin-A488 uptake in the presence of substrate, incubate [M] 1 micromolar ( $\mu$ M) Clusterin-A488 with the corresponding amount of substrate, e.g. denatured Luciferase or A $\beta$  aggregates, in PBS for 00:20:00 at \$\mathbb{8}\$ 37 °C or \$\mathbb{4}\$ 42 °C (denatured Luciferase) in a total volume of \$\mathbb{\L}\$ 30  $\mu$ L or  $\mathbb{4}$  in the case of HEK293T or iNeurons, respectively. After the incubation time dilute the mix 1/10 in media and add it to the cells resulting in a final concentration of  $\mathbb{M}$  0.1 micromolar ( $\mathbb{\mu}$ M) Clusterin-A488 ( $\mathbb{L}$  5  $\mathbb{L}$  ).
- To monitor substrate uptake e.g., denatured luciferase or Aβ aggregates, in the presence of Clusterin, mix [M] 0.2 micromolar (μM) of denatured Luciferase-pHrodo or [M] 0.5 micromolar (μM) of Aβ-pHrodo aggregates with the corresponding amount of unlabeled Clusterin in a total volume of Δ 30 μL or Δ 40 μL in the case of HEK293T or iNeurons, respectively. Dilute the mix 1/10 in media. pHrodo Red dye is pH sensitive dye which fluoresces brightly only in acidic environments and therefore can be used to specifically monitor phagocytosis and endocytosis, but substrates labeled with A488 can be also used.
- The indicated Clusterin-A488 or substrate concentrations and incubation times for each cell line are tentative. These parameters should be experimentally tested to be in the linear range of the assay.



## **Materials**

- **⊠** ACCUTASE™ 100 mL **STEMCELL Technologies Inc. Catalog** #7920
- X Trypan Blue Solution 0.4% Thermo Fisher Scientific Catalog #15250061

# Troubleshooting



# Clusterin cellular uptake assay - HEK293T cells 25m 1 Plate 100,000 HEK293T cells per well in a 24-well plate. 2 On the next day, add $\perp$ 5 $\mu$ L of Clusterin-A488 together with $\perp$ 300 $\mu$ L of fresh DMEM (without fetal bovine serum, $\perp 4.5 \,\mu g$ in $\perp 4.300 \,\mu L$ medium) to the cells and place the cells back in the incubator. 3 After 04:00:00 incubation, place the plate on ice to stop endocytosis. 4h 4 Wash the cells gently with cold 1x PBS. 5 Add 🗸 100 uL TrypL Express Enzyme (Gibco). Incubate for few minutes 🖁 On ice. 6 Collect the cells with $\perp 400 \,\mu$ of cold medium and transfer them to an Eppendorf tube placed \ \ On ice \ . 7 Centrifuge at 1000 x g, 4°C, 00:05:00 5m 8 8 Discard the supernatant and fix the cells by resuspending the cell pellet with 4 200 µL 10m 4% Paraformaldehyde (PFA) in 1x PBS. Incubate for 00:10:00 at Room temperature 9 Centrifuge at 1000 x g, 4°C, 00:05:00 . 5m

Wash the cell pellet with 1x PBS.

10



11 Centrifuge at 1000 x g, 4°C, 00:05:00.

- 5m

### **iNeurons**



- Add  $\[ \] 5 \ \mu L \]$  of Clusterin-A488 to 250,000 iNeurons cultured in a well of a 12-well plate (add  $\[ \] 2 \ \mu g \]$  Clusterin-A488 to  $\[ \] 200 \ \mu L \]$  of fresh medium and add the mix to the well with cells containing  $\[ \] 200 \ \mu L \]$  conditioned medium) and place the cells back in the incubator.
- After 01:00:00 incubation place the plate on ice to stop endocytosis.

1h

15 Wash the cells gently with cold 1x PBS.

16 Add  $\perp$  100  $\mu$ L Accutase. Incubate for 5-10 minutes  $\parallel$  On ice .

- 17 Collect the cells with  $400 \, \mu L$  of cold medium and transfer them to Eppendorf low binding tubes placed  $700 \, \mu L$  On ice .
- Centrifuge at 1000 x g, 4°C for 00:05:00 (swing-bucked centrifuge preferred).

# 5m

#### Note

The use of low binding tubes and swing-bucked centrifuge significantly reduces cell loss.

Discard the supernatant and fix the cells by resuspending the cell pellet with Δ 200 μL 4% PFA in 1x PBS. Incubate for 00:10:00 at 8 Room temperature .



20 Centrifuge at 1000 x q, 4°C, 00:05:00 (swing-bucked centrifuge preferred). 5m 21 Wash the cell pellet with 1x PBS. 22 Centrifuge at 1000 x q, 4°C, 00:05:00 (swing-bucked centrifuge preferred). 5m 23 Resuspend the cell pellets with  $\perp$  160  $\mu$ L 1x PBS  $\bigcirc$  7.2 and store at  $\parallel$  4  $^{\circ}$ C until analyzed. iMicroglia 1h 15m 24 Dispense 150,000 iMicroglia cells per well with 🚨 300 μL medium into a Geltrexcoated 24-well plate. 25 Add 5  $\perp$  5  $\mu$ L of Clusterin-A488 (  $\perp$  1.5  $\mu$ g in  $\perp$  300  $\mu$ L medium) and place the cells back in the incubator. 26 After (5) 00:30:00 incubation place the plate on ice to stop endocytosis, collect the 30m cells and transfer them to Eppendorf low binding tubes placed \(\begin{aligned} \text{\$\graph\$} \text{On ice} \end{aligned}\). 27 Centrifuge at 1000 x q, 4°C, 00:05:00 (swing-bucked centrifuge preferred). 5m Note The use of low binding tubes and swing-bucked centrifuge significantly reduces cell loss. 28 Wash the cell pellet with 1x PBS.

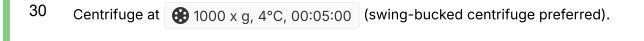
Discard the supernatant and fix the cells by resuspending the cell pellet with 4 200 µL

4% PFA in 1x PBS. Incubate for 6000:10:00 at 8 Room temperature .

29

10m







31 Wash the cell pellet with 1x PBS.



32 Centrifuge at 1000 x q, 4°C, 00:05:00 (swing-bucked centrifuge preferred).





# Uptake quantification



Quantify Clusterin or substrate uptake by measuring A488 or pHrodo Red intensity inside the cells by flow cytometry. If A488 is used, add  $\stackrel{\square}{=}$  50  $\mu$ L of Trypan blue solution 0.4% (refer materials section) right before measuring to quench the 488 fluorescence outside the cells.

#### Note

An Attune NxT flow cytometer (Thermo Fisher Scientific) can be used with the following settings:

- Alexa485: Excitation 488 nm Emission 550/30.
- pHrodo Red: Excitation 561 nm Emission 585/16.