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ChroPlate - IMAC V.2

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Abstract

Purification Guide for the Isolation of Histidine-tagged Proteins with ChroPlate Filtration Plates by DALEX Biotech.

ChroPlate-IMAC by DALEX Biotech offers a robust and convenient way to isolate polyhistidine-tagged proteins from bacterial, mamalian, and insect cell cultures.

Immobilized metal affinity chromatography (IMAC) is based on the interaction of the imidazole ring of histidine with transition metal ions immobilized on a solid support. Recombinant proteins with a 3 - 10 histidine fusion tag bind to these metal ions while unwanted proteins are removed by washing with excess binding buffer. Elution of the target protein is achieved by the addition of imidazole, EDTA or a low pH.

Easy and quick small scale fusion protein purification from various sources.

Low metal leaching from column.

Tolerates reducing reagents (e.g. DTT up to 10 mM).

The proprietary resin does not shrink or swell in aqueous buffers.

High pressure stability.

pH stability short term 2 - 8, long term 3 - 8.

Excellent thermal stability up to 15 minutes at 120 °C in aqueous buffers at neutral pH.

Can be dried for long term storage (80 °C for > 2 h).

Guidelines

It is advisable that all fractions are collected (Sample, flow through, wash, and eluate) in separate plates for analysis, e.g. SDS-PAGE

Materials

Materials provided in the kit:

ChroPlate

Dummy plate

Wash buffer

Elution buffer

Nickel solution

Cobalt solution


Materials not provided in the kit:

DNase/lysozyme

Denaturing wash buffer

Denaturing elution buffer

Safety warnings

 The buffers in the kit include sodium azide (CAS No. 26628-22-8) as a preservative.
For safety information on this chemical(s) check <http://www.dguv.de/ifa/gestis-database>

Before start

Make sure your sample is free of particulate matter. You can remove particles by centrifugation or filtration (0.45 μm).



- 1 How do you want to purify your protein?
Please choose below.

STEP CASE

Native Purification

10 steps

If you try to purify your protein for the first time or you already know that your protein is in the soluble fraction and the his-tag is accessible, use the "Native Purification" protocol (buffers are included in the kit).

Sample Preparation

- 2 Determine the weight of the frozen bacterial pellet and thaw it at room temperature. Resuspend the pellet in wash buffer by pipetting. For every gram of pellet add 3 - 5 milliliters buffer.
Add lysozyme and DNaseI (not included in the kit)
[M] 0.01 Mass / % volume 0.1mg/ml each
Alternatively, use ultrasonication according to the instructions of your instrument's manufacturer and skip the next step.

Note

Pre-chill an appropriate centrifuge to 4 °C.
- 3 Incubate for 15 minutes with gentle end-over-end mixing, stirring, or rocking at room temperature.
If your target protein is known to undergo proteolytic degradation or rapid denaturation, incubate at 4 - 8 °C for 30 minutes.

🌡️ 22 °C 🕒 00:15:00

or

🌡️ 4 °C 🕒 00:30:00
- 4 Centrifuge the lysate for 30 minutes at >10.000 g at 🌡️ 4 °C .
Carefully transfer the supernatant to a fresh tube.

**Note**

During centrifugation you can already proceed with the equilibration of filter plate.

- 5 Please choose below how you want to proceed.

STEP CASE**Purification by Centrifugation** 6 steps**Equilibration**

- 6 Add 200 µl of nickel or cobalt solution to each well. Place the ChroPlate on a deep-well plate and centrifuge at 400 g for 60 seconds in a swing-out rotor. For counterbalance of the centrifuge a dummy filter plate is included in the kit.

Note

Apart from Ni^{2+} and Co^{2+} , you can also use Cu^{2+} or Zn^{2+} .

The affinity of histidine towards the metal ions is in the order $\text{Cu} > \text{Ni} > \text{Zn} > \text{Co}$. However, the specificity is in the inverse order, i.e. copper will most likely result in best yields but with lower purity. In comparison, cobalt will result in a better purity but also lower yields.

- 7 Empty the deep-well plate and add 500 µl deionized water to each well of the ChroPlate. Centrifuge at 400 g for 2 minutes in a swing-out rotor. Repeat once more.
- 8 Empty the deep-well plate and add 500 µl wash buffer to each well of the ChroPlate. Centrifuge at 400 g for 2 minutes in a swing-out rotor.

Load and Wash

- 9 Place the ChroPlate onto a clean deep-well plate. Add up to 1.5 ml cleared lysate and centrifuge at 400 g for 2 - 10 minutes in a swing-out rotor.



Note

It is advisable to keep the flow-through and wash fractions for later analysis, e.g. SDS-PAGE.

Repeat the loading step when your sample volume is larger than 1.5 ml. This is particularly recommended when your target protein has low expression levels.

The centrifugation time depends on the sample's volume and viscosity. Volumes larger than 1 ml and viscous samples e.g. serum usually require more than 5 minutes centrifugation time.

- 10 Empty the deep-well plate or use a clean one, add 500 μ l wash buffer to each well of the ChroPlate, and centrifuge at 400 g for 2 minutes in a swing-out rotor. Repeat once more.

Note

For an increase in purity repeat this step a third time. However, the yield will be lowered.

Elution

- 11 Place the ChroPlate on a clean deep-well plate. Add 300 μ l elution buffer to each well and centrifuge 2 minutes at 400 g in a swing-out rotor. Elute two more times into the same deep-well plate.