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# Chemical extraction of sulfachloropyridazine from feather samples

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Ekaterina Pokrant<sup>1</sup>, Francisca Medina<sup>1</sup>, Aldo Maddaleno<sup>1</sup>, Betty San Martin<sup>2</sup>, Javiera Cornejo<sup>1</sup>

<sup>1</sup>Preventive Medicine Department, Faculty of Veterinary and Animal Sciences, University of Chile;

<sup>2</sup>Laboratory of Veterinary Pharmacology, Faculty of Veterinary and Animal Sciences, University of Chile



Javiera Cornejo

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**Protocol status:** Working

**We use this protocol and it's working**

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**Keywords:** Broiler chickens, feather, Sulfachloropyridazine, residues, mas spectrometry

## Abstract

Analytical methodology for the detection of sulfachloropyridazine (SCP) in samples of feathers via LC-MS/MS was implemented based on techniques previously published by other authors:

- 1- Hindle R. A validated atmospheric pressure chemical ionization method for analysing sulphonamides in pork muscle. 2013. Available at <http://www.youngin.com/application/0411-0045EN-E.pdf>. Accessed October 12, 2017.
- 2- Renew JE, Huang CH. Simultaneous determination of fluoroquinolone, sulfonamide, and trimethoprim antibiotics in wastewater using tandem solid phase extraction and liquid chromatography–electrospray mass spectrometry. *J Chromatogr A* 2004; 1042: 113-21.
- 3- Shao B, Dong D, Wu y, Hu J, Meng J, Tu X, Xu S. Simultaneous determination of 17 Sulfonamide residues in porcine meat, kidney and liver by solid phase extraction and liquid chromatography-tandem mass spectrometry. *Anal Chim Acta* 2005; 546:174-81.
- 4- Pang G, Cao YZ, Zhang JJ, Jia GQ, Fan CL, Li XM, Liu YM, Li ZY, Shi YQ. Determination of sulfonamides in honey by liquid chromatography- tandem mass spectrometry. *J AOAC Int* 2005; 88:1304-11.
- 5- Stubbings G, Bigwood T. The development and validation of a multi-class liquid chromatography tandem mass spectrometry (LC– MS/MS) procedure for the determination of veterinary drug residues in animal tissue using a QuEChERS (QUick, Easy, CHEap, Effective, Rugged and Safe) approach. *Anal Chim Acta* 2009; 637:68-78.
- 6- Bedendo GC, Jardim IC, Carasek E. A simple hollow fiber renewal liquid membrane extraction method for analysis of sulphonamides in honey samples with determination by liquid chromatography-tandem mass spectrometry. *J Chromatogr A* 2010; 1217:6449-54.
- 7- Yu H, Tao Y, Chen D, Wang Y, Huang L, Peng D, et al. Development of a high-performance liquid chromatography method and a liquid chromatography-tandem mass spectrometry method with the pressurized liquid extraction for the quantification and confirmation of sulphonamides in the foods of animal origin. *J Chromatogr B* 2011; 879:2653-62.

The method is based on a solid-liquid extraction with organic solvents. The clean up is carried out through aromatic sulfonic acid (Bakerbond spe™) disposable extraction columns of 6 mL. The analyte is concentrated using a water bath at 40-50°C under a mild nitrogen flow. For the instrumental analysis, a Symmetry C8 analytical column of 3.5µm and 2.1 × 100mm (Waters®) was fitted in an Agilent 1290 infinity series liquid-chromatograph equipment, coupled to an API 3200 (AB Sciex, Darmstadt, Germany) triple-quadrupole mass-spectrometer. The analytical data was then integrated using the Analyst® version 1.5 software package (SCIEX, Framingham, Massachusetts).



## Materials

### MATERIALS

✕ 1.5 mL Eppendorf tubes

✕ Falcon Tube (50 mL) **Fischer Scientific**

✕ BAKERBOND spe™ Aromatic Sulfonic Acid (C<sub>6</sub>H<sub>5</sub>SO<sub>3</sub>H) Disposable Extraction Columns **J.T. Baker Catalog #7090-29**

✕ Sulfamethazine-(phenyl-13C<sub>6</sub>) hemihydrate **Merck MilliporeSigma (Sigma-Aldrich) Catalog #32519**

✕ Sulfachloropyridazine **Merck MilliporeSigma (Sigma-Aldrich) Catalog #S9882**

✕ 1 ml syringe **Catalog #JD+01T2713**

✕ 10 mL syringe **Catalog #JD+10L2125-WEI**

✕ Glass wool **Merck Millipore (EMD Millipore) Catalog #1040860250**

✕ Millex Syringe Filter **Merck Millipore (EMD Millipore) Catalog #SLGVX13NK**

### STEP MATERIALS

✕ Ethyl Acetate **J.T. Baker Catalog #9280-03**

✕ BAKERBOND spe™ Aromatic Sulfonic Acid (C<sub>6</sub>H<sub>5</sub>SO<sub>3</sub>H) Disposable Extraction Columns **J.T. Baker Catalog #7090-29**

✕ Ethyl Acetate **J.T. Baker Catalog #9280-03**

✕ n-Hexane **Merck Millipore (EMD Millipore) Catalog #1037014000**

✕ Methanol **J.T. Baker Catalog #9093-03**

✕ Water **Merck Millipore (EMD Millipore) Catalog #1153334000**

✕ Methanol **J.T. Baker Catalog #9093-03**

✕ Ammonia solution 25% **Merck Millipore (EMD Millipore) Catalog #1054322500**

✕ Formic Acid 98-100% **Merck Millipore (EMD Millipore) Catalog #100264**

✕ Methanol **J.T. Baker Catalog #9093-03**

✕ Water **Merck Millipore (EMD Millipore) Catalog #1153334000**

✕ Ethyl Acetate **J.T. Baker Catalog #9280-03**

✕ BAKERBOND spe™ Aromatic Sulfonic Acid (C<sub>6</sub>H<sub>5</sub>SO<sub>3</sub>H) Disposable Extraction Columns **J.T. Baker Catalog #7090-29**

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⊗ Methanol **J.T. Baker Catalog #9093-03**

⊗ Water **Merck Millipore (EMD Millipore) Catalog #1153334000**



## Protocol materials

- ☒ Methanol **J.T. Baker Catalog #9093-03**
- ☒ 10 mL syringe **Catalog #JD+10L2125-WEI**
- ☒ 1.5 mL Eppendorf tubes
- ☒ Falcon Tube (50 mL) **Fischer Scientific**
- ☒ Sulfachloropyridazine **Merck MilliporeSigma (Sigma-Aldrich) Catalog #S9882**
- ☒ Millex Syringe Filter **Merck Millipore (EMD Millipore) Catalog #SLGVX13NK**
- ☒ Ethyl Acetate **J.T. Baker Catalog #9280-03**
- ☒ Water **Merck Millipore (EMD Millipore) Catalog #1153334000**
- ☒ Ethyl Acetate **J.T. Baker Catalog #9280-03**
- ☒ n-Hexane **Merck Millipore (EMD Millipore) Catalog #1037014000**
- ☒ Ammonia solution 25% **Merck Millipore (EMD Millipore) Catalog #1054322500**
- ☒ Methanol **J.T. Baker Catalog #9093-03**
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- ☒ Methanol **J.T. Baker Catalog #9093-03**
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- ☒ Methanol **J.T. Baker Catalog #9093-03**
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- ☒ Ethyl Acetate **J.T. Baker Catalog #9280-03**
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- ☒ Formic Acid 98-100% **Merck Millipore (EMD Millipore) Catalog #100264**
- ☒ Water **Merck Millipore (EMD Millipore) Catalog #1153334000**
- ☒ 1 ml syringe **Catalog #JD+01T2713**

- ✕ Glass wool **Merck Millipore (EMD Millipore) Catalog #1040860250**
- ✕ Ammonia solution 25% **Merck Millipore (EMD Millipore) Catalog #1054322500**
- ✕ Water **Merck Millipore (EMD Millipore) Catalog #1153334000**
- ✕ Sulfamethazine-(phenyl-13C6) hemihydrate **Merck MilliporeSigma (Sigma-Aldrich) Catalog #32519**
- ✕ Ethyl Acetate **J.T. Baker Catalog #9280-03**
- ✕ Ethyl Acetate **J.T. Baker Catalog #9280-03**
- ✕ n-Hexane **Merck Millipore (EMD Millipore) Catalog #1037014000**
- ✕ BAKERBOND spe™ Aromatic Sulfonic Acid (C6H5SO3H) Disposable Extraction Columns **J.T. Baker Catalog #7090-29**
- ✕ Water **Merck Millipore (EMD Millipore) Catalog #1153334000**
- ✕ Methanol **J.T. Baker Catalog #9093-03**
- ✕ Methanol **J.T. Baker Catalog #9093-03**
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- ✕ Formic Acid 98-100% **Merck Millipore (EMD Millipore) Catalog #100264**
- ✕ Methanol **J.T. Baker Catalog #9093-03**
- ✕ Water **Merck Millipore (EMD Millipore) Catalog #1153334000**

## Safety warnings

- ❗ Protect hands, eyes and face from organic solvents and liquid nitrogen during all the extraction steps with all the necessary materials.

## Before start

Feather samples must be cryogenically treated with liquid nitrogen and then ground in an industrial Robot Coupe® R4 table-top cutter food processor (Burgundy, France) to ensure their homogeneity.



Weigh  $2 \pm 0.02$  g of feather sample in a 50-mL polypropylene tube.


1

Fortifie each sample with the internal standard solution, Sulfamethazine-(phenyl-<sup>13</sup>C<sub>6</sub>) hemihydrate (SMZ-<sup>13</sup>C<sub>6</sub>) and the positive controls with the analyte Sulfamethazine .

2

Samples must be rested before extraction.

3

 00:15:00


Add 40 mL of ethyl acetate.

4

 Ethyl Acetate J.T. Baker Catalog #9280-03


Stirr the samples on a vortex-mixer.

5

 00:15:00


Sonicate the sample.

6

 00:05:00

Centrifuge at 1,800 g

7

 00:10:00

Filter the supernatant through glass wool, using a syringe of 10 mL. Pass the filtrate to a fresh 50-mL polypropylene tube.


8

Concentrate down to 15 mL using a water bath at 40-50 °C under a mild nitrogen flow.

9

Condition Aromatic sulfonic acid (Bakerbond spe™) disposable extraction columns with 6 mL of hexane and 6 mL of ethyl acetate.

10

 BAKERBOND spe™ Aromatic Sulfonic Acid (C<sub>6</sub>H<sub>5</sub>SO<sub>3</sub>H) Disposable Extraction Columns J.T. Baker Catalog #7090-29

 Ethyl Acetate J.T. Baker Catalog #9280-03

 n-Hexane Merck Millipore (EMD Millipore) Catalog #1037014000

Filter the samples through the columns.

11

Wash the column with 2 mL of water and 2 mL of methanol.


12

 Methanol J.T. Baker Catalog #9093-03

 Water Merck Millipore (EMD Millipore) Catalog #1153334000

Elute the column with 10 mL of a mixture of methanol and ammonia solution (97/3)

13

 Methanol J.T. Baker Catalog #9093-03

 Ammonia solution 25% Merck Millipore (EMD Millipore) Catalog #1054322500

Evaporate the samples using a water bath at 40-50°C under a mild nitrogen flow.





14

Reconstitute the sample with 300  $\mu$ L of a mixture of mobile phase A and B (15/85)

15

**Note**

Mobile phase A: 0.1% formic acid in methanol (pH  $2.9 \pm 0.3$ ).  
Mobile phase B: 0.1% formic acid in water (pH  $2.7 \pm 0.2$ ).


 Formic Acid 98-100% **Merck Millipore (EMD Millipore) Catalog #100264**

 Methanol **J.T. Baker Catalog #9093-03**

 Water **Merck Millipore (EMD Millipore) Catalog #1153334000**


Stirr the reconstituted solution on a vortex-mixer.

16

 00:05:00


Sonicate the reconstituted solution.

17

 00:05:00

Transfer the reconstituted solution into an Eppendorf tube and centrifuge at 17,000 g

18

 00:10:00

Filter the sample through 13 mm millex filters with 0.22  $\mu$ m polyvinylidene fluoride (PVDF) membranes and transfer into a glass vials.

19