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Characterization of iPSC

 In 1 collection

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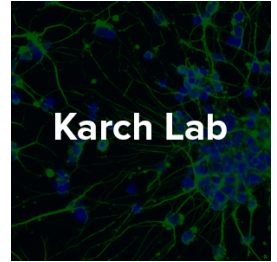
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Protocol status: Working

We use this protocol and it's working

Created: February 17, 2019

Last Modified: February 27, 2019

Protocol Integer ID: 20474



Attachments



Comprehensive

Genomi...

31KB

Guidelines

This protocols is part of the Screening Edited iPSC Clones collection.

Safety warnings









⚠ Please refer to the SDS (Safety Data Sheet) for information about hazards, and to obtain advice on safety precautions.

Before start




Starting Material: 1 confluent 6 well plate.

Use 2 wells to characterize the iPSC and 4 wells for freezing the iPSC.



- 1 Coat T25 flask, chamber slide and plates with Matrigel  01:00:00 prior to passaging.
- 2 Aspirate media from cell culture.
- 3 Wash with  1 mL PBS per well and aspirate.
- 4 Add  1 mL Accutase per well of 6 well plate.
- 5 Incubate in  37 °C for 3-4 minutes.  00:03:00
- 6 Collect cells from 2 wells with  3 mL DMEM/F12 per well and transfer into 15mL conical A.
- 7 Collect cells from 4 wells with  3 mL DMEM/F12 per well and transfer into 15mL conical B.
- 8 Spin cells at 750 rpm for  00:03:00 .
- 9 Aspirate media from each tube.

Tube A

- 10 To 15mL conical tube A, add  2 mL mTesR1 and distribute cells.
- 11 Karyotype: Add  2 mL of mTesR1 to T25 flask, then add  500 μ L cells .



- 12 gDNA pellet: 500 μL cells in 1.7 mL tube, spin down at max speed for 00:00:15 , aspirate media, store in -80 $^{\circ}\text{C}$.
- 13 RNA pellet: 900 μL cells in 1.7 mL tube, spin down at max speed for 00:00:15 , aspirate media, store in -80 $^{\circ}\text{C}$
- 14 ICC: Dilute 100 μL cells in 750 μL mTesR1. Plate 200 μL cells per well in 4 wells of a chamber slide.

Tube B

- 15 To 15mL conical tube B, add 4 mL mTesR1 and 4 mL of 2x Freezing Media (20% DMSO, FBS).
- 16 Add 1 mL cell suspension to each vial (1 well will freeze down into approximately 2 vials).

Note

Cryovials need to be labeled with the following before freezing down:

- Cell Type
- Line Name
- Passage #
- Date
- Your Name