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🌐 Cell dissociation of fresh human lung tissue for single-cell RNA-seq

DOI

<https://dx.doi.org/10.17504/protocols.io.zp2f5qe>

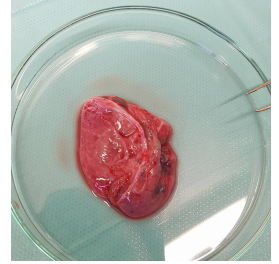
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Human Cell Atlas Metho...



Ilias Angelidis



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External link: <https://www.helmholtz-muenchen.de/ilbd/research/ilbdcpc-junior-research-groups/systems-medicine-of-chronic-lung-disease-schiller-lab/scientific-focus/index.html>

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Protocol status: Working

We use this protocol and it's working

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Guidelines

Human body fluids and tissue potentially contain blood borne viruses and other agents. Work with blood samples or tissue from individuals therefore carries a risk of infection if the material is not handled with care.

All practices must follow all safety guidelines regarding human tissue handling.

Materials

MATERIALS

✕ DNase I, RNase-free **Qiagen Catalog #79254**

✕ PBS without Ca² or Mg² **Gibco - Thermo Fisher Scientific Catalog #10010-031**

✕ Dispase **Corning Catalog #354235**

✕ Collagenase CLS I **Biochrom AG Catalog #C1-28**

✕ Elastase **Serva, Germany Catalog #20931**

✕ FBS **Biochrom AG Catalog #S 0615**

STEP MATERIALS

✕ RBC Lysis Buffer **Invitrogen - Thermo Fisher Catalog #00-4333-57**



Protocol materials

⊗ DNase I, RNase-free **Qiagen Catalog #79254**

⊗ PBS without Ca²⁺ or Mg²⁺ **Gibco - Thermo Fisher Scientific Catalog #10010-031**

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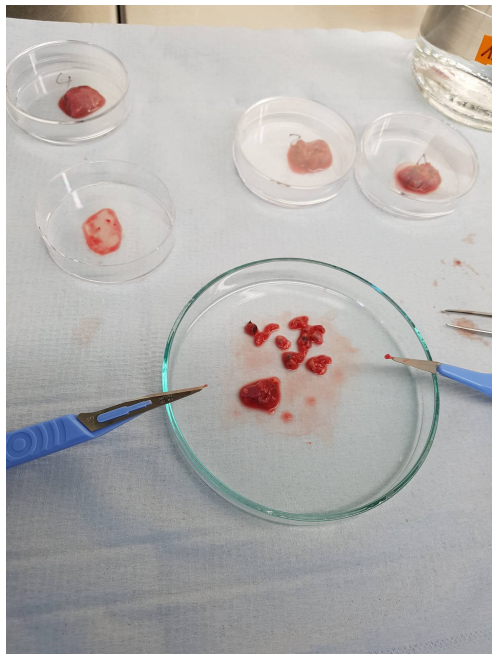
⊗ RBC Lysis Buffer **Invitrogen - Thermo Fisher Catalog #00-4333-57**

- 1 Transfer lung tissue in a petri dish and cut the required amount for your experiment (2×2×1)cm.

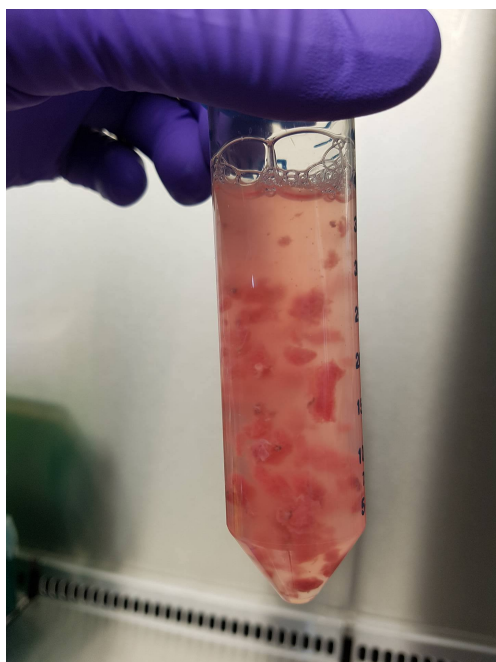


Desired amount of tissue is cut off

- 2 Mince the tissue into small pieces using a pair of scissors. Transfer tissue chunks in a 50mL Falcon tube containing  30 mL of ice cold PBS. This step is intended to wash away any remaining blood off the tissue.



Tissue is cut into small pieces



Tissue is washed in PBS

- 3 Remove PBS by passing the minced tissue through a 40µm strainer and keeping the tissue pieces on top.



Tissue is now minced, washed and ready for Digestion.

- 4 Transfer the tissue into a new 50mL Falcon tube containing  8 mL of **Enzyme Mix** (should suffice for 2×2×1 cm of tissue).

	Enzyme	Cat. Number	Final Concentration
	Dispase	354235 (Corning)	50 caseinolytic units/ml
	Collagenase	C1-28 (Merck)	0,6 mg/ml

	Elastase	20931 (Serva)	0,02 mg/ml
	DNaase	79254 (QIAGEN)	17 units/ml

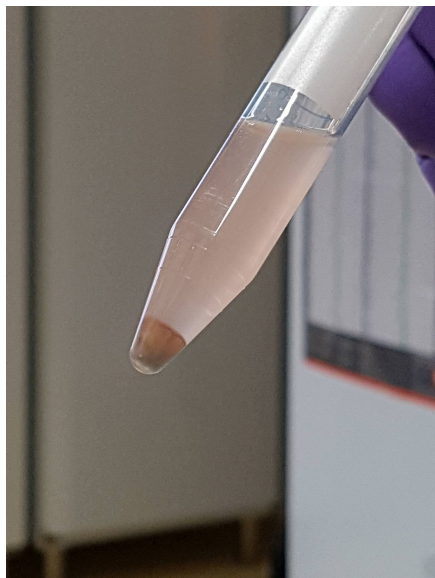
Enzyme Mix

- 5 Incubate for:  00:50:00 at  37 °C with constant shaking at 750rpm.
- 6 After enzymatic digestion add  7 mL of ice cold **Inactivation Buffer**. Mix well using a 10mL serological pipette and pass the cell suspension from the digested tissue through a 70µm strainer into a 15mL Falcon tube (keep flow-through).

	Reagent	Cat. Number	Final Concentration
	PBS	10010-015 (gibco)	1X
	FBS	S0615 (Merck)	10%

Inactivation Buffer

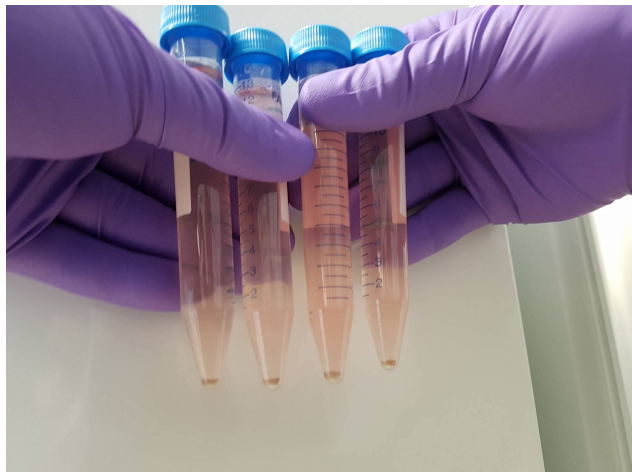
- 7 Centrifuge the cell isolate at **300g** for 00:05:00 at 4 °C .



Typical cell pellet

- 8 Resuspend cell pellet in 2 mL of **RBC Lysis Buffer** at RT for 00:02:00 to remove remaining red blood cells.

 RBC Lysis Buffer Invitrogen - Thermo Fisher Catalog #00-4333-57



Cell pellet after RBC

- 9 Add  10 mL of ice cold **Inactivation Buffer** to inhibit the activity of the RBC Lysis Buffer.



- 10 Centrifuge the cell isolate at **300g** for  00:05:00 at  4 °C .
- 11 Resuspend in  1 mL of ice cold **Inactivation Buffer** and count the cells.
- 12 Assess cell viability using Trypan Blue staining. (cell viability must be around 85%-95% in order to yield high quality single cell libraries)
- 13 Proceed with preferred scRNA-seq platform using the appropriate number of cells.