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 We use this protocol and it's working

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Cell deposition and drug sensitivity assay

In 2 collections

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Protist Research to Optimize Tools in Genetics (PROT-G)

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ABSTRACT

This protocol describes how to deposit several cells of *S. robusta* in droplets on a petri dish, allowing the experimenter to track the growth of approximately single cell isolates with high throughput. This was used to determine drug sensitivity.

We found that in, liquid culture, *S. robusta* is sensitive to 10 ug/mL puromycin and 250 ug/mL glyphosate. One caveat to keep in mind is that *S. robusta* appears more tolerant to antibiotics when grown on agar plates, being insensitive to up to 20 ug/mL of puromycin, for example. We recommend selection in liquid culture.

MATERIALS

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⊗ Puromycin dihydrochloride **US Biological Catalog #P9200**

⊗ Hygromycin B **Phytotech Labs Catalog #h397**

⊗ Glyphosate **Phytotech Labs Catalog #g345**

Measure density of floating cells in culture

- 1 Grow 100 mL volume of cells in a 175 cm² flask for three to four days, until there is a substantial number of healthy, floating cells
- 2 Stand flask vertically and take a 15 mL volume of the floating culture (so as to not disturb the adherent cells) to measure cell density
- 3 Spin cells down at 6000xg to pellet them. Scrape and resuspend in 1 mL
- 4 Measure cell density using a hemocytometer. The target concentration for distributing cells is 400-500 floating cells/mL, so dilute as necessary. The volume needed is 2 mL, including any drugs you want to add.

Deposit cells in droplets on a petri dish

- 5 b. Take the 2 mL, containing the desired number of cells, in an Eppendorf tube.
- 6 Centrifuge using a personal mini-centrifuge for two minutes. At this speed (which I believe is 6000 rpm), many cells remain in the supernatant. Those are the cells we want. Do not disturb any pellet that forms from the centrifugation.
- 7 Using a Pipetteman, withdraw 40µl aliquots from near the top of the Eppendorf tube. Deposit 40µl drops in a 6x8 array in a Petri dish.

Count initial cells

8 Record the approximate number of cells deposited in each drop.

9 In our hands, this is ~4-10 cells per droplet on average

10 Wrap petri dish with parafilm and place in incubator

Assay

11 After three days of treatment, count cell numbers/determine cell viability in each droplet