Prepare 1.5ml tube by adding a few glass beads.

To each tube add ~200mg of leaf material

Freeze in liquid N2 and grind using a homogeniser

Prepare CTAB buffer:

Add 2% beta-mercaptoethanol to the CTAB buffer just before use (200μl/10ml)

CTAB Buffer:
2% CTAB
2% PVP-40
100mM Tris-HCL pH 8.0
25mM EDTA-Na
2M NaCl
0.5g/L spermidine
5. Add 1ml CTAB buffer to the sample, mix, incubate 15min at 50°C

6. Centrifuge 5min full speed, transfer 900ul supernatant to fresh 2ml tube

7. Add 900 ul Chloroform:isoamyl alcohol (24:1), mix, centrifuge for 10min full speed

8. Transfer 800ul supernatant to fresh tube and add equal volume of Chloroform:isoamy alcohol (24:1), mix, centrifuge 10 min full speed

9. Transfer 650ul supernatant to fresh 1.5 ml tube, add 390 ul (0.6 volume) cold EtOH

10. Incubate at -80°C for 30 min or at -20C overnight.

11. Centrifuge 30 min full speed at 4°C

12. Remove supernatant

13. Wash the pellet in 1 ml 80% ETOH

14. Centrifuge 5min and vacuum dry the pellet.

15. Dissolve the pellet in 100 ul DEPC-treated water. *continue on ice

16. Measure the DNA concentration using a NanoDrop.

17. Pipette out as much total NA as required.
18 Continue with the following steps for RNA precipitation

19 Add 1/5th volume of 10M LiCl

20 Incubate at -20 for at least 1 hour

21 Centrifuge full speed (4C) for 30 min

22 Wash with 80% EtOH

23 Centrifuge 5min at 4000 rpm, at 4C, and vacuum dry the pellet.

24 Resuspend in 30-50ul of H2O