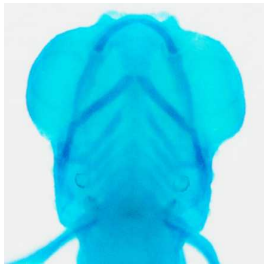
Dec 18, 2023 Version 1

Cartilage staining V.1



DOI
https://dx.doi.org/10.17504/protocols.io.n2bvj3je5lk5/v1

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Fish behavior and physi... Zebrafish Research

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Protocol Citation: Satheeswaran Balasubramanian, Ekambaram Perumal 2023. Cartilage staining. **protocols.io** https://dx.doi.org/10.17504/protocols.io.n2bvj3je5lk5/v1Version created by Satheeswaran Balasubramanian

Manuscript citation:
Balasubramanian S, Rangasamy S, Vivekanandam R, Perumal E. Acute exposure to tenorite nanoparticles induces phenotypic and behavior alterations in zebrafish larvae. Chemosphere. 2023 ;339:139681. https://doi.org/10.1016/j.chemosphere.2023.139681

Protocol status: Working
We use this protocol and it's working

Created: November 25, 2023

Last Modified: December 18, 2023

Protocol Integer ID: 91415

Keywords: Cartilage, zebrafish, alcian blue, development of cartilage, alcian blue, staining technique, bone in zebrafish embryo, cartilage, cartilage through an electrostatic interaction, charged dye, zebrafish embryo, alcian, developmental biologist, bone

Funders Acknowledgements:
ICMR-SRF
Grant ID: No.45/51/2020-Nan/BMS

Abstract

The Alcian blue staining technique is widely used among developmental biologists to observe the development of cartilage and bone in zebrafish embryos and larvae. Alcian blue is a positively charged dye that stains the cartilage through an electrostatic interaction with negatively charged acidic mucopolysaccharides in the presence of magnesium ions (Scott et al. 1996).

Guidelines

This protocol is optimized for 5 days post-fertilization (dpf) of zebrafish larvae. It can be adapted for higher dpf.

Materials

	A	B	C
	Abbreviation	**Chemical Name**	**Molecular Weight (g/mol)**
	Na2HPO4	Disodium hydrogen phosphate	141.96
	MgCl2	Magnesium chloride	95.21
	KCl	Potassium chloride	74.55
	KH2PO4	Potassium dihydrogen phosphate	136.09
	KOH	Potassium hydroxide	56.11
	NaCl	Sodium chloride	58.44
	dH2O	Distilled water	-
	EtOH	Ethanol	46.07
	PFA	Paraformaldehyde	30.03
	PBS	Phosphate-buffered saline	-
	PBST	PBS plus 0.1% Tween 20	-
	MeOH	Methanol	32.04

1. 10X PBS stock; pH 7.4: To prepare 1 L stock, add 1.37 M (80 g) NaCl, 27 mM (0.2 g) KCl, 100 mM (14.4 g) Na_2HPO_4, and 18 mM (0.24 g) KH2PO4 in 800 ml of dH2O. Make up the volume to 1000 ml and autoclave. Store at room temperature. To prepare 1X PBS, add 100 ml of 10X PBS to 900 ml dH2O.
2. 1X PBST: Add 0.1 ml of Tween 20 to 100 ml of 1X PBS and mix.
3. 4 % PFA; pH 7.4: Dissolve 4 g of PFA in 80 ml of PBS at 60 °C, until the solution clears. Cool the solution and make it to 100 ml. To avoid repeated thawing and freezing, PFA can be aliquoted in 5 ml tubes and stored at −20 °C.
4. Bleach solution: For 1 ml, add 100 µl of 30% solution of H2O2, 250 µl of 2% KOH and 650 µl of dH2O.
5. Staining solution: For 1 ml, add 100 µl of 10X alcian blue (0.01%), 200 µl of 1 M MgCl2 (60 mM), and 700 µl of 100% EtOH (70%). a) 10X of alcian blue solution (8 GX; 05500, Sigma Aldrich): For 10 ml solution, add 10 mg alcian blue and 10 ml of methanol. b) 300 mM of MgCl2: Add 0.29 g of MgCl2 to 10 ml of dH2O. c) 100% EtOH.
6. Storage solution: Keep 100% glycerol and 1% KOH (100 mg in 10 ml) separately.
   a. 25% solution: For 1 ml, mix 250 µL glycerol, 250 µl of KOH, and 500 µl of dH2O.
   b. 50% solution: For 1 ml, mix 500 µL glycerol, 250 µl of KOH, and 250 µl of dH2O.

Safety warnings

Make sure to read all Safety Data Sheets for the reagents. Hydrogen peroxide and paraformaldehyde solution must be prepared under a fume hood at all times.

Before start

1. Every protocol needs to be validated in the laboratory when first introduced. The present protocol describes validation steps that were taken in the Molecular Toxicology lab, at Bharathiar University.
2. Always wear personal protective equipment.
3. All the steps are optimized in a 24-well plate for multiple groups such as control and treated.

Larval fixation

3h

1 At the desired stage, take the zebrafish larvae and wash in PBS for 5 minutes. 5m

2 Euthanize the larvae using the cold shock method (Keep at 4^{o}C for 5-10 minutes). 10m

3 Transfer the euthanized larvae into 4% PFA and keep it in the rocker for 2 hours at room temperature (if required, it can be kept overnight at 4^{o}C). 2h

4 Wash the fixed larvae with PBST for 5 minutes, two times. 10m

5 Dehydrate the larvae using 50% ethanol and keep it in the rocker for 10 minutes at room temperature. 10m

Staining

8h

6 Add 1 ml of alcian blue staining solution to the dehydrated larvae and keep it in a rocker overnight at room temperature. 8h

Bleaching

40m

7 Rinse the stained larvae once using PBST. 1m

8 Transfer the larvae to a bleach solution and incubate in dark at room temperature. Assess the bleaching process (using a microscope) and stop the reaction when the larvae become transparent (approximately 20-30 min for 4 dpf and above). 30m

Clearing

9h

9 Remove the bleach solution and rinse once using PBST. 1m

10 Clear the beached larvae with 25% glycerol and 0.25% KOH. Keep them in a rocker at room temperature for 30 minutes to overnight. 30m

11 Replace the solution with 50% glycerol and 0.25% KOH. Keep them in a rocker at room temperature for 2 hours to overnight. 8h

Storage

12 Store larvae in a solution of 50% glycerol and 0.1% KOH at 4^{o}C. Avoid long-term storage as it will diminish the stain. Capture images of the cartilage using any bright field microscope (preferably a stereo microscope).

Results

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Expected result

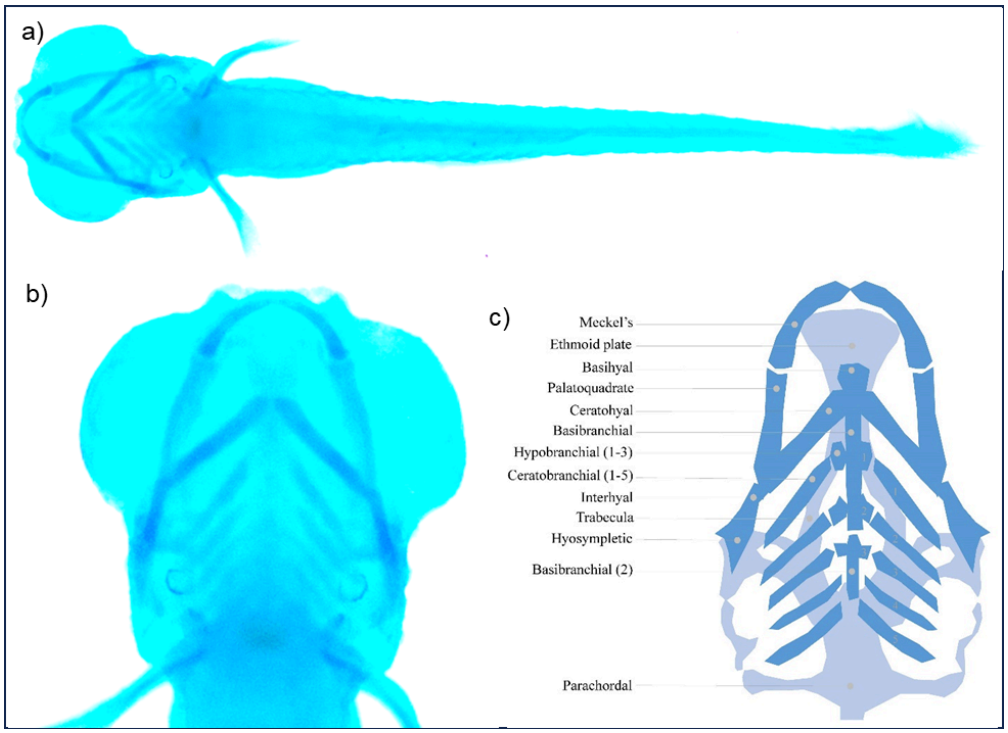

Figure 1. Representative staining images of 5 dpf larvae. a) Whole larvae; b) head region (taken in molecular toxicology lab, Bharathiar University); c) Schematic representation of cartilaginous structures in the head region of zebrafish (adapted from Raterman et al. 2020).

Protocol references

Dr Mb Walker & Cb Kimmel (2007) A two-color acid-free cartilage and bone stain for zebrafish larvae, Biotechnic & Histochemistry, 82:1, 23-28, DOI: 10.1080/10520290701333558