

Oct 08, 2025

CALCIUM IMAGING PROTOCOL

DOI

<https://dx.doi.org/10.17504/protocols.io.e6nvw4k22lmk/v1>

Carmen Perez De Nanclares¹

¹University of Minnesota

Team Lee



Jane Balster

ASAP - Team Lee

Create & collaborate more with a free account

Edit and publish protocols, collaborate in communities, share insights through comments, and track progress with run records.

Create free account

OPEN  ACCESS



DOI: <https://dx.doi.org/10.17504/protocols.io.e6nvw4k22lmk/v1>

Protocol Citation: Carmen Perez De Nanclares 2025. CALCIUM IMAGING PROTOCOL. protocols.io
<https://dx.doi.org/10.17504/protocols.io.e6nvw4k22lmk/v1>

License: This is an open access protocol distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

Protocol status: Working

We use this protocol and it's working

Created: July 21, 2025

Last Modified: October 10, 2025

Protocol Integer ID: 223838

Keywords: ASAPCRN, electrophysiology, calcium imaging protocol, calcium imaging protocol this protocol detail, activity in astrocyte, astrocyte, ca1 region, stratum radiatum of the ca1 region, mouse hippocampus, imaging

Abstract

This protocol details Ca²⁺ imaging to analyze activity in astrocytes of the stratum radiatum of the CA1 region of the mouse hippocampus.

Attachments



Calcium Imaging Prot...

172KB



- 1 Locate Ca^{2+} levels in astrocytes in the stratum radiatum of the CA1 region of the hippocampus are monitored using the Ca^{2+} indicator fluo-4.
- 2 Load Astrocytes with the dye by incubating the slices with fluo-4-AM ([M] 2 micromolar (μM) and 0.01% of pluronic) for 🕒 00:45:00 - 🕒 01:00:00 at 🌡 Room temperature (Araque et al. 2002; Kang et al. 1998; Nemani et al. 2010; Nett et al. 2002; Navarrete and Araque 2008).
- 3 This protocol has been proven to be suitable for measuring the calcium activity of astrocytes (Nanclares et al. 2023).
- 4 Briefly, fluo-4-positive cells present typical electrophysiological properties for astrocytes, this is, low input resistance, a quasi-linear voltage–current relationship under voltageclamped conditions, and absence of action potentials.
- 5 On the contrary, recordings from fluo-4-negative cells in the stratum radiatum exhibit typical neuronal characteristics, such as higher input resistances, a non-linear voltage–current curve, and firing of action potentials.
- 6 Additionally, incubate the slices with fluo-4 and sulforhodamine 101 (SR101), a widely used astrocyte marker known to label astrocytes in the hippocampus (Schnell, Hagos, and Hülsmann 2012), provide good co-localization between SR101 and fluo-4-positive cells (Nanclares et al. 2023).
- 7 Perform the recordings in the presence of a cocktail of neurotransmitter receptor antagonists to minimize neuronal contribution to astrocyte Ca^{2+} activity (CNQX [M] 20 micromolar (μM) , AP5 [M] 50 micromolar (μM) , MPEP [M] 50 micromolar (μM) , LY367385 [M] 100 micromolar (μM) , picrotoxin [M] 50 micromolar (μM) , CGP54626 [M] 1 micromolar (μM) , atropine [M] 50 micromolar (μM) , CPT [M] 2 micromolar (μM) , suramin [M] 100 micromolar (μM) , flupenthixol [M] 30 micromolar (μM) , AM251 [M] 2 micromolar (μM) and TTX [M] 1 micromolar (μM) to block glutamatergic, GABAergic, cholinergic, purinergic, endocannabinoid and dopaminergic transmission).
- 8 Image Astrocytes using a custom-made confocal microscope (Thorlabs).
- 9 Obtain the videos at 512×512 resolution with a sampling interval of 🕒 00:00:01 .

1h 45m



1s

- 10 Use ImageJ software (NIH) for quantitative fluorescence measurements.
- 11 Estimate Ca^{2+} variations as changes in the fluorescence signal over baseline (DF/F0), and cells are considered to show a Ca^{2+} event when DF/F0 increases two times the standard deviation of the baseline.
- 12 Quantify the astrocyte Ca^{2+} signal as the number of Ca^{2+} events per min within  00:10:00 of recording. 10m
- 13 Obtain Astrocyte Ca^{2+} events from 5 to 24 astrocytes in the field of view during the recording.

Protocol references

- Araque, Alfonso, Eduardo D. Martín, Gertrudis Perea, Jon I. Arellano, and Washington Buño. 2002. "Synaptically Released Acetylcholine Evokes Ca^{2+} Elevations in Astrocytes in Hippocampal Slices." *The Journal of Neuroscience* 22 (7): 2443–50. <https://doi.org/10.1523/JNEUROSCI.22-07-02443.2002>.
- Kang, Jian, Li Jiang, Steven A. Goldman, and Maiken Nedergaard. 1998. "Astrocyte-Mediated Potentiation of Inhibitory Synaptic Transmission." *Nature Neuroscience* 1 (8): 683–92. <https://doi.org/10.1038/3684>.
- Nanclares, Carmen, Jonah Poynter, Hector A. Martell-Martinez, et al. 2023. "Dysregulation of Astrocytic Ca^{2+} Signaling and Gliotransmitter Release in Mouse Models of α -Synucleinopathies." *Acta Neuropathologica* 145 (5): 597–610. <https://doi.org/10.1007/s00401-023-02547-3>.
- Navarrete, Marta, and Alfonso Araque. 2008. "Endocannabinoids Mediate Neuron-Astrocyte Communication." *Neuron* 57 (6): 883–93. <https://doi.org/10.1016/j.neuron.2008.01.029>.
- Nemani, Venu M., Wei Lu, Victoria Berge, et al. 2010. "Increased Expression of α -Synuclein Reduces Neurotransmitter Release by Inhibiting Synaptic Vesicle Reclustering after Endocytosis." *Neuron* 65 (1): 66–79. <https://doi.org/10.1016/j.neuron.2009.12.023>.
- Nett, Wolfgang J., Scott H. Oloff, and Ken D. McCarthy. 2002. "Hippocampal Astrocytes In Situ Exhibit Calcium Oscillations That Occur Independent of Neuronal Activity." *Journal of Neurophysiology* 87 (1): 528–37. <https://doi.org/10.1152/jn.00268.2001>.