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Blood Test

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We use this protocol and it's working

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Abstract

Blood Test

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1 Transfer the mice into a cage with fresh bedding and fast them for 4h before testing (8:30 am-12:30 pm), while ensuring that the mice have access to drinking water.

2 **Glucose measurement**

2.1 Carefully cut off 1-2 mm of the tail tip using sharp scissors.

2.2 Wipe off the first drop of blood to avoid hemolysis or contamination with tissue fluid before taking new blood samples for blood glucose determination.

2.3 Draw a small blood sample (~3 µl) for the measurement of the basal blood glucose level with the glucometer.

3 **Blood sampling**

3.1 Collect a blood sample (around 30 µl) using a fresh capillary tube (keep the capillary tube horizontal).

3.2 Empty the capillary tube using a pipette by putting the pipette tip at the top of the capillary tube end and carefully pushing the collected blood into a well of the 96-well plate, while avoiding air bubbles.

3.3 Repeat this procedure for all mice - one at a time.

4 After the experiment, centrifuge blood samples at 2,500 x g, 30 min, 4 °C. Transfer the supernatant (plasma) to empty wells of the plate and store it at -20 °C until analysis.

5 Determine plasma insulin levels using a commercially available ELISA kit following the manufacturer's instructions of the kit.