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Blood Test

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Protocol status: Working

We use this protocol and it's working

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Abstract

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- 1 Transfer the mice into a cage with fresh bedding and fast them for 4h before testing (8:30 am-12:30 pm), while ensuring that the mice have access to drinking water.
- 2 **Glucose measurement**
 - 2.1 Carefully cut off 1-2 mm of the tail tip using sharp scissors.
 - 2.2 Wipe off the first drop of blood to avoid hemolysis or contamination with tissue fluid before taking new blood samples for blood glucose determination.
 - 2.3 Draw a small blood sample (~3 μ l) for the measurement of the basal blood glucose level with the glucometer.
- 3 **Blood sampling**
 - 3.1 Collect a blood sample (around 30 μ l) using a fresh capillary tube (keep the capillary tube horizontal).
 - 3.2 Empty the capillary tube using a pipette by putting the pipette tip at the top of the capillary tube end and carefully pushing the collected blood into a well of the 96-well plate, while avoiding air bubbles.
 - 3.3 Repeat this procedure for all mice - one at a time.
- 4 After the experiment, centrifuge blood samples at 2,500 x g, 30 min, 4 °C. Transfer the supernatant (plasma) to empty wells of the plate and store it at -20 °C until analysis.
- 5 Determine plasma insulin levels using a commercially available ELISA kit following the manufacturer's instructions of the kit.