

Aug 14, 2019

Biochemical Measures of Neuropathy - Glutathione Peroxidase

DOI

<https://dx.doi.org/10.17504/protocols.io.3qrgmv6>



Eva Feldman¹

¹University of Michigan - Ann Arbor

Diabetic Complications Consortium
Tech. support email: rmcindoe@augusta.edu



Lili Liang

Create & collaborate more with a free account

Edit and publish protocols, collaborate in communities, share insights through comments, and track progress with run records.

Create free account

OPEN  ACCESS



DOI: <https://dx.doi.org/10.17504/protocols.io.3qrgmv6>

External link: <https://www.diacomp.org/shared/document.aspx?id=54&docType=Protocol>

Protocol Citation: Eva Feldman 2019. Biochemical Measures of Neuropathy - Glutathione Peroxidase. **protocols.io**
<https://dx.doi.org/10.17504/protocols.io.3qrgmv6>

License: This is an open access protocol distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

Protocol status: Working

We use this protocol and it's working

Created: June 04, 2019

Last Modified: August 14, 2019

Protocol Integer ID: 24049

Keywords: Biochemical Measures of Neuropathy, diabetic neuropathy, Glutathione Peroxidase, biochemical measures of neuropathy, glutathione peroxidase summary, neuropathy, increased levels of glucose overload mitochondria, oxidative stress, antioxidant capacity, glucose overload mitochondria, hyperglycemia, glutathione, biochemical measure, axons of the peripheral nervous system, diabetic complication, excess glucose, metabolic change, flow of excess glucose, cellular pathway

Abstract

Summary:

Oxidative stress is highly correlated with the metabolic changes caused by hyperglycemia. Increased levels of glucose overload mitochondria and result in the production of reactive oxygen species (ROS). In addition, the flow of excess glucose through cellular pathways decreases the cell's normal ability to detoxify ROS. As a result, the neurons and axons of the peripheral nervous system contain increased levels of ROS and decreased antioxidant capacity. The following assays are used to measure these changes in rodent models of diabetic neuropathy.

Diabetic Complication:



Neuropathy

Materials

MATERIALS

 Cayman Glutathione Peroxidase Assay Kit **Cayman Chemical Company Catalog #703102**

Reagents & Supplies: HPLC-grade water

Reagent Preparation:

Assay Buffer (10X): Dilute 2 mL of Assay buffer concentrate with 18 mL of HPLC-grade water. Store at 4°C. Stable for 2 months.

Sample Buffer (10X): Dilute 2 mL of Sample buffer concentrate with 18 mL of HPLC-grade water. Store at 4°C. Stable for 1 month.

Glutathione Peroxidase (Control): Dilute 10 μ L of supplied enzyme with 490 μ L of diluted sample buffer. Aliquot 70 μ L into 0.5 mL centrifuge tubes and store at -20°C .

Co-Substrate Mixture: Reconstitute the number of vials required by adding 2 mL of HPLC-grade water to each vial and vortex. Each vial will have enough reagent for 40 wells.

Cumen Hydroperoxide: Ready to use as supplied. Store at -20°C .

Note:

Cayman Chemical ([RRID:SCR_008945](https://www.caymanchem.com/RRID:SCR_008945))

Sample Preparation — Tissue:

1. Homogenize the tissue in 5–10 mL of cold buffer (i.e., 50 mM Tris-HCl, pH 7.5, 5 mM EDTA, and 1 mM DTT) per gram of tissue. **General Equation: $\mu\text{L Buffer} = \text{mg Tissue} \times 10$**
2. Centrifuge at 10,000 x g for 15 minutes at 4°C.
3. Remove supernatant for assay and store on ice. Sample can be stored at –80°C for at least one month.

Performing Assay:

1. Turn on Multiskan and open file gpx.sed.
2. **Background Wells:** add 120 μL of Assay Buffer and 50 μL of co-substrate mixture to three wells.
3. **Positive Control Wells:** add 100 μL of Assay Buffer, 50 μL of co-substrate mixture, and 20 μL of diluted GPx (control) to three wells.
4. **Sample Wells:** add 100 μL of Assay Buffer, 50 μL of co-substrate mixture, and 20 μL of sample to three wells.
5. Initiate reactions by adding 20 μL of cumen hydroperoxide to all wells being used as quickly as possible. Note precise time the reaction is initiated.
6. Place plate onto Multiskan holder and click **START**.
7. Save raw data as an Excel file into the GPx data folder. Use the naming convention gxXXXX.xls, where XXXX is the date in mmdd format.
8. Select Process>Organize. Choose the appropriate data to organize (usually Measure1), then click **OK**. This rearranges the data into columns.
9. Save organized data as an Excel file into the GPx data folder. Use the naming convention gxXXXXor.xls, where XXXX is the date in mmdd format.