



Jul 23, 2025

Binding Assay (His-tagged proteins)

DOI

<https://dx.doi.org/10.17504/protocols.io.261gednqdv47/v1>

Terran Stenger¹, Philippa Kennedy¹

¹University of Minnesota Medical School



Terran D Stenger

University of Minnesota Medical School

Create & collaborate more with a free account

Edit and publish protocols, collaborate in communities, share insights through comments, and track progress with run records.

Create free account

OPEN  ACCESS



DOI: <https://dx.doi.org/10.17504/protocols.io.261gednqdv47/v1>

Protocol Citation: Terran Stenger, Philippa Kennedy 2025. Binding Assay (His-tagged proteins). **protocols.io**
<https://dx.doi.org/10.17504/protocols.io.261gednqdv47/v1>

License: This is an open access protocol distributed under the terms of the **Creative Commons Attribution License**, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

Protocol status: Working

We use this protocol and it's working

Created: August 01, 2023

Last Modified: July 23, 2025

Protocol Integer ID: 85806

Keywords: binding, binding assay, tag present on the trike construct, flow cytometry, tagged protein, functional characterization of these therapeutic molecule, antigen interaction, 10x polyhistidine, insights into trike, therapeutic molecule, trike construct, assessment of trike, trike

Abstract

To assess the binding specificity of tri-specific killer engagers (TriKEs) to their intended target, we utilize a flow cytometry-based binding assay leveraging the 10x polyhistidine (His) tag present on the TriKE construct, as previously described by our lab (see references). This assay enables assessment of TriKE binding efficiency and specificity, providing insights into TriKE-antigen interactions that inform further optimization and functional characterization of these therapeutic molecules.

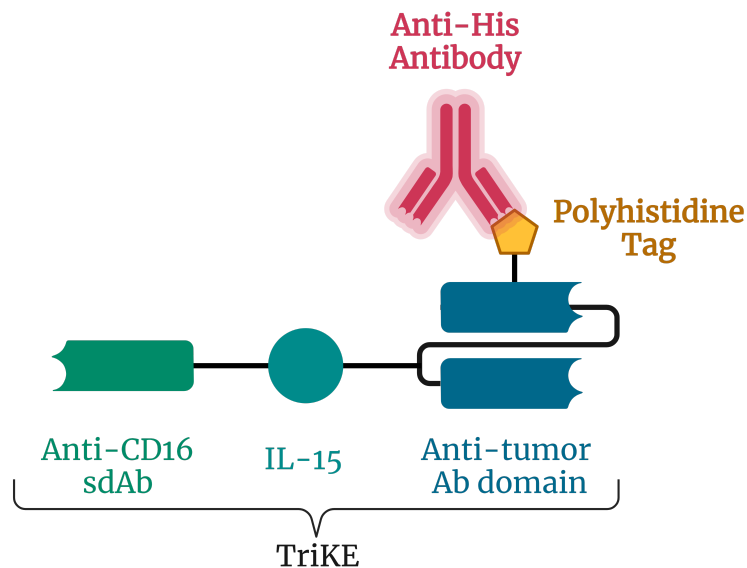


Fig 1. Schematic of the binding assay (figure made in BioRender).

Guidelines

Processed tissue additional controls:

Fc Receptor Blocking Solution, cat. 422302, BioLegend, (1/100 dilution)



Materials

- R10: RPMI (Gibco Cat. No. 2240-089) + 10% fetal bovine serum (Gibco Cat. No. 26140079) + 100 U/mL Penicillin and Streptomycin (Gibco Cat. No. 15140122)
- Flow buffer: 1% human AB serum, 0.5 mM EDTA in PBS
- PE anti-His tag antibody, cat. 362603, Biolegend
- PE mouse IgG2A Isotype control, cat. 400212, Biolegend
- 2% paraformaldehyde/PBS



Cell Preparation

- 1 Cells are resuspended in R10 at 1.5 million cells/mL, and  200 μL (300k cells) are plated in each well in a U bottom 96 well plate.
- 2  300 x g, 00:05:00 The plate is spun in a centrifuge at 300g for  00:05:00 (this is approximately 1200rpm in a large benchtop centrifuge). The supernatant is removed. 5m

Incubation with His-Tagged Protein 30m

- 3  50 μL of 4X histidine(His)-tagged treatments are added to the pellet.
- 3.1 Recommended final concentrations: 0.3 nM, 3 nM, 30 nM for TriKEs and 3 nM, 30 nM and 300 nM for TriKE-PACC.
- 3.2 Prepare 4X drug in R10.
- 4  150 μL of R10 is added. Resuspend.
- 5 Incubate for  00:30:00 at 37°C. 30m
- 6 After the incubation, samples are washed twice with flow buffer to remove excess, unbound protein.

Extracellular + Live/Dead Staining 30m

- 7 Cells are stained in  50 μL of flow buffer (1% AB serum, 0.5 mM EDTA in PBS) containing:
 -  .05 μL LIVE/DEAD Fixable Near-IR Dead Cell Stain, cat. L34976, ThermoFisher
 -  5 μL PE anti-His Tag Antibody, cat. 362603, Biolegend

Note: As controls, untreated cells are stained with the anti-His antibody, and TriKE-treated cells are stained with an isotype control antibody.

- 8 Incubate for 00:20:00 at 4°C. After the incubation, samples are washed twice with flow buffer to remove excess antibody. 20m
- 9 Cells are resuspended in 100 μL of 2% paraformaldehyde/PBS and incubated at room temperature in the dark for 00:10:00 to fix them. Afterwards, wells are topped up to 200 μL with flow buffer and washed once. 10m

Flow Cytometry

- 10 Samples are analyzed via flow cytometry.

10.1

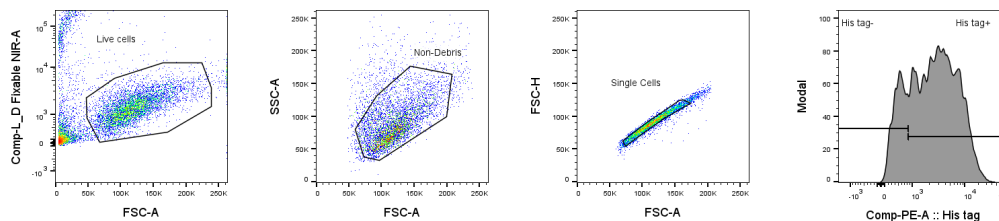


Fig. 2: Example gating strategy in FlowJo

Protocol references

- 1.) Felices M, Lenvik TR, Davis ZB, Miller JS, Vallera DA. Generation of BiKEs and TriKEs to Improve NK Cell-Mediated Targeting of Tumor Cells. *Methods Mol Biol.* 2016;1441:333-46. doi: 10.1007/978-1-4939-3684-7_28. PMID: 27177679; PMCID: PMC5823010.
- 2.) Phung SK, Zorko NA, Soignier Y, Waller RL, Shackelford M, Walker JT, Nelson TD, Selleck C, Bendzick LE, Kotz LE, Kile QM, Bozicevich AJ, Miller SE, Khaw M, Shetty M, Hinderlie P, Ehrhardt M, Li Y, Luo X, Dehm SM, Antonarakis ES, Kennedy PR, Miller JS, Felices M. A PSMA-Targeted Tri-Specific Killer Engager Enhances NK Cell Cytotoxicity against Prostate Cancer. *Cancer Immunol Res.* 2025 Feb 3;13(2):258-272. doi: 10.1158/2326-6066.CIR-24-0273. PMID: 39545924; PMCID: PMC11790377.