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Version 3

Axenic Diatoms cultures protocol V.3

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Protocol status: Working

We use this protocol and it's working

Created: October 24, 2019



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Abstract

Axenic cultures protocol

Axenicity of cultures is obtained by multi-antibiotic treatment: by adding antibiotics directly to the medium in which the strain will be inoculated. After a variable amount of days we refresh the coltures with new medium.

Troubleshooting

antibiotics concentration

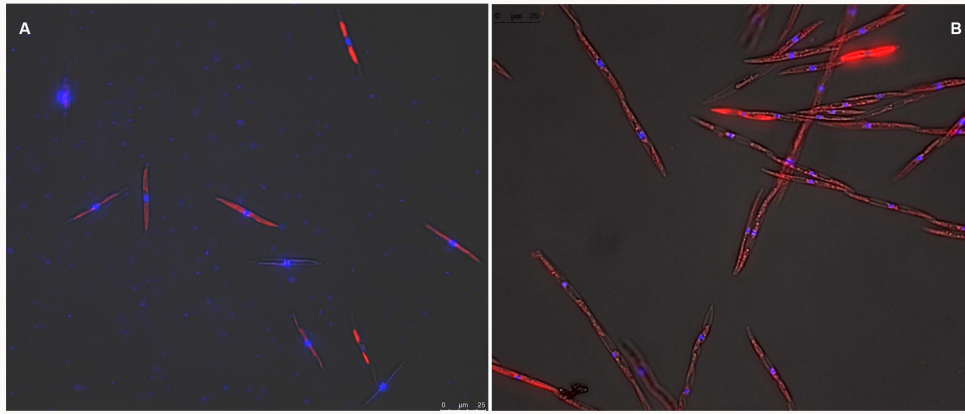
- 1 The antibiotics used are streptomycin, penicillin and ampicillin;
We used the following final concentration:
 - Streptomycin 0,5 mg/ml
 - Penicillin 0,05 mg/ml
 - Ampicillin 0,03 mg/ml
- 2 Add antibiotics in F/2 medium.
Start from 20/50.000 cells in 25 ml of F/2 medium with antibiotics.
Refresh 3 times the cultures every 3 days with new medium with antibiotics.
- 3 Transfer the cultures into a variable higher volume flask containing F/2 medium and antibiotics.
Arrive progressively at the final volume.

contamination test

- 4 Assess axenicity of the strains by fluorescence microscopy using DAPI staining (4',6-diamidino-2-phenylindole-a DNA stain).

(DAPI stains DNA, if bacteria are present in the culture bacterial nucleoids can be visualised as fluorescent spots).

To fix cells use neutralized formaldehyde 1.6%
DAPI final concentration 1:1000



Pseudo-nitzschia multistriata fluorescence microscopy images using DAPI staining.

- A. Normal culture with bacteria
- B. Axenic culture without bacteria.