VERSION 2

AUG 16, 2018

OPEN ACCESS

DOI:

dx.doi.org/10.17504/protocols.io.sk7eczn

Protocol Citation: Tomasz Suchan 2018. AMPure purification protocol.

protocols.io

<https://dx.doi.org/10.17504/protocols.io.sk7eczn>

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Protocol status: Working
correcting some typos

Created: Aug 14, 2018

Last Modified: Aug 16, 2018

PROTOCOL integer ID:
14719

AMPure purification protocol V.2

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GUIDELINES

The protocol below works for 200 µl PCR tubes or plates. If working with 1.5 ml tubes, use larger volumes of EtOH for the washes (step 6 and 7) and Tris/water for the final elution (step 9).

- 1 Remove beads from the fridge, equilibrate to the room temperature, mix thoroughly but do not vortex.

- 2 Add desired ratio of AMPure beads to the DNA to be purified and mix well by pipetting.
- 3 Incubate 5 minutes.
- 4 Place on the magnetic rack.
- 5 Let it stand for 5 minutes on the rack, aspirate and discard supernatant.
- 6 Add 200 µl of freshly prepared 70% EtOH, incubate 30 seconds, aspirate and discard EtOH.
- 7 Repeat the wash: add 200 µl of freshly prepared 70% EtOH, incubate 30 seconds, aspirate and discard EtOH.
- 8 Wait until the pellet dries out completely. DO NOT remove the plate/tubes from the magnetic rack, as the dried AMPure might pop out from the tubes!
- 9 Add desired volume of Tris 10 mM or water, wetting the dried AMPure pellets (add 1 µl to the final volume to avoid pipetting out the beads)

- 10** Remove from the magnetic rack.
- 11** Resuspend by pipetting or vortexing.
- 12** Incubate 10 minutes, incubating in 37°C can improve DNA yield.
- 13** Place on the magnetic rack.
- 14** Let it stand for 5 minutes, pipette out and save supernatant. The eluted DNA is in the supernatant, do not discard it!