Aug 16, 2018 Version 2

AMPure purification protocol V.2

DOI
dx.doi.org/10.17504/protocols.io.sk7eczn

Tomasz Suchan[1]

[1]Institute of Botany, Polish Academy of Sciences

Molecular Biogeography...

Tomasz Suchan
W. Szafer Institute of Botany, Polish Academy of Sciences

DOI: dx.doi.org/10.17504/protocols.io.sk7eczn

Protocol Citation: Tomasz Suchan 2018. AMPure purification protocol. **protocols.io https://dx.doi.org/10.17504/protocols.io.sk7eczn**

Protocol status: Working
correcting some typos

Created: August 14, 2018

Last Modified: August 16, 2018

Protocol Integer ID: 14719

Guidelines

The protocol below works for 200 µl PCR tubes or plates. If working with 1.5 ml tubes, use larger volumes of EtOH for the washes (step 6 and 7) and Tris/water for the final elution (step 9).

1 Remove beads from the fridge, equilibrate to the room temperature, mix thoroughly but do not vortex.

2 Add desired ratio of AMPure beads to the DNA to be purified and mix well by pipetting.

3 Incubate 5 minutes.

4 Place on the magnetic rack.

5 Let it stand for 5 minutes on the rack, aspirate and discard supernatant.

6 Add 200 µl of freshly prepared 70% EtOH, incubate 30 seconds, aspirate and discard EtOH.

7 Repeat the wash: add 200 µl of freshly prepared 70% EtOH, incubate 30 seconds, aspirate and discard EtOH.

8 Wait until the pellet dries out completely. DO NOT remove the plate/tubes from the magnetic rack, as the dried AMPure might pop out from the tubes!

9 Add desired volume of Tris 10 mM or water, wetting the dried AMPure pellets (add 1 µl to the final volume to avoid pipetting out the beads)

10 Remove from the magnetic rack.

11 Resuspend by pipetting or vortexing.

12 Incubate 10 minutes, incubating in 37°C can improve DNA yield.

13 Place on the magnetic rack.

14 Let it stand for 5 minutes, pippete out and save supernatant. The eluted DNA is in the supernatant, do not discard it!