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Protocol status: Working
We use this protocol and it's working

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Adult mouse skin dissociation protocol (on ice) V.2

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ABSTRACT

This protocol was developed to dissociate adult (8-10 wk) mouse skin "on ice". It utilizes two layers of digestion with a *Bacillus licheniformis* protease cocktail, combined with mechanical disruption from a dounce homogenizer. The cell yield is 3000 cells/mg.

ATTACHMENTS

[Skin adult mouse dissociation protocol.pdf](#)

GUIDELINES

***Bacillus Licheniformis* Enzyme Mix (1 mL per 23 mg tissue):**

100 µL b. lich 100 mg/mL (10 mg/mL final) (Sigma, P5380)

1 µL 0.5 M EDTA (Sigma, A8806)

899 µL DPBS (no Ca, Mg) ThermoFisher (cat. #14190)

Preparing enzymes:

The enzyme is made up in DPBS (#14190). It is aliquoted and stored at -80 °C at 100 mg/mL in 100 µL aliquots..

Reagents

Enzymes, trypsin inhibitor, BSA and DNase are made up in DPBS (no Ca, no Mg) from Thermo Fisher (14190).

Bovine Serum Albumin - Sigma (A8806).

Hypothermosol FRS

Required supplies:

2 mL dounce homogenizer – Bellco (1984-10002)

Centrifuge for 1.5 mL, 15 mL conicals

Pipettes and pipet tips

15 ml Conicals (MLS)

1.5 mL tubes (MLS)

30 µM filters - Miltenyi (130-098-458)

Petri dishes (MLS)

Razor blades (MLS)

Ice bucket w/ice

Hemocytometers - InCyto Neubauer Improved (DHC-NO1-5)

Isolating tissue

- 1 After euthanizing mouse, remove hair using Nair: dab with Nair, wait 30 secs, wipe with wet paper towel.
- 2 Isolate tissue and place in ice-cold hypothermosol.
- 3 Scrape off underlying layer of fatty / connective tissue using scalpel before proceeding.

- 4 Mince skin tissue thoroughly on petri dish on ice for 3-4 min on ice into 1-mm³ pieces using razor blade while manipulating tissue with forceps – you will need to use grinding motion and vigorously break up tissue.

00:04:00 mincing

1st digest layer

- 5 Place 23 mg minced tissue into 1 mL B. Lich enzyme cocktail. Incubate on ice.

23 mg minced tissue

- 6 Shake every min; triturate 10x every 2 min with p1000 w/tip cut (start triturating at 2 min) for 20 min.

00:20:00 digest

00:01:00 shake 00:02:00 triturate 10X

- 7 After 20 mins of triturating on ice, use pipet to transfer digest mix to 2 mL dounce homogenizer. Use 10 strokes of Pestle A every 2 min (4 series total, 8 min). Digest mix should become turbid.

00:02:00 dounce homogenize 00:08:00 digest using dounce

- 8 Transfer back to 1.5 mL tube using 1 mL serological pipet. Mix thoroughly and allow to settle on ice 2 min.

00:02:00 settle on ice

- 9 Save 70% (700 μ L) of supernatant, leaving chunks at the bottom of the tube; apply to 30 μ M filter on 15 mL conical. Rinse filter w/5 mL ice-cold PBS/BSA 0.04%. Save flow through on ice and keep filter on tube for 2nd layer.

700 μ L save supernatant 5 mL rinse filter w/ice-cold PBS/BSA 0.04%

2nd digest layer

- 10 Add additional 1 mL b. Lich enzyme mix to residual tissue chunks.
1 mL b. lich enzyme mix
- 11 Triturate 10x every 2 min, shake every min while incubating on ice for 20 additional mins. (50 min. total digest time).
00:20:00 additional digest time
00:02:00 triturate 10x 00:01:00 shake every min.
- 12 Transfer entire volume to same 30 μ M filter on 15 mL conical. Rinse with additional 5 mL ice-cold PBS/BA 0.04%.
5 mL ice-cold PBS/BSA 0.04%

Preparing cells for single cell analysis

- 13 Centrifuge at 300 g for 5 min at 4 °C. Remove supernatant & re-suspend in 100 μ L PBS/BSA 0.04%. Examine using hemocytometer with trypan blue.
00:05:00 centrifuge at 300 g 100 μ L re-suspend in ice-cold PBS/BSA 0.04%
- 14 Adjust concentration to 1,000 cells/ μ L for Chromium or 100 cells/ μ L for DropSeq.