



Immunoprecipitation Protocol V.1

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External link:

http://www.biolegend.com/media_assets/support_protocol/BioLegend_Immunoprecipitation_protocol_012715.pdf

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Guidelines

Application Notes:

Immunoprecipitation is a procedure by which proteins or peptides that react specifically with an antibody are removed from solution and examined for quantity or physical characteristics. Immunoprecipitation can also be used to “enrich” a protein population prior to Western Blotting. For example, one can perform immunoprecipitation with a pan-specific antibody against a protein of interest followed by Western blotting with a modification-specific antibody (such as a phospho-specific antibody or an acetylation-specific antibody).

Tips:

1. The choice of lysis buffer depends on the location of the protein (membrane, cytosolic, nuclear).
2. Immunoprecipitates allowed to incubate overnight may have a higher background than ones processed for shorter periods of time due to time-dependent aggregation or denaturation of cellular proteins.
3. Always use an isotype-matched irrelevant control antibody (monoclonal) or same-species serum from a non-immunized animal to remove non-specific antibody binding in cellular lysates.

Solutions and Reagents:

1X Cell Lysis Buffer:

25 mM Tris-HCl, pH 7.5	1 µg/ml aprotinin
150 mM NaCl	1 mM Na ₃ PO ₄
0.1% Triton X-100	1 mM PMSF
2 mM EDTA	5 mM NaF
1 µg/ml leupeptin	3 mM Na ₄ P ₂ O ₄

5X SDS sample Buffer:

312.5 mM Tris-HCl (pH 6.8) 10% SDS (w/v)

250 mM DTT

50% Glycerol

0.05% Bromophenol Blue (w/v)

Use at 1X

10X SDS Running Buffer:

Dissolve 144 g of Glycine, 30 g of Tris base and 10 g SDS in 800 ml of distilled H₂O.

Add distilled H₂O to 1 liter

Use at 1X

Transfer Buffer:

2.25 g Tris base

10.5 g Glycine 1 g SDS




200 ml Methanol

Add distilled water to 1.0 L




Troubleshooting



Preparation of antibody-protein G agarose beads

- 1 Wash protein G agarose beads with cell lysis buffer by pulsing in a microcentrifuge tube (two minutes at 5,000 rpm).
 00:02:00
- 2 Aspirate and discard supernatant.
- 3 Wash the beads with cell lysis buffer (1/3).
- 4 Wash the beads with cell lysis buffer (2/3).
- 5 Wash the beads with cell lysis buffer (3/3).
- 6 Adjust antibody concentration to 5-10 $\mu\text{g/ml}$ in PBS and transfer 500 μl of diluted antibody to 5-10 μl of agarose beads for each sample.
- 7 Place the antibody-protein G agarose mix on a shaker and rotate at 4°C for one hour.
 01:00:00
- 8 Spin down the protein G beads for two minutes at 5,000 rpm and wash the antibody-beads three times with cell lysis buffer.
 00:02:00

Preparation of cell extract

- 9 Collect cells and centrifuge at 1200 rpm for 5 minutes at 4°C.
 00:05:00
- 10 Discard the supernatant and immediately add 800 μl of ice-cold lysis buffer to the cells and vortex, then incubate for 30 minutes on ice.
 00:30:00
- 11 Freeze and thaw the samples with dry ice for two more cycles or sonicate for 15 seconds to ensure the full release of the proteins from the cells.
 00:00:15
- 12 Spin lysates at 14,000 rpm in a pre-cooled centrifuge for 10 minutes and keep the supernatant.



00:10:00

Immunoprecipitation

- 13 Adjust the protein concentration of the supernatant to 1-2 mg/ml with lysis buffer.
- 14 Mix 100-500 μ l of cell extract with antibody-protein G agarose and rotate the samples at 4°C for about two hours.
 02:00:00
- 15 Collect the agarose beads by pulsing in a microcentrifuge tube (two minutes at 5,000 rpm, 4°C).
 00:02:00
- 16 Aspirate and discard the supernatant.
- 17 Wash the beads with ice-cold cell lysis buffer (1/3).
- 18 Wash the beads with ice-cold cell lysis buffer (2/3).
- 19 Wash the beads with ice-cold cell lysis buffer (3/3).
- 20 After the final wash, remove the supernatant and add 20 μ l of 2X SDS sample buffer.
- 21 Boil for 5 minutes at 95°C.
 00:05:00
- 22 Spin down the beads at maximum speed in a microcentrifuge for 5 minutes at room temperature.
 00:05:00
- 23 Carefully pipette off the supernatant.
- 24 Load 30 μ l of sample in each well of a 1.5 mm thick gel. Run the gel according to manufacturer's recommendations and continue with immunoblotting using [BioLegend's](#)



Western Blotting protocol (alternately, radiolabeled proteins prepared from target cells can be used to directly visualize the immunoprecipitated protein).