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Version 1

## Digestion with NEBNext dsDNA Fragmentase (M0348) V.1

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## Abstract

Protocol for digestion with NEBNext dsDNA Fragmentase (M0348)

## Guidelines

Adequate mixing of NEBNext dsDNA Fragmentase is important for the success of this reaction. NEBNext dsDNA Fragmentase should be vortexed for 3 seconds prior to use.

For tough digestions, add 1  $\mu$ l of 200 mM MgCl<sub>2</sub> to the reaction. Additional MgCl<sub>2</sub> can be added if necessary.

The protocol listed below is for fragmentation of 5 ng–3  $\mu$ g of DNA.

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**End Repair:** Clean up the fragmented DNA (e.g. column purification, or using SPRI) then proceed with desired DNA end repair protocol.

**Agarose Gel Size Selection/Analysis:** Samples can be loaded directly on to an agarose gel. It is not necessary to clean up the reactions prior to loading.

**Polyacrylamide Gel Analysis:** Clean up the fragmented DNA (e.g. column purification) prior to loading the samples on a PAGE gel.

**Long Term Storage:** Clean up the fragmented DNA (e.g. column purifications, or SPRI Beads\*) prior to long term storage.

\*Note: If using SPRI Beads for sample purification, it is recommended to dilute the sample 1:1 with sterile water to allow for faster collection of beads to the magnet.

## Materials

### MATERIALS

 NEBNext dsDNA Fragmentase - 50 rxns **New England Biolabs Catalog #M0348S**

## Troubleshooting



## Before start

Adequate mixing of NEBNext dsDNA Fragmentase is important for the success of this reaction. NEBNext dsDNA Fragmentase should be vortexed for 3 seconds prior to use.

The protocol listed below is for fragmentation of 5 ng–3 µg of DNA.



## 1 Vortex NEBNext dsDNA Fragmentase for 3 seconds

### Note

This protocol is for fragmentation of 5 ng–3 µg of DNA.

### Note

Adequate mixing of NEBNext dsDNA Fragmentase is important for the success of this reaction.

## 2 Quick spin and place on ice

## 3 Mix together the following components in a sterile PCR tube:

### Protocol

NAME  
 **M0348 Digestion Mixture**

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Preview

### 3.1 DNA (5 ng–3 µg), **1–16 µl**

#### Note

If the starting material is 100 ng or less, incubation times should be increased by 10 minutes.

### 3.2 10X Fragmentase Reaction Buffer v2, **2 µl**

 2 µL

**Note**


The Fragmentase Reaction Buffer v2 now contains BSA, eliminating the need to add it separately. The buffer also has increased Mg++ which improves the uniformity of fragmentation across different conditions.

**3.3 Sterile Water to 18 µl****4 Add 2.0 µl dsDNA Fragmentase** 2 µL**5 Vortex the mixture for 3 seconds****6 Incubate at 37°C for the recommended times below to generate the desired fragment size**

	<b>Desired Fragment Size (bp)</b>	<b>Incubation Time (min)</b>
	50–200	25–35
	200–1,000	15–25
	1,000–2,000	10–15

**Note**

If starting material is 100 ng or less, incubation times should be increased by 10 minutes.

**7 Add 5 µl of 0.5 M EDTA to stop the reaction.** 5 µL**Note**

DNA fragments are ready for DNA end repair, size selection or analysis (see guidelines for more information).