

Feb 02, 2025

Version 1

5-ethynyl uridine labeling of nascent mitochondrial genome transcription V.1

DOI

<https://dx.doi.org/10.17504/protocols.io.261gero6wl47/v1>

Tejashree Waingankar¹, Samantha C Lewis¹

¹UC Berkeley



Samantha C Lewis

UC Berkeley

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Protocol Citation: Tejashree Waingankar, Samantha C Lewis 2025. 5-ethynyl uridine labeling of nascent mitochondrial genome transcription. **protocols.io** <https://dx.doi.org/10.17504/protocols.io.261gero6wl47/v1>

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Protocol status: Working

We use this protocol and it's working as of 1/1/2025.

Created: November 04, 2024



Last Modified: February 02, 2025

Protocol Integer ID: 111778

Keywords: Fluorescence microscopy, Transcription, mitochondria, mtDNA, click chemistry, fluorescent readout of localized mitochondrial gene transcription, mitochondrial genome transcription, localized mitochondrial gene transcription, mitochondrial genome transcription this protocol, ethynyl uridine labeling of nascent, immunofluorescence detection of mtdna, ethynyl uridine labeling, mtdna, genome, proteins in cultured cell

Funders Acknowledgements:

Sloan Research Foundation

Grant ID: FG-2023-20384

NSF

Grant ID: 2339182

Abstract

This protocol details a method for 5-ethynyl uridine labeling of nascent mitochondrial genome transcription coupled with immunofluorescence detection of mtDNA-binding proteins in cultured cells. The result is a fluorescent readout of localized mitochondrial gene transcription compatible with high-resolution microscopy.

Guidelines

This protocol was developed to work with cultured mammalian cell lines. It also works with cultured mammalian neurons.

Materials

Buffers and reagents:

- 1 mM Triptolide diluted in DMSO
- 0.5 M 5-ethynyl uridine (EU) in DMSO
- Mitotracker Deep Red (MTDR)

 MitoTracker[®] Deep Red FM - Special Packaging **Thermo Fisher Catalog #M22426**
-  Click-iT[®] Plus EdU Alexa Fluor[®] 647 Imaging Kit **Thermo Fisher Catalog #C10640**
- **Permeabilization solution:** DPBS + 0.1% Triton X-100
- **Blocking buffer:** 0.1% TBST + 1% BSA
- **culture media:** Complete MEM: For 50 ml

	A	B
	Horse serum	5 ml
	Pen/strep antibiotic	500 µl
	L-Glutamine	500 µl
	MEM vitamin supplement	500 µl
	MEM	43.5 ml



Method

1d 14h 35m

- 1 Culture cells in vitro for 24:00:00 in 300 μL complete MEM on a coverslip or in a 35 mm glass bottom culture dish. 1d
- 2 Remove the 300 μL of culture media and add pre-warmed 1 mL of complete culture media.
- 3 Add 1 μL of 1 millimolar (mM) Triptolide to the media and incubate at 37 $^{\circ}\text{C}$ for 01:00:00 . 1h
- 4 Add 1 μL of 0.5 Mass Percent EU to the media and incubate for 03:00:00 at 37 $^{\circ}\text{C}$. 3h
- 5 After 2h, add 50 nanomolar (nM) MTDR to the culture media.
- 6 Remove the media and replace it with fresh, complete culture media. Incubate at 37 $^{\circ}\text{C}$ for 00:10:00 . 10m
- 7 Fix with pre-warmed 4% PFA for 00:20:00 at Room temperature . 20m
- 8 Add 200 μL of 1 Mass Percent Glycine to quench the reaction for 00:05:00 at Room temperature . 5m
- 9 Add 2 mL of permeabilization solution for 00:10:00 at Room temperature . 10m
- 10 Replace the permeabilization solution with the blocking buffer for 00:10:00 at Room temperature . 10m



- 11 Replace the blocking buffer solution with primary antibody solution diluted in blocking buffer; incubate at 4 °C Overnight . 8h
- 12 Prepare a click reaction buffer mixture as described in the manufacturer's protocol.
- 13 Incubate at Room temperature for 00:30:00 while protected from light. 30m
- 14 Remove the solution completely and wash gently with 1 mL of blocking buffer.
- 15 Replace with freshly prepared secondary antibody solution diluted in blocking buffer; incubate for 01:00:00 at Room temperature , protected from light. 1h
- 16 Wash once with the blocking buffer for 00:10:00 . 10m
- 17 Cover cells in 500 µL DPBS and proceed with fluorescence microscopy imaging.

Protocol references

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Acknowledgements

We thank the Lewis Lab for helpful discussion.