Jun 15, 2023

2-step PCR mixture and conditions (Barcoded-head primers for seqs pooling)

Forked from 2-step PCR mixture and conditions (Barcoded-head primers for seqs pooling)

DOI
https://dx.doi.org/10.17504/protocols.io.e6nvwdp32lmk/v1

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OPEN ACCESS

DOI: https://dx.doi.org/10.17504/protocols.io.e6nvwdp32lmk/v1

Protocol Citation: Yin-Tse Huang, Tsu-Chun Hung 2023. 2-step PCR mixture and conditions (Barcoded-head primers for seqs pooling). **protocols.io** https://dx.doi.org/10.17504/protocols.io.e6nvwdp32lmk/v1

Manuscript citation:
Herbold CW, Pelikan C, Kuzyk O, Hausmann B, Angel R, Berry D, Loy A. 2015. A flexible and economical barcoding approach for highly multiplexed amplicon sequencing of diverse target genes. Front. Microbiol. [Internet] 6:731. Available from: http://dx.doi.org/10.3389/fmicb.2015.00731

Protocol status: Working
We use this protocol and it's working

Created: June 15, 2023

Last Modified: September 13, 2023

Protocol Integer ID: 83462

Keywords: pcr mixture, step pcr mixture, powerpol 2x pcr mix, pcr, head primers for seq, head primer, primer, mix, seq

Abstract

PCR mixture and condition (PowerPol 2X PCR Mix)

1 Wear glove, clean up the working bench w. 1% bleach

For 1' PCR head-primers

2 Prepare 1' PCR master mixutre for **head-primers** (**prepare 1.2X of solutions for pipetting error if needed**)

PCR mixture for head-primers for each reaction

	A	B	C	D
	Component	**Volume**	**Volume (1.2X)**	**Final conc.**
	Forward Primer (10 µM)	0.5 µl	1.2 µl	0.2 µM
	Reverse Primer (10 µM)	0.5 µl	1.2 µl	0.2 µM
	PowerPol 2X PCR Master Mix	12.5 µl	15 µl	-
	ddH20	10.25 µl	11.1 µl	-
	Total volume	23.75 µl	28.5 µl	-

Note

Negative control ALWAYS NEEDED! For example, if you have 5 PCR reactions to run, prepare master mixture for 6 reactions (5 DNA template + 1 negative control).

3 Mix the 1' PCR master mixture gently by pippeting. Quick spin the tube.

4 Transfer 23.75 µL 1' PCR master mixutre in 8-strip PCR tubes.

5 Add 1.25 µL DNA template in 8-strip PCR tubes, resulting in a 25 µL reaction mixture for 1' PCR.

Note

Negative control contains only 23.75 µL master mixture but not DNA template

6 Mix the reaction mixture gently by tapping the tubes. Quick spin the tubes.

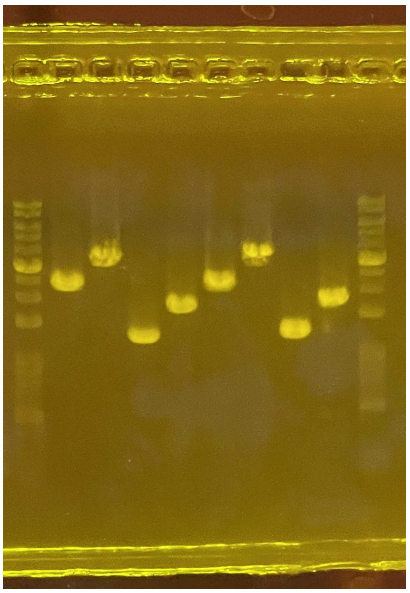
7 Carry out PCR using the following condition:

1' PCR condition for **head-primers**

	A	B	C	D
	Step	**Temp**	**Sec**	**Cycle**
	Initial denaturation	98 ºC	45	
	Denaturation	98 ºC	10	25 cycles
	Annealing	60-66 ºC varied (b)	30	
	Extension	72 ºC	150	
	Final extension	72 ºC	300	
	Preservation	4 ºC	∞	

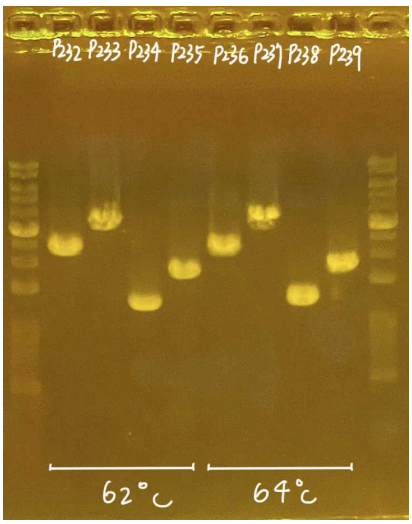
b. Annealing varied, **60-66C** is working; Refer to 1' PCR primers for annealing temperature
c. 1kb ~ 1min extension; enough time allow full extension of sequence

8 Carry out **electrophoresis** for inspection of DNA products



Gel before markdown

9 Markdown wells and upload the pictures to the Lab Google drive



Marked gel picture go to the Lab Google drive

For 2' PCR barcoded-head primers

10 Prepare 2' PCR master mixutre for **barcoded-primers** (**prepare 1.2X of solutions for pipetting error if needed**)

PCR mixture for barcoded-primers for each reaction **(NO PRIMERs at this point!!)**

	A	B	C	D
	Component	**Volume**	**Volume (1.2X)**	**Final conc.**
	ZEJU PCR Master Mix	7.5 µL	9 µL	-
	ddH20	5.55 µL	6.66 µL	-
	Total volume	13.05 µL	15.66 µL	-

Note

Negative control ALWAYS NEEDED! For example, if you have 5 PCR reactions to run, prepare master mixture for 6 reactions (5 DNA template + 1 negative control).

11 Mix the 2' PCR master mixture gently by pippeting. Quick spin the tube.

12 Transfer 13.05 µL of the 2' PCR master mixture to 8-strip PCR tubes.

13 Add 1.2 µL **pre-mixed barcoded-head primers** (Forward + Reverse) to each PCR tubes.

14 Add 0.75 µL of **1' PCR product as template**, resulting in 15 µL reaction mixture for 2' PCR.

Negative control contains only 14.25 µL master mixture and premixed barcoded-head primers but not DNA template

15 Mix gently by tapping the tubes. Quick spin the tubes.

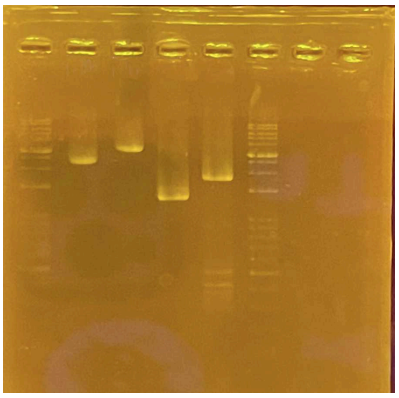
16 Carry out 2' PCR using the following condition:

2' PCR condition for **barcoded-head primers**

	A	B	C	D
	Step	**Temp**	**Sec**	**Cycle**
	Initial denaturation	98 ºC	30	
	Denaturation	98 ºC	15	12 cycles
	Annealing	64-68 ºC varied (a)	15	
	Extension	72 ºC	20 (b)	
	Final extension	72 ºC	210	
	Preservation	Preservation	4 ºC	∞

a. Annealing varied, **65 C** is working based on test on 220531; Refer 2' PCR primers for annealing temperature
b. 1kb ~ 1min extension; enough time allow full extension of sequence

17 Carry out **electrophoresis** for inspection of DNA products



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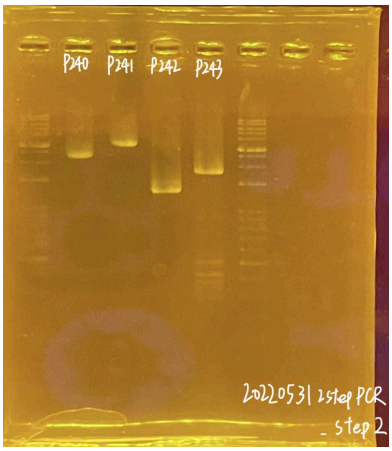

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