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18S Metagenomics in a Field Setting

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Stefan Prost^{1,2}, Gideon Erkenwick^{3,4}, Mrinalini Watsa^{3,5,4}, Aaron Pomerantz^{6,7}

¹LOEWE-Center for Translational Biodiversity Genomics, Senckenberg Museum, 60325 Frankfurt, Germany;

²South African National Biodiversity Institute, National Zoological Garden, Pretoria 0184, South Africa;

³University of Missouri - Saint Louis; ⁴Field Projects International; ⁵Washington University in Saint Louis;

⁶University of California, Berkeley, CA, USA, Department of Integrative Biology;

⁷Marine Biological Laboratory, Woods Hole, MA - USA



Mrinalini Watsa

San Diego Zoo Wildlife Alliance

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Protocol status: Working

We use this protocol and it's working

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Abstract

This protocol is was used to conduct DNA 18S metagenomics on FPI's Genomics in the Jungle - 2018 field course at the Green Lab, located and Inkaterra Guides Field Station, Madre de Dios, Peru.

Guidelines

This protocol starts from already extracted DNA

Materials

STEP MATERIALS

 Agencourt Ampure XP **Beckman Coulter Catalog #A63880**

Protocol materials

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Amplification


1 **Remove samples and the following reagents and let thaw, once thawed keep on ice block**

- 10mM DNTPs
- 25 mMgCl
- 5x Go Taq Buffer
- Forward primer 5'-CAGCAGCCGCGGTAATTCC-3' (10uM)
- Reverse primer 5'-CCCGTGTTGAGTCAAATTAAGC-3' (10uM)
- GoTaq Hotstart Polymerase 5u/ul

Make PCR cocktail for # of samples * 1.1 (10% extra). Don't forget to include 1 PCR negative control for each separate PCR

Run PCR according to the following cycle conditions:

- Initial denaturation  95 °C for 120s
- 25 cycles of  98 °C for  00:00:30 ,  51 °C for  00:00:30 ,  72 °C for  00:00:45
- Final extension  72 °C for  00:05:00

Upon completion remove, label, and store at  4 °C , or take directly to electrophoresis

2 **Equipment**


- BlueGel system
- MiniOne system



Create .8 - 1.0% agarose 1 gel with 13 combs

- Measure 1 g of agarose
- Mix agarose with 100 mL of 1xTBE
- Microwave the mixture until agarose is completely dissolved (1-3 min)
- Pour the agarose gel into the tray with the comb in place.
- Allow the agarose gel to harden (20-30 min)



Insert the agarose gel into electrophoresis equipment and add 1xTBE buffer until the agarose gel is submerged

Spot check with  2 μL **of each sample**



Mix  1 μL **of loading dye to**  2 μL **of each sample and load the gel.** (If Green Taq buffer with built in loading dye was used, skip this step).

Load  5 μL **of 100bp ladder into the agarose gel.**











Turn on the electrode and let the DNA run until the band is identifiable

Barcoding PCR

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- A barcoding PCR was run to attach barcodes from the 96-barcode kit for the MinION to each sample
- We did not use special PCR mastermix at this stage, using instead a mix similar to that of the PCRs above
- We used  1 μL of each barcode primer and  2 μL of every positive PCR amplicon in a total volume of 25 μL

We ran the PCR at the following conditions:

- Initial denaturation of hotstart taq at  95 $^{\circ}\text{C}$ for  00:02:00
- Denaturation at  95 $^{\circ}\text{C}$ for  00:00:30
- Annealing at  62 $^{\circ}\text{C}$ for  00:00:30
- Extension at  72 $^{\circ}\text{C}$ for  00:00:45
- Total number of cycles - 18
- Final Extension at  72 $^{\circ}\text{C}$ for  00:05:00

Quantification

4

Equipment

Equipment

Quantus Fluorometer

NAME

DNA and RNA Quantifier

TYPE

Promega

BRAND

E6150

SKU

<https://www.promega.com/products/fluorometers-luminometers-multimode-readers/fluorometers/quantus-fluorometer/?catNum=E6150>

LINK



- dsDNA dye
- Qubit Assay Tubes
- Your DNA sample


Pulse vortex your sample and spin down.

Add  200 μL **of dsDNA dye into the qubit assay tube.**

- Make sure to cover up the tube from light

Transfer  1 μL **of DNA sample into qubit assay tube**

Pulse vortex and spin down

Let it sit for  00:05:00 **in room temperature**

- Covered from the light

Gently vortex and spindown the sample

Calibrate the Quantus Fluormeter by standard and reference testing.

- Follow the same process but use  1 μL ddH₂O and Lambda DNA.

Insert the qubit assay tube into the Quantus Flurometer



Normalisation and Pooling

5 **Dilute each sample to 50 nM**

- Mix 5 uL of the sample with calculated amount of ddH₂O (based on DNA concentration above) to make each sample the same concentration of 50 nM
- Then pool 5 uL of each dilution into a single tube.
- This is now your library.

SPRI Cleanup

- 6 Run a SPRI cleanup of the library using your choice of bead purification systems/kits in a 1:1 ratio. Resuspend in the same volume.



Agencourt Ampure XP **Beckman Coulter Catalog #A63880**